

### IntechOpen

## Neuroprotection

Edited by Raymond Chuen-Chung Chang and Yuen-Shan Ho





## Neuroprotection

### Edited by Raymond Chuen-Chung Chang and Yuen-Shan Ho

Published in London, United Kingdom













## IntechOpen





















## Supporting open minds since 2005



Neuroprotection http://dx.doi.org/10.5772/intechopen.77296 Edited by Raymond Chuen-Chung Chang and Yuen-Shan Ho

#### Contributors

Patrick Flood, Monika Sharma, Maria Bogdan, Liliana Mititelu-Tartau, Victor Gheorman, Gratiela Eliza Popa, Gabriela Rusu, Beatrice Buca, Liliana Pavel, Ina Cristofor, Liliana Foia, Ancuta Goriuc, Cosmin-Gabriel Tartau, Amy Lo, Shen Nian, Arry Yanuar, Widya Dwi Aryati, Rezi Riadhi Syahdi, Nabilah Nurtika, Raymond Chuen-Chung Chang, Yuen-Shan Ho

#### © The Editor(s) and the Author(s) 2019

The rights of the editor(s) and the author(s) have been asserted in accordance with the Copyright, Designs and Patents Act 1988. All rights to the book as a whole are reserved by INTECHOPEN LIMITED. The book as a whole (compilation) cannot be reproduced, distributed or used for commercial or non-commercial purposes without INTECHOPEN LIMITED's written permission. Enquiries concerning the use of the book should be directed to INTECHOPEN LIMITED rights and permissions department (permissions@intechopen.com).

Violations are liable to prosecution under the governing Copyright Law.

#### CC BY

Individual chapters of this publication are distributed under the terms of the Creative Commons Attribution 3.0 Unported License which permits commercial use, distribution and reproduction of the individual chapters, provided the original author(s) and source publication are appropriately acknowledged. If so indicated, certain images may not be included under the Creative Commons license. In such cases users will need to obtain permission from the license holder to reproduce the material. More details and guidelines concerning content reuse and adaptation can be found at http://www.intechopen.com/copyright-policy.html.

#### Notice

Statements and opinions expressed in the chapters are these of the individual contributors and not necessarily those of the editors or publisher. No responsibility is accepted for the accuracy of information contained in the published chapters. The publisher assumes no responsibility for any damage or injury to persons or property arising out of the use of any materials, instructions, methods or ideas contained in the book.

First published in London, United Kingdom, 2019 by IntechOpen IntechOpen is the global imprint of INTECHOPEN LIMITED, registered in England and Wales, registration number: 11086078, The Shard, 25th floor, 32 London Bridge Street London, SE19SG - United Kingdom Printed in Croatia

British Library Cataloguing-in-Publication Data A catalogue record for this book is available from the British Library

Additional hard and PDF copies can be obtained from orders@intechopen.com

Neuroprotection Edited by Raymond Chuen-Chung Chang and Yuen-Shan Ho p. cm. Print ISBN 978-1-78984-736-9 Online ISBN 978-1-78984-737-6 eBook (PDF) ISBN 978-1-78985-925-6

# We are IntechOpen, the world's leading publisher of **Open Access books** Built by scientists, for scientists

Open access books available

4,200+ 116,000+ 125M+

International authors and editors

Downloads

15 Countries delivered to

Our authors are among the lop 1%

most cited scientists

12.2%





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science<sup>™</sup> Core Collection (BKCI)

### Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



## Meet the editor



Dr. Chang is the Lab Chief for the Laboratory of Neurodegenerative Diseases in the School of Biomedical Sciences, LKS Faculty of Medicine, the University of Hong Kong. Dr. Chang has organized the International Alzheimer's Disease Conference every year since 2000. He has published over 142 peer-reviewed papers, 14 book chapters, and edited 3 books on neurodegenerative diseases. Dr. Chang is the chief editor for the *American Journal* 

of Alzheimer's Disease and Other Dementias, senior editor for the Journal of Neuroimmune Pharmacology, and associate handling editor for Frontiers in Neurology/ Neurosciences/Psychiatry. He is on the Scientific Advisory Board of the International AD/PD Symposium, a member of the editorial boards of more than 20 different journals, and a grant reviewer for different grant agencies/foundations.



Dr. Ho has been working for some time on aging-associated neurodegenerative diseases. She is a registered Chinese medicine practitioner with rich experience in both laboratory and clinical research. She has worked as an assistant professor in Macau and is now at the School of Nursing in Hong Kong Polytechnic University. Her research interest includes the use of Chinese herbal medicine and acupuncture to prevent neurodegeneration (neu-

roprotection). Dr. Ho focuses her research on topics including, but not limited to, disease progression and risk factors leading to neurodegenerative diseases. She has expertise in combining Western and Chinese medicine to elicit holistic effects on the body to exert neuroprotection. She is a member of the editorial boards of four journals and a constant grant reviewer for different journals and funding agencies.

### Contents

Preface	XIII
Section 1	
Neurotransmitter Receptor as Target for Neuroprotection	1
<b>Chapter 1</b> Introductory Chapter: Concept of Neuroprotection - A New Perspective <i>by Raymond Chuen-Chung Chang and Yuen-Shan Ho</i>	3
<b>Chapter 2</b> Current Therapeutic Approaches from Imidazoline and Opioid Receptors Modulators in Neuroprotection <i>by Liliana Mititelu-Tartau, Maria Bogdan, Victor Gheorman, Liliana Foia,</i> <i>Ancuta Goriuc, Gabriela Rusu, Beatrice Buca, Liliana Pavel, Ana Cristofor,</i> <i>Cosmin-Gabriel Tartau and Gratiela Eliza Popa</i>	13
<b>Chapter 3</b> The Role and Development of the Antagonist of Adenosine A <sub>2A</sub> in Parkinson's Disease by Widya Dwi Aryati, Nabilah Nurtika Salamah, Rezi Riadhi Syahdi and Arry Yanuar	31
<b>Chapter 4</b> Adrenergic Receptors as Pharmacological Targets for Neuroinflammation and Neurodegeneration in Parkinson's Disease <i>by Monika Sharma and Patrick M. Flood</i>	45
Section 2 Neuroprotection to the CNS	69
Chapter 5 Protecting the Aging Retina by Shen Nian and Amy C.Y. Lo	71

## Preface

Neuroprotection is a key approach to maintaining brain health to prevent the retraction of synapses, accumulation of neurodegenerative proteins (tau,  $\beta$ -amyloid peptide, α-synuclein, huntingtin, or any misfolded proteins), activation of neuroinflammation, or even the breakdown of the blood-brain barrier. Neuroprotection can be applied to chronic neurodegenerative diseases (Alzheimer's disease, Parkinson's disease, Lewy's body diseases, Huntington's disease, amyotrophic lateral sclerosis, or even brain tumors), acute neurological disorders (brain trauma, stroke, transient ischemia, epilepsy, or cerebral hemorrhage), or even mental disorders (psychosis, depression, or schizophrenia). The traditional view of neuroprotection is often restricted to the use of agonists or antagonists of neurotransmitter receptors. While this is still a great approach to safeguard neurons, neuroprotection strategy is not limited to modulating neurotransmitter receptors. Neuroprotection can be accomplished by (1) natural products, (2) herbal medicine, (3) different forms of exercise, (4) learning new skills or languages, or (5) even good sleeping patterns. From all these new perspectives, the concept of neuroprotection is limited not only to the protection of neurons in the pathogenesis of diseases but also to the prevention of any detrimental factors leading to neuronal cell loss.

The major aim of this book is to focus on different approaches to achieve neuroprotection. In this book, most of our authors review the advantages of neurotransmitter receptors. Mititelu-Tartau and Bogdan's group reviews the imidazoline ligands and opioid ligands in Alzheimer's disease. They review the neuroprotective effects of agmatine on memory. In addition, they also summarize the findings of using different opioid receptor ligands to elicit neuroprotection. Similarly, Yanuar's group reviews the beneficial effects of the adenosine  $A_{2A}$  receptor antagonist. It has been reported that the adenosine receptor antagonist elicits neuroprotective effects, partially because of its signaling pathways to activate the cytosolic fraction of cAMP. Yanuar's group reviews how  $A_{2A}$  antagonists provide neuroprotection.

Apart from modulating neurotransmitter receptors, Flood's group reviews another very important pathological factor leading to neurodegeneration: neuroinflammation. Microglial cells are the major line of cells to be activated to produce cytokines and free radicals to damage neurons. Interestingly, Flood's group reviews how different adrenergic receptor agonists and antagonists modulate microglial responses. This independent review suggests that modulation of neuroinflammation can also be achieved by intervening neurotransmitter receptors and their associated signaling events.

Neuroprotection can be disease specific but use different approaches. Nian and Lo review the neuroprotection of aging eyes, in which aging-associated macular degeneration often occurs in the elderly. With the progression of the disease and identification of biological targets at different states of the disease, the strategy of neuroprotection can be changed.

Taken together, neuroprotection receives increasing attention from different approaches in different states of neurodegenerative diseases, acute neurological

disorders, and even mental disorders. Even in disease progression, neuroprotection can prevent the spread of neuronal damage or neuronal loss from one region to other regions. This book will give insights to scientists in the field to stimulate their research, medical professionals to review their clinical practices, and others who would like to learn more about different neuroprotective approaches.

#### Raymond Chuen-Chung Chang, PhD

Laboratory of Neurodegenerative Diseases, School of Biomedical Sciences, LKS Faculty of Medicine, The University of Hong Kong, Pokfulam, Hong Kong

#### Yuen-Shan Ho, PhD

School of Nursing, Faculty of Health and Social Sciences, The Hong Kong Polytechnic University, Kowloon, Hong Kong Section 1

Neurotransmitter Receptor as Target for Neuroprotection

#### Chapter 1

## Introductory Chapter: Concept of Neuroprotection - A New Perspective

Raymond Chuen-Chung Chang and Yuen-Shan Ho

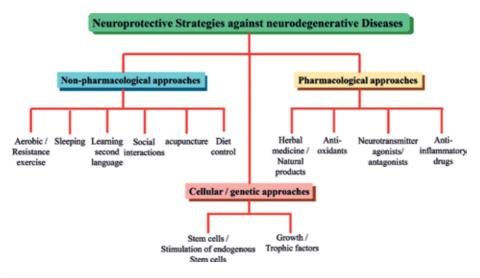
#### 1. Introduction

Neuroprotection is an approach to preserve neurons so that neurons cannot be hurt by different pathological factors in neurodegenerative diseases. It can be an approach before the onset of the disease so that neurons cannot be affected by any risk factors. It can also be an approach during the progression of the disease to prevent spreading of injury from one neuron to neighboring neurons. Therefore, neuroprotection can also be one approach as "disease-modifying agent" to delay and even stop progressive neurodegeneration.

The concept of neuroprotection has long been confined to intervene neurotransmitter receptors via agonists and antagonists. A very well-known example is the neuroprotective effect of caffeine via adenosine receptor, because caffeine is an A2 receptor antagonist. Caffeine can exert neuroprotection via adenosine A2 receptor to protect dopaminergic neurons in an experimental model of Parkinson's disease (PD) by using 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) as a toxin agent [1]. Therefore, drinking coffee has been considered to be neuroprotection. Apart from adenosine receptor on neurons, it is now known that microglial cells also express A2 receptor, and A2 receptor antagonist or caffeine can reduce activation of microglial cells [2]. Since caffeine or A2 receptor antagonists can protect neurons and minimize activation of microglial cells, research is still very active in this direction. Indeed, investigation of other neurotransmitter receptor antagonists, agonists, blockers, or even partial blockers is still a very active research area in neuroprotection.

While it is exciting to reveal differential neuroprotective effects via modulating receptors for different neurotransmitters, we should not restrict ourselves to this perspective only. In fact, a wider scope of neuroprotection has been evolved. The concept of neuroprotection can now be categorized into three groups: (A) pharmacological intervention, (B) non-pharmacological intervention, and (C) cellular and genetic approaches (**Figure 1**).

In group A neuroprotective approaches, pharmacological intervention includes modulation of neurotransmitter receptors (as introduced above), anti-oxidative stress, and anti-inflammatory responses [3]. These are classical pharmacological approaches. These methods have been explored for decades and can be used for both neuroprotection and as disease-modifying agents. Their effects can be target-specific for one single protein (e.g., receptor for neurotransmitter or one type of cytokines) or pathway-specific (anti-oxidative responses using nuclear factor erythroid 2-related factor 2, Nrf2). Since single biological target may limit the beneficial effects and may not be able to intervene the complexity of the disease



#### Figure 1.

A summary of different neuroprotective approaches.

progression, neuroprotective effects from herbal medicine or natural products receive increasing attention in scientific research because of their multiple effects on different biological targets [4–6]. For example, it has been shown that triterpenoids extracted from *Prunella vulgaris* has anti-inflammatory effects [7]. We have proved that polysaccharide part of *Prunella vulgaris* can modulate the immune responses of microglia and macrophages [8, 9]. Therefore, *Prunella vulgaris* exerts neuroprotection toward neurodegenerative diseases against neuroinflammation.

In group B neuroprotective approaches, non-pharmacological approaches receive increasing attention. Since it is quite difficult to ask a healthy adult or even elderly to constantly take medicine even though it is herbal, non-pharmacological approaches to earn neuroprotection are most welcome. Among these different methods, exercise is the best to prevent nearly all kinds of diseases. Exercise can be further divided into aerobic and resistance exercises, in which aerobic exercise is suitable for motor-healthy adults or elderly. However, there is a large population of elderly who have difficulty in their motor functions. Resistance exercise becomes an excellent tool for them. Both aerobic and resistance exercises can affect the metabolisms of the body and induce trophic factors like myokines, released from muscles and liver [10, 11]. Since body metabolism is important, diet control is also important to reduce any risk factors leading to the development of neurodegenerative diseases, such as diabetes, hyperlipidemia, or hypercholesterolemia. All of these risk factors are related to vascular components, which may further add on vascular dementia [12–14]. Apart from exercise, it is well-known that, maintaining social activity is essential to keep our brain healthy [15]. In addition, sleeping can promote clearing of pathological factors, such as  $\beta$ -amyloid (A $\beta$ ) peptide and phosphorylated tau protein [16, 17]. Also, it has been recently shown that learning second language or bilingualism can be neuroprotective [18]. Furthermore, increasing lines of evidence have shown that acupuncture can help adjusting the body metabolism and immunity [18, 19]. Therefore, acupuncture can also be considered to be one method in non-pharmacological approaches. One key point should be noticed that all these neuroprotective approaches are multiple targets. This may be why they are so effective in minimizing neurodegeneration to preserve neurons.

Neuroprotective approaches in group C are the new extension of multiple effects from trophic factors secreted by genetically engineered cells or viral vector. Alternatively, stimulating the proliferation and differentiation of endogenous stem cells, application of induced pluripotent stem cells (iPSC), or mesenchymal stem cells are popular trends in scientific research to prevent neurodegeneration and neuronal loss [20–22].

Since there are several chapters about the effects of neurotransmitter receptors, we will focus on some neuroprotective approaches only, as this is an introductory chapter.

#### 2. Neuroprotective effects of herbal medicine and natural products

This direction of neuroprotection is the most controversial approach among different pharmacological tools. This is because most of the herbal medicine or natural products have a wide array of effects. If we use traditional way of thinking to target a specific protein or signaling pathway, herbal medicine fails to do so. However, if we are aware of neurodegenerative diseases that are usually multifactorial, we can then accept that herbal medicine should be the direction to prevent neuronal loss and spreading of neurodegeneration. Our group has long been working on discovering herbal medicine as neuroprotective agents and investigating their underlying mechanisms. Since most of the herbal medicines are in decoction form, for oral consumption, the effective components in different herbs are not limited to one single chemical. The small molecules being extracted from one herb can also be found from other herbs. There are many laboratories investigating those small molecules and have Research and Development (R&D) to be commercial products. In contrast, big molecules such as polysaccharides have been an unexplored area. In fact, a large portion of components from herbs is polysaccharides. When we use hot water to prepare decoction, what we can easily absorb is polysaccharides. Therefore, we have investigated polysaccharides extracted from different herbs [23].

We have first found that polysaccharides from *Nerium indicum* exert neuroprotective effects against  $\beta$ -amyloid (A $\beta$ ) neurotoxicity cultured neurons [24, 25]. Then, we also found that neuroprotective effects of Verbena officinalis and Ganoderma lucidum [26, 27]. Having investigated different herbal medicine, we found that the polysaccharides from Lycium barbarum (Wolfberry) are potent [28, 29]. Polysaccharides can be extracted in hot water or from alkaline condition and elicit neuroprotection [30]. It should be noted that not all polysaccharides, but only some sub-fractions provide neuroprotective effects. Apart from neurons in the brain, polysaccharides from wolfberry can also protect the retina against experimental glaucoma and stroke [31–33]. As harvesting and planting processes have good agricultural practice and the extraction can be done under stringent control of good manufacturing practice, we choose wolfberry as our long-term study. Wolfberry is indeed an anti-aging Chinese herbal medicine. We would like to understand the concept of "anti-aging." Therefore, we have investigated its application in different aging-associated neurodegenerative diseases [34]. In fact, not all aging-associated neurodegenerative diseases can be attenuated by the polysaccharide fraction of Wolfberry. In Parkinson's disease (PD), more potent antioxidants are required to provide neuroprotection.

In experimental PD using 6-hydroxydopamine (6-OHDA) as toxin agent, we employed oxyresveratrol, which is a natural product but a structural analog of resveratrol, and found that it can attenuate neurodegeneration [35, 36]. For this kind of neurodegenerative disease requiring high levels and high capacity of antioxidant, another method is to employ pro-drug approach so that oxidant cleave the precursor and the product becomes potent antioxidant. Interestingly, polyphenol (-)-epigallocatechin-3-gallate (EGCG) from green tea falls in this category [37].

The advantages of using herbal medicine or natural products are to make use of their multiple effects. In neurodegenerative diseases, a wide array of stress responses is stimulated because of free radicals or accumulation of misfolded or badly folded proteins. Most of the herbal medicine and natural products attenuate many stress kinases. They may not be a good candidate to clear accumulation of bad proteins; however, they can inhibit the cascades of stress responses (e.g., activation of c-Jun-N-terminal kinase, JNK; endoplasmic reticulum stress pathways), which usually lead to neuronal apoptosis. Some of the herbal medicine or natural products can even strengthen the survival signaling pathways such as mTOR or Akt pathways.

In addition to their direct effects on neurons, there are many herbal medicine or natural products that can modulate body immune responses. For example, polysaccharides fraction from *Prunella vulgaris* L. can modulate innate immune responses of macrophages [8, 9]. It has been increasingly aware that activation of body (systemic) immune responses are the origin of our sickness responses leading to low appetites and even fever [38]. Long-term effects of systemic immune responses can even result in psychological depression or acute delirium [39]. Experimentally, we have shown that infection in the body or immune responses triggered by surgery (laparotomy) can modulate cognitive functions because of stimulation of neuroimmune responses [40, 41]. Therefore, modulation of body immunity can be a powerful method to minimize neuroimmune responses and then reduce cognitive dysfunctions.

#### 3. Aerobic and resistance exercises

As emphasized above, the advantages of herbal medicine and natural products are their multiples and wide array effects. Similarly, exercise is another method to achieve this goal. Exercise can stimulate production of different trophic/growth factors, e.g., vascular endothelial growth factor (VEGF), brain-derived neurotrophic factor (BDNF), insulin-like growth factor-1 (IGF-1), peroxisome proliferatoractivated receptor gamma coactivator-1 $\alpha$  (PGC-1 $\alpha$ )/neuronal fibronectin type III domain-containing protein 5 (FNDC5). Some trophic factors can be secreted by muscles or liver, which is collectively called myokines. It has been shown that muscle cells can produce PGC-1 $\alpha$ , irisin, cathepsin B, myostatin (growth differentiation factor 8), fibroblast growth factor 21 (FGF-21), IL-6, IL-15, myonectin, and SPARC (osteonectin) as novel brain-beneficial myokines [10, 42]. Some of them, such as FGF21, can also be secreted by the liver or even in the brain upon exercise [43]. They all can affect muscle, liver, adipose tissues, and brain cells, leading to preservation of cognitive functions, modulating the lipid metabolism, and immune responses [44]. This kind of myokine can pass through the blood-brain barrier.

For motor-healthy subject, aerobic exercise is beneficial to majority of people. As discussed above, myokines can be induced so that they provide neuroprotective effects to the brain and attenuate any systemic immune responses. While aerobic exercise is known to be good, a great number of elderly and demented patients could not enjoy the benefit of aerobic exercise. Those patients may have prior knee replacement or fall down hurting the legs. They may be lying down on the bed. No matter how we advocate the beneficial effects of aerobic exercise, this will not be beneficial to this group of patients. Does it mean that they will not have any beneficial effects of exercise? This question leads us to investigate the beneficial effects of resistance exercise.

In a systematic review, aerobic exercise shows beneficial effects on cognitive functions and executive functions, which is better than that from resistance exercise [45]. Furthermore, aerobic exercise seems to elicit strong anti-inflammatory effect than that of resistance exercise [46]. However, resistance exercise seems to particularly increase the volume of hippocampus [47]. A study in laboratory animal

Introductory Chapter: Concept of Neuroprotection - A New Perspective DOI: http://dx.doi.org/10.5772/intechopen.85631

has shown that aerobic exercise can markedly increase BDNF; whereas, resistance exercise can significantly increase IGF-1 [48]. Although resistance exercise may have some limitations, this is still a good choice for those elderly and dementia patients to have this form of exercise. IGF-1 can still elicit multiple functions in the brain to preserve neurons.

#### 4. Concluding remarks

Since this is an introductory chapter, we do not intend to discuss every single method of neuroprotection in detail. We should keep in mind that neurodegenerative diseases are multiple-hit processes. Therefore, no single biological target can afford all neuroprotective needs. Approaches in using herbal medicine and natural products remind us that multiple biological targets may be the way to exert effective neuroprotection. Therefore, any non-pharmacological approaches including exercise and even the new stem cell approaches exert multiple effects. This can be the way for our effective neuroprotective strategies.

#### **Author details**

Raymond Chuen-Chung Chang<sup>1,2\*</sup> and Yuen-Shan Ho<sup>3</sup>

1 Laboratory of Neurodegenerative Diseases, LKS Faculty of Medicine, The University of Hong Kong, Pokfulam, Hong Kong SAR

2 State Key Laboratory of Brain and Cognitive Sciences, The University of Hong Kong, Pokfulam, Hong Kong SAR

3 Faculty of Health and Social Sciences, School of Nursing, The Hong Kong Polytechnic University, Kowloon, Hong Kong SAR

\*Address all correspondence to: rccchang@hku.hk

#### IntechOpen

© 2019 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/ by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

#### References

[1] Chen JF, Xu K, Petzer JP, et al. Neuroprotection by caffeine and A(2A) adenosine receptor inactivation in a model of Parkinson's disease. The Journal of Neuroscience. 2001;**21**:RC143

[2] Caetano L, Pinheiro H, Patrício P, et al. Adenosine A2A receptor regulation of microglia morphological remodeling-gender bias in physiology and in a model of chronic anxiety. Molecular Psychiatry. 2017;**22**:1035-1043

[3] Chang RCC, Chiu K, Ho YS, So KF. Modulation of neuroimmune responses on glia in the central nervous system: Implication in therapeutic intervention against neuroinflammation. Cellular & Molecular Immunology. 2009;**6**:317-326

[4] Ho YS, So KF, Chang RCC. Antiaging herbal medicine—How and why can they be used in aging-associated neurodegenerative diseases? Ageing Research Reviews. 2010;**9**:354-362

[5] Ho YS, So KF, Chang RCC. Drug discovery from Chinese medicine against neurodegeneration in Alzheimer's and vascular dementia. Chinese Medicine. 2011;**6**:15. DOI: 10.1186/1749-8546-6-15

[6] Chao J, Leung Y, Wang M, Chang RCC. Nutraceuticals and their preventive or potential therapeutic value for Parkinson's disease. Nutrition Reviews. 2012;**70**:373-386

[7] Wang YY, Yang YX, Zhe H, He
ZX, Zhou SF. Bardoxolone methyl
(CDDO-me) as a therapeutic agent:
An update on its pharmacokinetic and pharmacodynamic properties. Drug
Design, Development and Therapy.
2014;8:2075-2088

[8] Fang X, Yu MS, Yuen WH, Zee SY, Chang RCC. Immune modulatory effects of *Prunella Valgaris L.* on monocytes/macrophages. International Journal of Molecular Medicine. 2005;**16**:1109-1116

[9] Fang X, RCC C, Yuen WH, Zee SY. Immune modulatory effects of *Prunella Valgaris L*. International Journal of Molecular Medicine. 2005;**15**:491-496

[10] Liu Y, Yan T, Chu JMT, Chen Y, Dunnett S, Ho YS, et al. The beneficial effects of physical exercise in the brain and related pathophysiological mechanisms in neurodegenerative diseases. Laboratory Investigation. 2019. DOI: 10.1038/s41374-019-0232-y. In press

[11] Marston KJ, Brown BM, Rainey-Smith SR, Peiffer JJ. Resistance exerciseinduced responses in physiological factors linked with cognitive health. Journal of Alzheimer's Disease. 2019. DOI: 10.3233/JAD-181079. In press

[12] Li G, Mayer CL, Morelli D, et al. Effect of simvastatin on CSF Alzheimer disease biomarkers in cognitively normal adults. Neurology. 2017;89:1251-1255

[13] Appleton JP, Scutt P, Sprigg N, Bath PM. Hypercholesterolaemia and vascular dementia. Clinical Science (London, England).
2017;131:1561-1578

[14] Besser LM, Alosco ML, Ramirez Gomez L, et al. Late-life vascular risk factors and Alzheimer disease neuropathology in individuals with normal cognition. Journal of Neuropathology and Experimental Neurology. 2016;75:955-962

[15] Rege SD, Geetha T, Broderick TL, Babu JR. Can diet and physical activity limit Alzheimer's disease risk? Current Alzheimer Research. 2017;**14**:76-93 Introductory Chapter: Concept of Neuroprotection - A New Perspective DOI: http://dx.doi.org/10.5772/intechopen.85631

[16] Zhang F, Zhong R, Li S, Chang RCC, Le W. The missing link between sleep disorders and age-related dementia: Recent evidence and plausible mechanisms. Journal of Neural Transmission. 2017;**124**:559-568

[17] Cox SR, Bak TH, Allerhand M, Redmond P, Starr JM, Deary IJ, et al. Bilingualism, social cognition and executive functions: A tale of chickens and eggs. Neuropsychologia. 2016;**91**:299-306

[18] Cao Y, Zhang LW, Wang J, Du SQ, Xiao LY, Tu JF, et al. Mechanisms of acupuncture effect on Alzheimer's disease in animal based researches. Current Topics in Medicinal Chemistry. 2016;**16**:574-578

[19] Leung MC, Yip KK, Ho YS, Siu FK, Li WC, Garner B. Mechanisms underlying the effect of acupuncture on cognitive improvement: A systematic review of animal studies. Journal of Neuroimmune Pharmacology. 2014;9:492-507

[20] Robbins JP, Price J. Human induced pluripotent stem cells as a research tool in Alzheimer's disease. Psychological Medicine. 2017;**47**:2587-2592

[21] Arber C, Lovejoy C, Wray S. Stem cell models of Alzheimer's disease: Progress and challenges. Alzheimer's Research & Therapy. 2017;**9**:42. DOI: 10.1186/s13195-017-0268-4)

[22] Peng X, Xing P, Li X, Qian Y, Song F, Bai Z, et al. Towards personalized intervention for Alzheimer's disease. Genomics, Proteomics & Bioinformatics. 2016;**14**:289-297

[23] Ho YS, Poon DCH, Chan TF, Chang RCC. From small to big molecules: How do we prevent and delay progression of aging-related neurodegeneration? Current Pharmaceutical Design. 2012;**18**:15-26 [24] Yu MS, Lai SW, Lin KF, Fang JN, Yuen WH, Chang RCC. Characterization of polysaccharides from the flowers of *Nerium indicum* and their neuroprotective effects. International Journal of Molecular Medicine. 2004;**14**:917-924

[25] Yu MS, Wong AYY, So KF, Fang JN, Yuen WH, Chang RCC. New polysaccharide from *Nerium indicum* protects neurons via stress kinase signaling pathway. Brain Research. 2007;**1153**:221-230

[26] Lai SW, Yu MS, Yuen WH, Chang RCC. Novel neuroprotective effects of the aqueous extracts from *Verbena officinalis Linn*. Neuropharmacology. 2006;**50**:641-650

[27] Lai CSW, Yu MS, Yuen WH, So KF, Zee SY, Chang RCC. Neuroprotective effects of anti-aging fungus *Gandoderma lucdium*. Brain Research.2008;**1190**:215-224

[28] Yu MS, Leung SKY, Che CM, Zee SY, So KF, Yuen WH, et al. Neuroprotective effects of an anti-aging Chinese medicine *Lycium barbarum* against  $\beta$ -amyloid peptide neurotoxicity. Experimental Gerontology. 2005;**40**:716-727

[29] Yu MS, Ho YS, So KF, Yuen WH, Chang RCC. Cytoprotective effects of *Lycium barbarum* against reducing stress on endoplasmic reticulum. International Journal of Molecular Medicine. 2006;**17**:1157-1162

[30] Ho YS, Yu MS, Lai CSW, So KF, Yuen WH, Chang RCC. Characterizing the neuroprotective effects of from *Lycium barbarum* on  $\beta$ -amyloid peptide neurotoxicity. Brain Research. 2007;**1158**:123-134

[31] Chan HC, Chang RCC, Ip AKC, Chiu K, Yuen WH, Zee SY, et al. Neuroprotective effects of *Lycium barbarum Lynn*, a traditional Chinese herbal medicine in protecting retinal ganglion cells in an ocular hypertension model of glaucoma. Experimental Neurology. 2007;**203**:269-273

[32] Chiu K, Zhou Y, Yeung SC, Lok CKM, Chan OOC, Chang RCC, et al. Up-regulation of crystallins is involved in neuroprotective effects of wolfberry on survival of retinal ganglion cells in rat ocular hypertension model. Journal of Cellular Biochemistry. 2010;**110**:311-320

[33] Li SY, Yang D, Yeung CM, Yu WY, Chang RCC, So KF, et al. *Lycium barbarum* polysaccharides reduce neuronal damage, blood-retinal barrier disruption and oxidative stress in retinal ischemia/reperfusion injury. PLoS One. 2011;**6**:e16380

[34] Chang RCC, So KF. Use of anti-aging herbal medicine, *Lycium barbarum*, against aging-associated diseases. What do we know so far? Cellular and Molecular Neurobiology. 2008;**28**:643-652

[35] Chao J, Yu MS, Ho YS, Wang M, Chang RCC. Dietary oxyresveratrol prevents parkinsonian mimetic
6-hydroxydopamine neurotoxicity.
Free Radical Biology & Medicine.
2008;45:1019-1026

[36] Chao J, Li H, Cheng KW, Yu MS, Chang RCC, Wang M. Protective effects of pinostilbene, a resveratrol methylated derivative, against 6-hydroxydopamineinduced neurotoxicity in SH-SY5Y cells. The Journal of Nutritional Biochemistry. 2010;**21**:482-489

[37] Chao J, Lau KW, Huie MJ, Ho YS, Yu MS, Lai CSW, et al. A prodrug of the green tea polyphenol (–)-epigallocatechin-3-gallate (EGCG) prevents differentiated SH-SY5Y cells from toxicity induced by 6-hydroxydopamine. Neuroscience Letters. 2010;**469**:360-364 [38] Poon DCH, Ho YS, Chiu K, Chang RCC. Cytokines: How important are they in mediating sickness? Neuroscience and Biobehavioral Reviews. 2013;**37**:1-10

[39] Poon CH, Ho YS, Chiu K, Wong HL, Chang RCC. Sickness: From the focus on cytokines, prostaglandins, and complement factors to the perspectives of neurons. Neuroscience and Biobehavioral Reviews. 2015;**57**:30-45

[40] Poon DCH, Ho YS, You R, Tse J, Chu K, Chang RCC. PKR deficiency alters E. Coli-induced sickness behaviors but does not exacerbate neuroimmune responses or bacterial load. Journal of Neuroinflammation. 2015;**12**:212. DOI: 10.1186/s12974-015-0433-2

[41] Huang CX, Irwin MG, Wong GTC, Chang RCC. Evidence of the impact of systemic inflammation on neuroinflammation from a nonbacterial endotoxin animal model. Journal of Neuroinflammation. 2018;**15**:147. DOI: 10.1186/ s12974-018-1163-z

[42] Lee JH, Jun HS. Role of myokines in regulating skeletal muscle mass and function. Frontiers in Physiology. 2018;**10**:42. DOI: 10.3389/ fphys.2019.00042

[43] Tanimura Y, Aoi W, Takanami Y, Kawai Y, Mizushima K, Naito Y, et al. Acute exercise increases fibroblast growth factor 21 in metabolic organs and circulation. Physiological Reports. 2016;4:e12828. DOI: 10.14814/ phy2.12828

[44] Fisher FM, Maratos-FlierE. Understanding the physiology of FGF21. Annual Review of Physiology.2016;78:223-241

[45] Barha CK, Davis JC, Falck RS, Nagamatsu LS, Liu-Ambrose T. Sex differences in exercise efficacy to improve cognition: A systematic review Introductory Chapter: Concept of Neuroprotection - A New Perspective DOI: http://dx.doi.org/10.5772/intechopen.85631

and meta-analysis of randomized controlled trials in older humans. Frontiers in Neuroendocrinology. 2017;**46**:71-85

[46] King DE, Carek P, Mainous AG 3rd, Pearson WS. Inflammatory markers and exercise: Differences related to exercise type. Medicine & Science in Sports & Exercise. 2003;**35**:575-581

[47] Kim YS, Shin SK, Hong SB, Kim HJ. The effects of strength exercise on hippocampus volume and functional fitness of older women. Experimental Gerontology. 2017;**97**:22-28

[48] Cassilhas RC, Lee KS, Fernandes J, Oliveira MG, Tufik S, Meeusen R, et al. Spatial memory is improved by aerobic and resistance exercise through divergent molecular mechanisms. Neuroscience. 2012;**202**:309-317

#### **Chapter 2**

## Current Therapeutic Approaches from Imidazoline and Opioid Receptors Modulators in Neuroprotection

Liliana Mititelu-Tartau, Maria Bogdan, Victor Gheorman, Liliana Foia, Ancuta Goriuc, Gabriela Rusu, Beatrice Buca, Liliana Pavel, Ana Cristofor, Cosmin-Gabriel Tartau and Gratiela Eliza Popa

#### Abstract

Due to brain plasticity, the nervous system is capable of manifesting behavioral variations, adapted to the influences from both external and internal environment. Multiple neurotransmitters are involved in the mediation of pathological processes at the molecular, cellular, regional, and interregional levels participating in cerebral plasticity, their intervention being responsible for various structural, functional, and behavioral disturbances. The current therapeutic strategies in neuroprotection aim at blocking on different levels, the molecular cascades of the pathophysiological mechanisms responsible for neuronal dysfunctions and ultimately for neuronal death. Different agents influencing these neurotransmitters have demonstrated beneficial effects in neurogenesis and neuroprotection, proved in experimental animal models of focal and global ischemic injuries. Serotonin, dopamine, glutamate, N-methyl-D-aspartate, and nitric oxide have been shown to play a significant role in modulating nervous system injuries. The imidazoline system is one of the important systems involved in human brain functioning. Experimental investigations have revealed the cytoprotective effects of imidazoline I2 receptor ligands against neuronal injury induced by hypoxia in experimental animals. The neuroprotective effects were also highlighted for kappa and delta receptors, whose agonists demonstrated the ability to reduce architectural lesions and to recover neuronal functions of animals with experimentally induced brain ischemia.

**Keywords:** neuroprotection, neurodegenerative diseases, ischemic stroke, imidazoline, opioids, nitric oxide

#### 1. Introduction

Increase in life expectancy has led to aging of the population and consequently to an expansion of the prevalence of neurodegenerative diseases (NDDs) [1].

Clinical type	Disease/disorder
Signs of progressive dementia	
with no other neurological signs	Alzheimer's disease
(absent/inconspicuous)	<ul> <li>Frontotemporal dementias</li> </ul>
	Some cases of Lewy-body disease
	Posterior cortical atrophy (visuospatial dementia)
Signs of progressive dementia	
accompanied by other neurological abnormalities	<ul> <li>Huntington's disease (chorea)</li> </ul>
	<ul> <li>Lewy-body disease (Parkinsonian features)</li> </ul>
	<ul> <li>Some cases of Parkinson's disease</li> </ul>
	<ul> <li>Corticobasal ganglionic degeneration</li> </ul>
	<ul> <li>Cortical-striatal-spinal degeneration (Jakob's disease)</li> </ul>
	Dementia-Parkinson-amyotrophic lateral sclerosis complex
	Cerebrocerebellar degeneration
	<ul> <li>Familial dementia with spastic paraparesis, amyotrophy, or</li> </ul>
	myoclonus
	Polyglucosan body disease
	Frontotemporal dementia with parkinsonism or ALS
Signs of movement disorders or	
other posture abnormalities	Parkinson's disease
	Multiple system atrophy
	Essential tremor
	Progressive supranuclear palsy
	Dystonia musculorum deformans
	<ul><li>Huntington's disease (chorea)</li><li>Acanthocytosis with chorea</li></ul>
	Corticobasal ganglionic degeneration
	<ul><li> Lewy-body disease</li><li> Restricted dystonia</li></ul>
Signs of progressive ataxia	·
	Spinocerebellar ataxias
	Cerebellar cortical ataxias
	Complicated hereditary and sporadic cerebellar ataxias
Signs of slowly developing	
muscular weakness and atrophy	<ul> <li>Motor disorders with amyotrophy</li> </ul>
	Spastic paraplegia without amyotrophy
Sensory and sensorimotor	
disorders	Hereditary sensorimotor neuropathies
	<ul> <li>Pure or predominantly sensory or motor neuropathic</li> </ul>
	Riley-Day autonomic degeneration
Signs of progressive blindness	
with or without other	Pigmentary degeneration of retina
neurological disorders	Stargardt's disease
	Age-related macular degeneration
Signs characterized by	
degenerative neurosensory deafness	Pure neurosensory deafness
	Hereditary hearing loss with retinal diseases
	<ul> <li>Hereditary hearing loss with system atrophies of the nervous</li> </ul>
	system

\*Adapted from Ropper A, Samuels M, Klein J. Adams and Victor's Principles of Neurology. 10th ed. McGraw-Hill Education; 2014.

#### Table 1.

Neurodegenerative diseases: main clinical types.

Neurodegeneration represents a loss of neurons and their structural components (dendrites, axons, and synapses) with a corresponding gradual atrophy in neuronal function [2].

#### Current Therapeutic Approaches from Imidazoline and Opioid Receptors Modulators... DOI: http://dx.doi.org/10.5772/intechopen.81951

NDDs (e.g., Alzheimer's disease, Parkinson's disease, Huntington's disease, dementia with Lewy bodies) determine cognitive and memory deterioration or alteration of the ability to move, speak, and breathe. These chronic and progressive disorders are an important cause of reduced quality of life, morbidity, caregiver burden and also, of the increase in total healthcare expenditure [3–5]. **Table 1** lists the main clinical types of NDDs [6].

Neuroprotection can be defined as a "relative preservation of neuronal structure and/or function" or as an action that aims "to prevent neuronal damage over time (either acute or chronic)" [7]. Neuroprotective action is primary if it is exerted directly on the neuron, or secondary if it appears from an activity on an intermediary that endangers neuronal function [8].

The mechanisms by which most agents with efficiency in NDDs act are not fully elucidated, requiring multiple and in-depth experimental and clinical studies.

Several experimental investigations highlight the multiple and various interrelations between adrenergic, serotoninergic, dopaminergic, glutamatergic, opioid, imidazoline systems, and the nitric oxide pathways, which may elucidate the effects of different compounds involved in the mediation of pathogenic mechanisms responsible for numerous structural, functional, and behavioral disturbances.

This chapter presents a brief overview of the most studied mechanisms related to neuroprotection and details the possibilities to pharmacologically influence through the main known neurotransmitters the pathophysiological mechanisms linked to various NDDs.

#### 2. The imidazoline system

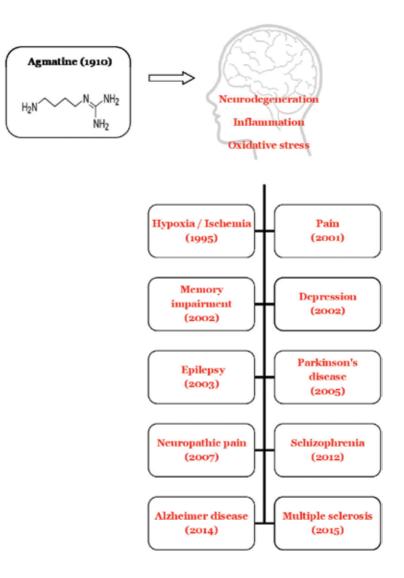
Imidazoline receptors are located not only in the mammalian central nervous system (CNS) cells but also in the peripheral nervous system [9], being involved in the mediation of various physiological processes in the body. It is currently known that there are four types of imidazoline receptors: I1, I2, I3, I4 (non I1-non I2), from which the first three have been mostly studied [10].

It has been emphasized that these receptors play an essential role in cell proliferation, regulation of adipose tissue formation, body temperature maintenance, mediation of gastrointestinal motility, neuroprotection, inflammation, nociceptive sensitivity, and some neurological or psychiatric disorders (such as depression) [11]. Moreover, it is known that these imidazoline receptor subtypes exert control over the activity of the hypothalamic-pituitary-adrenal and noradrenergic axis [12, 13].

A number of different endogenous ligands have been characterized: agmatine, the best known and largely studied, harmane and harmalane (derivatives of the beta-carboline group), and the newly discovered ribotide (acetic acid imidazole). Agmatine, the potent neurotransmitter of the imidazoline system, has an important role in the mediation of body's response to stress, analgesia, drug addiction, and abstinence syndrome, in modulation of seizures development, and in neuroprotection [14, 15].

Endogenous agmatine is produced in response to stress (in conditions of ischemia, prolonged exposure to cold) and/or to inflammation [16]. It is assumed that agmatine is also an effective neurotransmitter, due to its concentration in the brain similar to classical neurotransmitters [17, 18].

Literature data have revealed that agmatine stimulates the activity of endothelial nitric oxide synthase [16], this effect being also proved by its level in the rat brain after cerebral ischemia [19, 20].



#### Figure 1.

Neuroprotective effects of agmatine and their first discovery.

Along with evidence of its neuromodulatory and neuroprotective properties, there are numerous preclinical studies demonstrating the beneficial effects of exogenous administration of agmatine in depression, anxiety, hypoxic ischemia, pain, morphine tolerance, memory impairment, Parkinson's disease, Alzheimer's disease, epilepsy, and other related conditions with traumatic brain injuries (**Figure 1**) [21–23]. All these are arguments in favor of the potential of agmatine as a new pharmacological agent for the treatment of various neurological diseases and NDDs [24].

## 3. The involvement of the imidazoline system in the mediation of cognitive functions

Electrophysiological studies involving various brain areas, performed on laboratory animals with experimentally induced cerebral alterations, have demonstrated the neurotropic effects of agmatine [25].

*In vitro* experimental researches have shown that activation of I2 receptors via the agmatine endogenous ligand exerts neuroprotective effects by increasing the

#### Current Therapeutic Approaches from Imidazoline and Opioid Receptors Modulators... DOI: http://dx.doi.org/10.5772/intechopen.81951

expression of glial fibrillary acidic protein in astrocyte cultures and by inhibiting MAO activity [26, 27]. Moreover, the beneficial effects of agmatine have been observed on ischemic-hypoxic lesions, on glutamate-induced neurotoxicity by activating the imidazoline receptors [28, 29]. It was also demonstrated that agmatine administration improves learning activity and memory of rats in experimental models of Alzheimer's disease and streptozotocin-induced type two diabetes mellitus [22, 30].

Other experimental investigations highlight the neuroprotective effect of intranasal administration of agmatine in elderly female rats, with a significant improvement of neurological status and increase of survival rate [28, 29].

The neuroprotective effects of agmatine on the morphological changes determined by repeated induced stress on medial prefrontal cortex and hippocampus of the rat were also investigated [31]. It was emphasized that under constant stress conditions, morphological alterations of the brain are associated with the reduction of endogenous agmatine levels [measured by high-performance liquid chromatography (HPLC)] and with an increase of arginine-decarboxylase level in the prefrontal cortex, hippocampus, striatum, and hypothalamus [32].

The exogenous administration of agmatine lowers brain morphological impairment, suggesting thus its neuroprotective effects against structural changes in the rat brain, under recurrent stress circumstances [32]. Moreover, elevated levels of agmatine have been evidenced in the blood, cortex, hippocampus, and hypothalamus, immediately after brain hypoxic ischemia. Other studies emphasize the neuroprotective influences of agmatine, highlighted by the increase of its brain levels, in rats subjected to prolonged cold-exposure stress conditions [33].

The neuroprotective potential of agmatine was also highlighted in the experimental model of 1-methyl-4-phenyltetrahydropyridine (MPTP)-induced Parkinson's disease in mice [29]. The use of agmatine attenuates the loss of cellular dopamine from the black substance and repeated treatment improves short-term memory impairment induced by MPTP in elderly mice. The behavioral benefits of agmatine are associated with the decrease in MPTP-induced glutamate capture in the hippocampal area, suggesting thus its involvement in modulation of glutamate recapture, the possible mechanisms responsible for lowering glutamate extracellular levels, thereby alleviating its neurotoxicity [29].

It is known that alteration of spatial memory in Parkinson's disease and schizophrenia is attributed to several factors, including hypofunction of glutamate and reduction of hippocampal volume [34]. Literature data report that the administration of the N-methyl-D-aspartate (NMDA) receptor antagonists (phencyclidine, also coded MK801) frequently impairs the late alternation performance in a standardized behavioral model of cognitive functions alteration similar to schizophrenia in laboratory animals [34]. The use of the glutamate/NMDA receptor antagonist phencyclidine induces a spectrum of behavioral, neurochemical, and anatomical changes, manifested by locomotor hyperactivity, motor-negative deficits, and cognition alterations (with memory impairment and visual attention) in laboratory animals. This substance was used to induce the experimental schizophrenia in laboratory animals [35, 36]. Agmatine attenuates cognitive and behavioral deficiency in rats with experimental phencyclidine-induced schizophrenic manifestations [37].

The effects of agmatine on memory alterations similar to those found in Alzheimer's disease have been evaluated in rats; in the pathogenesis of this degenerative disorder (which causes cognitive deficits in rodents), the fragment beta amyloid A $\beta$ 2 25–35 plays an essential role. Studies have shown that agmatine significantly reduces the alterations in memory and spatial learning induced by the beta amyloid A $\beta$ 2 25–35 fragment (the neurotoxic component of beta amyloid A $\beta$  1–42) in various behavioral experimental models, such as: the swimming test, the radial arm maze test, and the object recognition test [38].

#### Neuroprotection

It has been revealed that agmatine diminishes the activation of hippocampal caspase-3 (the early indicator of neuronal apoptosis) and prevents the alteration of spatial memory induced by lipopolysaccharides, in the swimming test in rat [39], suggesting its neuroprotective effects.

The neurotropic activity of agmatine has been also evidenced in the structural and cognitive alterations after the administration of NMDA (N-methyl-D-aspartate) in rats [40]. The use of high performance liquid chromatography (HPLC) and electrochemical detection allowed highlighting that the treatment with NMDA is associated with low concentrations of monoamines (epinephrine, norepinephrine, dopamine, and serotonin) in rat PC12 cells [29, 40]. In this experimental model (swimming test), agmatine protects against NMDA-induced PC12 cell lesions, augmenting the levels of epinephrine, norepinephrine, and dopamine, but not influencing serotonin values, together with lowering intracellular Ca<sup>2+</sup> overload. These results indicate that the neuroprotective action of agmatine may be related to NMDA-receptor modulation and/or to controlling the decrease in monoamine content and NMDA-induced intracellular Ca<sup>2+</sup> overload [40].

Immunohistochemical studies and electrophysiological investigations performed on the brain have validated the neuroprotective actions of both imidazoline receptor antagonists idazoxan and efaroxan in rats with cerebral damages caused by the use of quinolinic acid [41], and also in mice with experimentally induced autoimmune encephalomyelitis, confirming the improvement of brain structural alterations and blood brain barrier lesions curtail [42].

A new (+)2-(ethyl-2,3-dihydrobenzofuranyl)-2-imidazoline derivative dexefaroxan—the (+) enantiomer of efaroxan has been characterized. It has a potent and selective  $\alpha$ 2 antagonist activity, with facultative effects on cognitive functions in the passive avoidance test in rats with memory-deficiency induced by scopolamine, diazepam, or by the 2-adrenergic agonist UK 14,304.

Dexefaroxan improves the cognitive performances in the passive avoidance test, facilitates spatial memory in the Morris swimming test in rats, and increases the object recognition ability in the specific behavioral test in mice [43, 44]. It has also proved to ameliorate the animal's memory deficits in these tests, particularly by attenuation of spontaneous memory loss, and to improve their spatial recognition ability, rather than through the acquisition skills or various other non-cognitive effects.

After subcutaneous administration of dexefaroxan, its pharmacodynamic effects persist for about 21–25 days, indicating that tolerance does not occur during prolonged treatment. Moreover, it was emphasized that dexefaroxan exerts protective effects on the spatial memory deficit caused by cortical devascularization in the Morris swimming test in rats [43].

Dexefaroxan has also been shown to exhibit neuroprotective effects on the devascularization-induced neurodegeneration, to ameliorate the structural changes in the hippocampus, and to remove the cognitive deficits induced by cerebral ischemia in rats [45, 46]. Its neuroprotective effects were present also in the excitotoxic lesions produced at the region of the basal magnocellular nucleus, increasing the olfactory discriminative capacity of rats, suggesting thus the possibility of its use in the treatment of memory disturbances in Alzheimer's disease [43].

Studies performed on genetically modified animals revealed that dexefaroxan improves cognitive performance in knockout mice with Alzheimer's disease [47].

Literature data regarding the neuroprotective action of imidazoline agonist and antagonist agents in human studies are only few, and the mechanisms involved in these effects are not completely deciphered.

Some investigations suggested that agmatine manifests protective activity against brain cell injury in different *in vivo* models of Parkinson's disease, as well as

Current Therapeutic Approaches from Imidazoline and Opioid Receptors Modulators... DOI: http://dx.doi.org/10.5772/intechopen.81951

*in vitro* studies, performed on human-derived dopaminergic neuroblastoma cell lines. It was postulated that the neuromodulatory properties of agmatine are related to the protective effects on the dopaminergic neurons, to NMDA receptor blocking, and to the decrease in oxidative stress, due to the inhibition of nitric oxide synthase (NOS) activity [48, 23].

Other clinical trials highlighted that the treatment with agmatine was associated with cytoprotective actions, in patients with spinal cord injury, proved by lessening the glial weal construction, decreasing the collagen scar zone, relieving the neuronal alterations, and recovering remyelination [49]. Moreover, the beneficial effects of agmatine have been demonstrated in various CNS lesions such as: cerebrovascular accident, brain trauma, neuropathic pain, lumbar degenerative disc disease, and different other types of neuropathy [50–52].

The administration of dexmedetomidine has also shown neuroprotective effects in humans with acute cerebral lesions [53].

In patients with dementia due to brain frontal lesions, idazoxan alleviates attentional and executive dysfunctions evoked by classical cognitive function tests [54].

## 4. The interrelation between the adrenergic and the imidazoline systems in the mediation of cognitive functions

Clonidine, both a non-specific  $\alpha$ 2 adrenergic and imidazoline receptor agonist, decreases the cognitive function alterations induced by phencyclidine and MK801, facilitating spatial memory in the radial arm maze test in rats [35], but does not influence the behavioral and cognitive deficits in the experimental NMDA-induced excitotoxic dorsal hippocampal lesions [35]. Such findings indicate that clonidine improves memory alterations caused by glutamate hypofunction, but not by hippocampal injury, implying that multiple and distinct mechanisms are involved in the development of memory disorders.

The administration of the  $\alpha 2$  adrenergic receptor agonist clonidine or guanfacine prevents some of the behavioral effects of NMDA antagonists, proving that the monoaminergic system mediates a number of aspects of the cognitive deficit. Clonidine and guanfacine improve the lack of visual attention and spatial memory induced by phencyclidine in rats [34, 36]. It was demonstrated that low doses of clonidine recover the animal's ability to accurately choose the object and prevent the performance deficit induced by phencyclidine. At high doses, clonidine decreases the response time and induces a lack of the choice accuracy. These results indicate that clonidine treatment can alleviate phencyclidine-induced deficit of attention and of working memory, probably by preventing some of the neurochemical and anatomical effects of this psychotomimetic drug [34].

On the other hand, the use of only the selective  $\alpha 2$  adrenergic receptor antagonist does not impair the animal's spatial memory, but dramatically aggravates the phencyclidine-induced memory deficit [36]. These data demonstrate that  $\alpha 2$  adrenergic receptors mediate the inhibition of spatial memory disturbances, suggesting their important role in cognitive deficits associated to NMDA receptor hypofunction [34, 36].

The role of moxonidine (an  $\alpha$ 2 adrenergic imidazoline I1 receptor agonist) on cognitive function in rats with Huntington's disease experimentally induced with 3-nitropropionic acid (3-NPA) was investigated in the Morris swimming test and in the elevated plus maze test. The administration of 3-NPA induces degenerative brain damage, progressive motor dysfunction, loss of grip force, emotional disturbances, weight loss, anxiety, and impairment of learning activity and memory. An increase in cerebral acetylcholinesterase level, enhancement of oxidative stress, and

impairment of the activity of mitochondrial enzyme complexes I, II, and IV were also noted [28]. The treatment with moxonidine resulted in the alleviation of disturbances caused on animal weight, motor activity, gripping ability, anxiety, impairment of learning ability and memory, and biochemical disturbances, thus indicating that substances modifying the activity of I1 receptors may be potential pharmacological agents for the treatment of degenerative brain disorders [55].

The effects of clonidine have also been evaluated in mice with subacute brain ischemia obtained after permanent ligation of common carotid arteries. The subsequent brain damages consisted of expansion of cerebral infarction areas, assessed by computed tomography scans. This experimentally induced chronic cerebral hypoperfusion was associated with a significant impairment of animal's learning ability and memory in the Morris swimming test [25, 28]. Subacute treatment with clonidine for 7 days increases the expression of neuronal nuclei, glutamic acid-decarboxylase-67, and gamma-aminobutyric acid (GABA) B receptor (GABA<sub>B</sub>1) in hippocampal subregion cornu amonis (CA<sub>1</sub>) but does not influence the level of these elements in the hippocampal area CA<sub>3</sub>, nor in the dentate gyrus. These data support the idea that clonidine exerts neuroprotective effects on chronic cerebral ischemic lesions, by regulating GABA<sub>B</sub>1 receptors and the activity of glutamic acid-decarboxylase-67 [25].

Additionally, the decrease in superoxide dismutase (SOD), catalase (CAT), and glutathione levels as well as the increase of both malondialdehyde (MDA) level and cerebral acetylcholinesterase activity were noted in animals with brain ischemic lesions [28].

Both moxonidine and clonidine have shown a decrease in histopathological changes, oxidative stress, central cholinesterase activity, as well as a reduction in memory disturbances and learning deficits in mice with vascular dementia induced by subacute ischemia after permanent bilateral cerebral artery ligation [28, 40].

*In vitro* cell culture studies from the rat frontal cortex with glutamate-induced neurotoxicity revealed the partial neuroprotective effects of moxonidine, with a significant decrease in the number of dead cells [26]. Moxonidine has shown beneficial effects on cerebral spasm in an experimental rabbit model of subarachnoid hemorrhage [40, 56].

### 5. The interrelations between the imidazoline system and the oxidative stress in the mediation of cognitive functions

Different pathological conditions of the body, as well as the physiological process of aging, can cause cognitive impairment and free oxygen radicals production, being responsible for abnormal functioning and cell death. Subsequently, a new idea has emerged claiming that nitric oxide (NO), along with the free radicals, plays a key role in the aging process, due to neurotoxic effects on the brain exerted by its excessive levels [57]. Nitric oxide is generated from L-arginine under the action of nitric oxide synthase (NOS). The three isoforms of NOS have different roles in the body: neuronal NOS (nNOS) is responsible for synaptic plasticity, learning and memory processes; endothelial NOS (eNOS) provides stabilization and regulation of vascular micro-environment and contributes to neuroplasticity [58]; and inducible NOS (iNOS) is involved in various pathophysiological conditions [57].

Numerous experimental researches reveal that NOS activity is significantly elevated in the brain of elderly rats, being associated with existing cognitive alterations [57, 59]. Mediated by the competitive inhibition of nNOS and iNOS, and correlated with the stimulation of NOS, agmatine contributes to the improvement of cognitive functions [19, 60, 61], while exhibiting neuroprotective effects [38, 39].

#### Current Therapeutic Approaches from Imidazoline and Opioid Receptors Modulators... DOI: http://dx.doi.org/10.5772/intechopen.81951

Literature data have shown that agmatine eliminates neuroinflammation and lipopolysaccharide-induced memory impairment (which is known to stimulate iNOS activity and, implicitly, the NO production) in laboratory animals. It prevents cognitive alterations, probably as a result of inhibition of iNOS activity [39]. Other researchers have disproved these results by showing that agmatine can cause cognitive impairment due to the inhibition of NMDA receptors and of NO, important elements in the modulation of learning and memory processes [62, 63].

On the other hand, it is known that the central cholinergic system plays a crucial role in the mediation of cognitive functions. Cognitive deficits have been induced in laboratory animals by using an anticholinergic agent, scopolamine, its administration producing a significant reduction in NOS activity, and an increase in arginase activity, of L-ornithine and putrescine levels in the hippocampus [51]. It has been observed that agmatine eliminates the scopolamine-induced alterations of memory and learning capacity [64, 65].

Although glutamatergic activity is required for cognitive processes, it is assumed that the increase of glutamate levels or of NMDA activity would also be responsible for the scopolamine-induced cognitive disturbances [66].

Knowing that agmatine blocks NMDA receptors and also interferes with the pathways of NO, NOS, and L-arginine, it was assumed that the removal of scopolamine-induced cognitive deficits can be attributed to its modulating effect on NO/NOS activity, on L-arginine, and also to the antagonization of NMDA receptors, with subsequent suppression of excessive glutamatergic activity [61, 65].

Abnormal release and disturbances of neuromodulatory activities due to variation in cerebral agmatine levels may be correlated to different CNS diseases (such as schizophrenia). Interactions of agmatine with other central neurotransmitter systems (such as glutamatergic and nitrergic) appear to be particularly important in the pathophysiological mechanisms of CNS disorders associated with brain damage and cognitive functions deficit.

#### 6. The opioid system

Neurodegeneration can be caused by chronic disease progression or by acute injury (cerebral ischemia—stroke or trauma) [67]. Ischemic stroke represents a vascular ailment with neurological consequences produced by the obstruction of the arteries in a part of the brain, therefore by blood supply privation [68]. Stroke can determine long-term neurological and psychiatric impairments, its therapy being focused on confining secondary injury processes [67].

In a recent review article, Chamorro presented that ischemic stroke is "the first cause of permanent disability in adult people, the second single most frequent cause of death for people older than 60 years, the second most common cause of dementia, representing approximately 3% to 7% of the total health-care expenditure in high income countries" [69].

A superpose of pathologies in different neurological disorders was proposed since NDDs and ischemic stroke are frequently concomitant, hence the neuroprotective therapy could be similar [70].

Opioids are substances with morphine-like action binding to specific opioid receptors (ORs). In the early 1990s, three important opioid receptor families [ $\mu$  (MOR),  $\kappa$  (KOR), and  $\delta$  (DOR)] were identified, and in 1994 another opioid receptor was discovered [nociceptin, orphanin FQ receptor (NOP), or the opioid receptor-like orphan receptor (ORL)]. ORs are found in the nervous system, lungs, heart, liver, and gastrointestinal and reproductive tracts. They have been intensively studied and it was emphasized that they not only are related to

Opioid receptor	Agonist
δ	DADLE [D-Ala2, D-Leu5]-enkephalin
	DPDPE (D-Pen2,D-Pen5)-enkephalin
	SNC80
	Tan-67
	Remifentanil
ĸ	BRL 52537
	CI-977
	GR89696
	Salvinorin A
	U-50,488H
	Dynorphin
μ	DAMGO [D-Ala2,N-MePhe4,Gly-ol]-enkephalin
	Endorphin 1 and 2 (EM 1/2),
	Morphine

#### Table 2.

Opioid receptors and their agonists tested for neuroprotective action.

antinociceptive action, but also have a role in cell proliferation, ionic homeostasis, emotional response, immune function, epileptic seizures, feeding, obesity, respiratory and cardiovascular control, hibernation, and neuroprotection [71, 72].

In the last decades, researches have pointed out that the opioid system can be promising to get neuroprotective treatment in the event of stroke, through OR agonists at lower doses, to avoid tolerance and/or physical dependency. DOR agonists followed by KOR agonists have revealed the most intense neuroprotective efficacy [64]. Major OR agonists tested for neuroprotection are listed in **Table 2** [71, 72].

DOR activation is beneficial against ischemic, hypoxic, and excitotoxic injuries [73]; recent studies promote DOR and especially DADLE (an analog of endogenous delta-opioid enkephalin) as promising targets for treating NDDs like stroke and PD [74–76].

DADLE alleviates apoptotic pathways, supports not only cell survival of peripheral organs (such as lung, heart, kidney, and liver) but also neuronal survival, and protects neurons and glial cells from ischemia-induced cell death [76–78].

In a cellular model of PD, DADLE administration augmented cell survivability with concurrent downregulation of the unfolded protein response stress sensors and protein aggregates [79].

Findings from a rat middle cerebral artery occlusion (MCAO) stroke model proposed that neuroprotection of DADLE treatment was based on the activation of PI3K-Akt pathway by reducing nerve cell apoptosis [80].

Non-selective opioid receptor agonists were also tested: LYS739 (fluorinated enkephalin-fentanyl derivative) and the most promising compound—biphalin— which proved to be effective both *in vitro* and *in vivo* stroke models [72].

The latter is a dimeric enkephalin analog (Tyr-D-Ala-Gly-Phe-NH-)2 with high potency and affinity for MOR and DOR and low affinity for KOR. Biphalin crosses blood-brain barrier reaching spinal and supraspinal sites expressing OR and produces less physical dependence and tolerance compared to morphine [72, 81–84].

Current Therapeutic Approaches from Imidazoline and Opioid Receptors Modulators... DOI: http://dx.doi.org/10.5772/intechopen.81951

Different studies using a mouse MCAO stroke model reported that biphalin reduced brain edema and infarction, ROS production, and NMDA-induced excitotoxicity. It also increased locomotor activities and neurological score after stroke resembled to saline-treated animals [72, 81, 85, 86]. Biphalin notably diminished penumbral expression of Na<sup>+</sup>, K<sup>+</sup>, 2Cl<sup>-</sup> cotransporter (NKCC), and the translocation of the conventional isoforms of protein kinase C [81].

It has been hypothesized that biphalin's neuroprotective effects are more intense compared to subtype-selective agonists due to concomitant activation of the three types of OR [72, 85].

# 7. Conclusion

Various pharmacological substances influencing the pathways of the main neurotransmitters have confirmed valuable effects in neurogenesis and neuroprotection, being validated in different *in vitro* researches and *in vivo* experimental animal models of limited or extensive ischemic brain lesions.

Deciphering the roles of the neurotransmitters in central nervous system activity other than the signaling function will represent a starting point to deepen the knowledge about the complex mechanisms of the brain functions and to obtain new agents useful for protection of ischemic neurons and for preventing their irreversible damage.

# **Author details**

Liliana Mititelu-Tartau<sup>1†</sup>, Maria Bogdan<sup>2\*†</sup>, Victor Gheorman<sup>3</sup>, Liliana Foia<sup>1</sup>, Ancuta Goriuc<sup>1</sup>, Gabriela Rusu<sup>1</sup>, Beatrice Buca<sup>1</sup>, Liliana Pavel<sup>4</sup>, Ana Cristofor<sup>1</sup>, Cosmin-Gabriel Tartau<sup>1</sup> and Gratiela Eliza Popa<sup>5†</sup>

1 Faculty of Medicine, "Gr. T. Popa" University of Medicine and Pharmacy, Iasi, Romania

2 Faculty of Pharmacy, University of Medicine and Pharmacy, Craiova, Romania

3 Faculty of Medicine, University of Medicine and Pharmacy of Craiova, Romania

4 Faculty of Medicine, "Dunarea de Jos" University of Medicine and Pharmacy, Galati, Romania

5 Faculty of Pharmacy, "Gr. T. Popa" University of Medicine and Pharmacy, Iasi, Romania

\*Address all correspondence to: bogdanfmaria81@yahoo.com

† These authors contributed equally to this work.

# IntechOpen

© 2019 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/ by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. *Current Therapeutic Approaches from Imidazoline and Opioid Receptors Modulators...* DOI: http://dx.doi.org/10.5772/intechopen.81951

# References

[1] Jaafaru MS, Abd Karim NA, Enas ME, Rollin P, Mazzon E, Abdull Razis AF. Protective effect of glucosinolates hydrolytic products in neurodegenerative diseases (NDDs). Nutrients. 2018;**10**(5), 580:1-15

[2] Cummings J. Disease modification and neuroprotection in neurodegenerative disorders. Translational Neurodegeneration. 2017;**6**:25

[3] Lauterbach EC, Victoroff J, Coburn KL, Shillcutt SD, Doonan SM, Mendez MF. Psychopharmacological neuroprotection in neurodegenerative disease: Assessing the preclinical data. The Journal of Neuropsychiatry and Clinical Neurosciences. 2010;**22**:8-18

[4] Gitler AD, Dhillon P, Shorter J. Neurodegenerative disease: Models, mechanisms, and a new hope. Disease Models & Mechanisms. 2017;**10**:499-502

[5] Gao HM, Hong JS. Why neurodegenerative diseases are progressive: Uncontrolled inflammation drives disease progression. Trends in Immunology. 2008;**29**:357-365

[6] Ropper A, Samuels M, Klein J. Adams and Victor's Principles of Neurology. 10th ed. Boston: McGraw-Hill Education; 2014. pp. 1060-1131

[7] Wiendl H, Elger C, Förstl H, Hartung H-P, Oertel W, Reichmann H, et al. Gaps between aims and achievements in therapeutic modification of neuronal damage ("Neuroprotection"). Neurotherapeutics. 2015;**12**:449-454

[8] Cummings J, Fox N. Defining disease modifying therapy for Alzheimer's disease. The Journal of Prevention of Alzheimer's Disease. 2017;**4**:109-115

[9] Ferrari F, Fiorentino S, Mennuni L, Garofalo P, Letari O, Mandelli S, et al. Analgesic efficacy of CR4056, a novel imidazoline-2 receptor ligand, in rat models of inflammatory and neuropathic pain. Journal of Pain Research. 2011;**4**:111-125

[10] Nechifor M. Imidazoline receptors— Normal and pathological factors. Revista Medico-Chirurgicala a Societatii de Medici Si Naturalisti din Iasi. 2001;**105**: 438-443

[11] Dardonville C, Rozas I, Callado LF, Meana JJ. I(2)-imidazoline binding site affinity of a structurally different type of ligands. Bioorganic & Medicinal Chemistry. 2002;**10**:1525-1533

[12] Pypendop BH, Silverstein D,
Hopper K. 2 agonists and antagonists. In:
Small Animal Critical Care Medicine.
2nd ed. Missouri: Saunders Inc.; 2015.
pp. 866-871.Ch 165. SRC - BaiduScholar

[13] Thorn DA, An X-F, Zhang Y, Pigini M, Li J-X. Characterization of the hypothermic effects of imidazoline I2 receptor agonists in rats. British Journal of Pharmacology. 2012;**166**:1936-1945

[14] Smith KL, Jessop DS, Finn DP. Modulation of stress by imidazoline binding sites: Implications for psychiatric disorders. Stress (Amsterdam, Netherlands). 2009;**12**:97-114

[15] Ciubotariu D, Nechifor M. Involvement of imidazoline system in drug addiction. Revista Medico-Chirurgicala a Societatii de Medici Si Naturalisti din Iasi. 2012;**116**:1118-1122

[16] Halaris A, Plietz J. Agmatine: Metabolic pathway and spectrum of activity in brain. CNS Drugs. 2007;**21**: 885-900

[17] Haenisch B, von Kügelgen I, Bönisch H, Göthert M, Sauerbruch T, Schepke M, et al. Regulatory mechanisms underlying agmatine homeostasis in humans. American Journal of Physiology. Gastrointestinal and Liver Physiology. 2008;**295**:G1104-G1110

[18] Reis DJ, Regunathan S. Is agmatine a novel neurotransmitter in brain? Trends in Pharmacological Sciences. 2000;**21**: 187-193

[19] Mun CH, Lee WT, Park KA, Lee JE. Regulation of endothelial nitric oxide synthase by agmatine after transient global cerebral ischemia in rat brain. Anatomy & Cell Biology. 2010;**43**: 230-240

[20] Nissim I, Horyn O, Daikhin Y, Chen P, Li C, Wehrli SL, et al. The molecular and metabolic influence of long term agmatine consumption. The Journal of Biological Chemistry. 2014;**289**: 9710-9729

[21] Mancinelli F, Ragonese F, Cataldi S, Ceccarini MR, Iannitti RG, Arcuri C, et al. Inhibitory effects of agmatine on monoamine oxidase (MAO) activity: Reconciling the discrepancies. The EuroBiotech Journal. 2018;**2**:121-127

[22] Sirvanci-Yalabik M, Sehirli AO, Utkan T, Aricioglu F. Agmatine, a metabolite of arginine, improves learning and memory in streptozotocininduced Alzheimer's disease model in rats. Bulletin of Clinical Psychopharmacology. 2016;26(4): 342-354

[23] Neis VB, Rosa PB, Olescowicz G, Rodrigues ALS. Therapeutic potential of agmatine for CNS disorders. Neurochemistry International. 2017; 108:318e331

[24] Moretti M, Neis VB, Matheus FC, Cunha MP, Rosa PB, Ribeiro CM, et al. Effects of agmatine on depressive-like behavior induced by intracerebroventricular administration of 1-methyl-4-phenylpyridinium (MPP (+)). Neurotoxicity Research. 2015;**28**: 222-231 [25] Lu Y, Li C, Zhou M, Luo P, Huang P, Tan J, et al. Clonidine ameliorates cognitive impairment induced by chronic cerebral hypoperfusion via upregulation of the GABA<sub>B</sub>R1 and GAD67 in hippocampal CA1 in rats. Pharmacology, Biochemistry, and Behavior. 2015;**132**:96-102

[26] Bakuridze K, Savli E, Gongadze N, Baş DB, Gepdiremen A. Protection in glutamate-induced neurotoxicity by imidazoline receptor agonist moxonidine. The International Journal of Neuroscience. 2009;**119**:1705-1717

[27] Head GA, Mayorov DN. Imidazoline receptors, novel agents and therapeutic potential. Cardiovascular & Hematological Agents in Medicinal Chemistry. 2006;**4**:17-32

[28] Gupta S, Sharma B. Pharmacological modulation of I(1)-imidazoline and α2adrenoceptors in sub acute brain ischemia induced vascular dementia. European Journal of Pharmacology. 2014;**723**:80-90

[29] Matheus FC, Aguiar AS, Castro AA, Villarinho JG, Ferreira J, Figueiredo CP, et al. Neuroprotective effects of agmatine in mice infused with a single intranasal administration of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP). Behavioural Brain Research. 2012;**235**:263-272

[30] Kang S, Kim C-H, Jung H, Kim E, Song H-T, Lee JE. Agmatine ameliorates typ. 2 diabetes induced-Alzheimer's disease-like alterations in high-fat diet-fed mice via reactivation of blunted insulin signalling. Neuropharmacology. 2017;**113**:467-479

[31] Aricioglu F, Ercil E, Dulger G. Agmatine inhibits naloxone-induced contractions in morphine-dependent Guinea pig ileum. Annals of the New York Academy of Sciences. 2003;**1009**: 147-151 *Current Therapeutic Approaches from Imidazoline and Opioid Receptors Modulators...* DOI: http://dx.doi.org/10.5772/intechopen.81951

[32] Zhu M-Y, Wang W-P, Cai Z-W, Regunathan S, Ordway G. Exogenous agmatine has neuroprotective effects against restraint-induced structural changes in the rat brain. The European Journal of Neuroscience. 2008;**27**: 1320-1332

[33] Aricioglu F, Regunathan S.
Agmatine attenuates stress- and lipopolysaccharide-induced fever in rats. Physiology & Behavior. 2005;85: 370-375

[34] Jeutsch JD, Anzivino LA. A low dose of the alpha2 agonist clonidine ameliorates the visual attention and spatial working memory deficits produced by phencyclidine administration to rats. Psychopharmacology (Berl). 2004;**175**: 76-83

[35] Bardget ME, Points M, Ramsey-Faulkner C. The effects of clonidine on discrete-trial delayed spatial alternation in two rat models of memory loss. Neuropsychopharmacology. 2008;**33**: 1980-1991

[36] Marrs W, Kuperman J, Avedian T, Roth RH, Jentsch JD. Alpha-2 adrenoceptor activation inhibits phencyclidine-induced deficits of spatial working memory in rats. Neuropsychopharmacology. 2005;**30**: 1500-1510

[37] Unal G, Ates A, Aricioglu F. Agmatine-attenuated cognitive and social deficits in subchronic MK-801 model of schizophrenia in rats. Psychiatry and Clinical Psychopharmacology. 2018:1-10

[38] Bergin DH, Liu P. Agmatine protects against beta-amyloid25–35induced memory impairments in the rat. Neuroscience. 2010;**169**:794-811

[39] Zarifkar A, Choopani S, Ghasemi R, Naghdi N, Maghsoudi AH, Maghsoudi N, et al. Agmatine prevents LPSinduced spatial memory impairment and hippocampal apoptosis. European Journal of Pharmacology. 2010;**634**: 84-88

[40] Li YF, Gong ZH, Cao JB, Wang HL, Luo ZP, Li J. Antidepressant-like effect of agmatine and its possible mechanism. European Journal of Pharmacology. 2003;**469**(1–3):81-88

[41] Martel J, Chopin P, Colpaert F, Marien M. Neuroprotective effects of the alpha2-adrenoceptor antagonists, (+)-efaroxan and (+/-)-idazoxan, against quinolinic acid-induced lesions of the rat striatum. Experimental Neurology. 1998;**154**:595-601

[42] Wang X-S, Fang H-L, Chen Y, Liang S-S, Zhu Z-G, Zeng Q-Y, et al. Idazoxan reduces blood-brain barrier damage during experimental autoimmune encephalomyelitis in mouse. European Journal of Pharmacology. 2014;**736**: 70-76

[43] Chopin P, Debeir T, Raisman-Vozari R, Colpaert FC, Marien MR. Protective effect of the alpha2adrenoceptor antagonist, dexefaroxan, against spatial memory deficit induced by cortical devascularization in the adult rat. Experimental Neurology. 2004;**185**: 198-200

[44] Pauwels PJ, Rauly I, Wurch T. Dissimilar pharmacological responses by a new series of imidazoline derivatives at precoupled and ligand-activated alpha 2A-adrenoceptor states: Evidence for effector pathway-dependent differential antagonism. The Journal of Pharmacology and Experimental Therapeutics. 2003;**305**:1015-1023

[45] Rizk P, Salazar J, Raisman-Vozari R, Marien M, Ruberg M, Colpaert F, et al. The alpha2-adrenoceptor antagonist dexefaroxan enhances hippocampal neurogenesis by increasing the survival and differentiation of new granule cells. Neuropsychopharmacology. 2006;**31**: 1146-1157 [46] Debeir T, Marien M, Ferrario J, Rizk P, Prigent A, Colpaert F, et al. In vivo upregulation of endogenous NGF in the rat brain by the alpha2-adrenoreceptor antagonist dexefaroxan: Potential role in the protection of the basalocortical cholinergic system during neurodegeneration. Experimental Neurology. 2004;**190**:384-395

[47] Francis BM, Yang J, Hajderi E, Brown ME, Michalski B, McLaurin J, et al. Reduced tissue levels of noradrenaline are associated with behavioral phenotypes of the TgCRND8 mouse model of Alzheimer's disease. Neuropsychopharmacology. 2012;**37**: 1934-1944

[48] Condello S, Monica Currò M, Ferlazzo N, Caccamo D, Satriano J, Lentile R. Agmatine effects on mitochondrial membrane potential and NF-κB activation protect against rotenone-induced cell damage in human neuronal-like SH-SY5Y cells. Journal of Neurochemistry. 2011;**116**(1):67-75

[49] Park YM, Lee WT, Bokara KK, Seo SK, Park SH, Kim JH, et al. The multifaceted effects of agmatine on functional recovery after spinal cord injury through modulations of BMP-2/4/7 expressions in neurons and glial cells. PLoS One. 2013;**8**:e53911

[50] Tohidi V, Hassanzadeh B, Sherwood B, Ma W, Rosenberg M, Gilad V, et al.
Effect of agmatine sulfate on neuropathic pain. Neurology. 2014;82 (Suppl. 10):P7.094

[51] Piletz JE, Aricioglu F, Cheng JT, Fairbanks CA, Gilad VH, Haenisch B, et al. Agmatine: Clinical applications after 100 years in translation. Drug Discovery Today. 2013;**18**(17–18): 880-893

[52] Keynan O, Mirovsky Y, Dekel S, Gilad VH, Gilad GM. Safety and efficacy of dietary agmatine sulfate in lumbar disc-associated radiculopathy. An openlabel, dose-escalating study followed by a randomized, double-blind, placebocontrolled trial. Pain Medicine. 2010; **11**(3):356-368

[53] Himmelseher S, Kochs EF.Neuroprotection by dexmedetomidine.In: Gullo A, editor. Anaesthesia, Pain,Intensive Care and EmergencyA.P.I.C.E. Italia: Springer-Verlag; 2006.pp. 633-639. Ch. 56

[54] Coull JT, Sahakian BJ, Hodges JR. The  $\alpha$ 2 antagonist idazoxan remediates certain attentional and executive dysfunction in patients with dementia of frontal type. Psychopharmacology. 1996;**123**:239-249

[55] Gupta S, Sharma B. Pharmacological benefit of I(1)-imidazoline receptors activation and nuclear factor kappa-B (NF- $\kappa$ B) modulation in experimental Huntington's disease. Brain Research Bulletin. 2014;**102**:57-68

[56] Ilik MK, Kocaogullar Y, Koc O, EsenH. Beneficial effects of moxonidine on cerebral vasospasm after experimental subarachnoid. Turkish Neurosurgery.2014;24(6):873-879

[57] Liu P, Smith PF, Appleton I, Darlington CL, Bilkey DK. Potential involvement of NOS and arginase in age-related behavioural impairments. Experimental Gerontology. 2004;**39**: 1207-1222

[58] Rushaidhi M, Collie ND, Zhang H, Liu P. Agmatine selectively improves behavioural function in aged male Sprague-Dawley rats. Neuroscience. 2012;**218**:206-215

[59] Gupta N, Jing Y, Collie ND, Zhang H, Liu P. Ageing alters behavioural function and brain arginine metabolism in male Sprague-Dawley rats. Neuroscience. 2012;**226**:178-196

[60] Liu P, Jing Y, Collie ND, Chary S, Zhang H. Memory-related changes in

*Current Therapeutic Approaches from Imidazoline and Opioid Receptors Modulators...* DOI: http://dx.doi.org/10.5772/intechopen.81951

L-citrulline and agmatine in the rat brain. Hippocampus. 2009;**19**:597-602

[61] Moosavi M, Khales GY, Abbasi L, Zarifkar A, Rastegar K. Agmatine protects against scopolamine-induced water maze performance impairment and hippocampal ERK and Akt inactivation. Neuropharmacology. 2012;**62**(5–6):2018-2023

[62] Stewart LS, McKay BE. Acquisition deficit and time-dependent retrograde amnesia for contextual fear conditioning in agmatine-treated rats. Behavioural Pharmacology. 2000;**11**: 93-97

[63] McKay BE, Lado WE, Martin LJ, Galic MA, Fournier NM. Learning and memory in agmatine-treated rats. Pharmacology, Biochemistry, and Behavior. 2002;**72**:551-557

[64] Utkan T, Gocmez SS, Regunathan S, Aricioglu F. Agmatine, a metabolite of L-arginine, reverses scopolamineinduced learning and memory impairment in rats. Pharmacology, Biochemistry, and Behavior. 2012;**102**: 578-584

[65] Knox LT, Jing Y, Fleete MS, Collie ND, Zhang H, Liu P. Scopolamine impairs behavioural function and arginine metabolism in the rat dentate gyrus. Neuropharmacology. 2011;**61**: 1452-1462

[66] Barber TA, Haggarty MK. Memantine ameliorates scopolamineinduced amnesia in chicks trained on taste-avoidance learning. Neurobiology of Learning and Memory. 2010;**93**: 540-545

[67] Nguyen L, Lucke-Wold BP, Mookerjee S, Kaushal N, Matsumoto RR. Sigma-1 receptors and neurodegenerative diseases: Towards a hypothesis of sigma-1 receptors as amplifiers of neurodegeneration and neuroprotection. Advances in Experimental Medicine and Biology. 2017;**964**:133-152

[68] Xiong XY, Liu L, Yang QW. Refocusing neuroprotection in cerebral reperfusion era: New challenges and strategies. Frontiers in Neurology. 2018; **9**:249

[69] Chamorro A. Neuroprotectants in the era of reperfusion therapy. Journal of Stroke. 2018;**20**(2):197-207

[70] Wu Q J, Tymianski M. Targeting NMDA receptors in stroke: New hope in neuroprotection. Molecular Brain. 2018; **11**(1):15

[71] Feng Y, He X, Yang Y, Chao D, Lazarus LH, Xia Y. Current research on opioid receptor function. Current Drug Targets. 2012;**13**:230-246

[72] Vaidya B, Sifat AE, Karamyan VT, Abbruscato TJ. The neuroprotective role of the brain opioid system in stroke injury. Drug Discovery Today. 2018;**23**: 1385-1395

[73] Yang Y, Zhi F, He X, Moore ML, Kang X, Chao D, et al. Delta-opioid receptor activation and microRNA expression of the rat cortex in hypoxia. PLoS One. 2012;7:e51524

[74] Grant Liska M, Crowley MG, Lippert T, Corey S, Borlongan CV. Delta opioid receptor and peptide: A dynamic therapy for stroke and other neurological disorders. In: Handbook of Experimental Pharmacology. Berlin, Heidelberg: Springer; 2017

[75] Staples M, Acosta S, Tajiri N, Pabon M, Kaneko Y, Borlongan CV. Delta opioid receptor and its peptide: A receptor-ligand neuroprotection. International Journal of Molecular Sciences. 2013;**14**(9):17410-17419

[76] Liu H, Chen B, Li S, Yao J. Dosedependent neuroprotection of deltaopioid peptide [D-Ala(2), D-Leu(5)] enkephalin on spinal cord ischemiareperfusion injury by regional perfusion into the abdominal aorta in rabbits. Journal of Vascular Surgery. 2016;**63**(4): 1074-1081

[77] Lee JY, Liska MG, Crowley M, Xu K, Acosta SA, Borlongan CV, et al.
Multifaceted effects of delta opioid receptors and DADLE in diseases of the nervous system. Current Drug Discovery Technologies. 2018;15(2): 94-108

[78] Crowley MG, Liska MG, Lippert T, Corey S, Borlongan CV. Utilizing delta opioid receptors and peptides for cytoprotection: Implications in stroke and other neurological disorders. CNS & Neurological Disorders Drug Targets. 2017;**16**(4):414-424

[79] Moghal ETB, Venkatesh K, Sen D. The delta opioid peptide D-Alanine 2, Leucine 5 Enkephaline (DADLE)induces neuroprotection through crosstalk between the UPR and pro-survival MAPK-NGF-Bcl2 signaling pathways via modulation of several micro-RNAs in SH-SY5Y cells subjected to ER stress. Cell Biology International. 2018;**42**(5): 543-569

[80] Lv MR, Li B, Wang MG, Meng FG, Yu JJ, Guo F, et al. Activation of the PI3K-Akt pathway promotes neuroprotection of the delta-opioid receptor agonist against cerebral ischemia-reperfusion injury in rat models. Biomedicine & Pharmacotherapy. 2017;93:230-237

[81] Yang L, Shah K, Wang H, Karamyan VT, Abbruscato TJ. Characterization of neuroprotective effects of biphalin, an opioid receptor agonist, in a model of focal brain ischemia. The Journal of Pharmacology and Experimental Therapeutics. 2011;**339**(2):499-508

[82] Kawalec M, Kowalczyk JE,Beresewicz M, Lipkowski AW, ZablockaB. Neuroprotective potential of

biphalin, multireceptor opioid peptide, against excitotoxic injury in hippocampal organotypic culture. Neurochemical Research. 2011;**36**(11): 2091-2095

[83] Remesic M, Macedonio G, Mollica A, Porreca F, Hruby V, Lee YS. Cyclic biphalin analogues with a novel linker lead to potent agonist activities at mu, delta, and kappa opioid receptors. Bioorganic & Medicinal Chemistry. 2018;**26**(12):3664-3667

[84] Popiolek-Barczyk K, Piotrowska A, Makuch W, Mika J. Biphalin, a dimeric enkephalin, alleviates LPS-induced activation in rat primary microglial cultures in opioid receptor-dependent and receptor-independent manners. Neural Plasticity. 2017;**2017**:3829472

[85] Yang L, Islam MR, Karamyan VT, Abbruscato TJ. In vitro and in vivo efficacy of a potent opioid receptor agonist, biphalin, compared to subtypeselective opioid receptor agonists for stroke treatment. Brain Research. 1609; 2015:1-11

[86] Islam MR, Yang L, Lee YS, Hruby VJ, Karamyan VT, Abbruscato TJ. Enkephalin-fentanyl multifunctional opioids as potential neuroprotectants for ischemic stroke treatment. Current Pharmaceutical Design. 2016;**22**: 6459-6468

# Chapter 3

# The Role and Development of the Antagonist of Adenosine A<sub>2A</sub> in Parkinson's Disease

Widya Dwi Aryati, Nabilah Nurtika Salamah, Rezi Riadhi Syahdi and Arry Yanuar

## Abstract

Adenosine is a neuromodulator that regulates the body's response to dopamine and another neurotransmitter in the brain that is responsible for motoric, emotion, learning, and memory function. Adenosine is a G-protein-coupled receptor and has four subtypes, which are  $A_1$ ,  $A_{2A}$ ,  $A_{2B}$ , and  $A_3$ . Adenosine  $A_{2A}$  is located in the striatum of the brain. Antagonist interferes with GABA releasing, modulates acetylcholine and releases dopamine, and also facilitates dopamine receptor's signaling. Therefore, it can reduce motoric symptoms in Parkinson's disease. Adenosine  $A_{2A}$ antagonist is also believed to have neuroprotective effects. Several compounds have been reported and have undergone clinical test as selective adenosine  $A_{2A}$  antagonists, including istradefylline, preladenant, tozadenant, vipadenant, ST-1535, and SYN-115. Nonselective adenosine  $A_{2A}$  antagonists from natural compounds are caffeine and theophylline.

**Keywords:** adenosine  $A_{2A}$ , selective adenosine  $A_{2A}$  antagonists, Parkinson's disease, neuroprotective, natural compounds

# 1. Introduction

Adenosine is a neuromodulator that coordinates responses to dopamine and other neurotransmitters in areas of the brain responsible for motor function, mood, learning, and memory [1]. Adenosine consists of four receptor subtypes:  $A_1$ ,  $A_{2A}$ ,  $A_{2B}$ , and  $A_3$  belonging to the superfamily of G-protein-coupled receptor. Adenosine  $A_1$  and  $A_3$  receptors are coupled to inhibitory G proteins, while  $A_{2A}$  and  $A_{2B}$  receptors are coupled to stimulatory G proteins [2].

Adenosine  $A_1$  receptor can be found in adipose tissue, heart muscle, and inflammatory cells. The receptor mostly expressed by the central nervous system such as neocortex, cerebellum, hippocampus, and dorsal horn of the spinal cord [3]. The pre- and postsynaptic nerve terminals, mast cells, airway smooth muscle, and circulating leukocytes are the places where adenosine  $A_2$  receptor can be found. As the more widely dispersed receptor, adenosine  $A_2$  is divided into two receptors on the basis of high- and low-affinity for adenosine,  $A_{2A}$  and  $A_{2B}$  [4]. Striatal neurons are where the adenosine  $A_{2A}$  are highly enriched; however its lower levels can also be found in glial cells and neurons outside the striatum [5]. The adenosine  $A_{2B}$  receptors are highly expressed in the gastrointestinal tract, bladder, lung, and on mast cells. The most widely dispersed receptor is the  $A_3$  receptor which can be found in the kidney, testis, lung, mast cells, eosinophils, neutrophils, heart, and the brain cortex [4].

Adenosine  $A_{2A}$  receptors are found to be concentrates in GABAergic mediumsized spiny neurons in the dopamine-rich regions of the brain. The protein translated in the adenosine  $A_{2A}$  is carried by many other tissues such as blood vessels, endothelial, lymphoid cells, smooth muscle cells, and several neurons in sympathetic and parasympathetic systems [6]. Therefore, the dispersion of adenosine  $A_{2A}$  is not limited to the medium spiny neurons in the basal ganglia. It stimulates the modulation of cAMP production and increases the level of adenylyl cyclase. This receptor is essential in giving the medium of vasodilation of coronary arteries which then supports the combination of new blood vessels and giving protection for tissues from indirect inflammatory damage [7]. The role of the  $A_{2A}$  in the brain includes influencing the activity within the indirect pathway of the basal ganglia. The  $A_{2A}$  has complicated actions because it colocalizes and is physically combined with other unrelated G-protein-coupled receptors. Therefore, it can form heterodimers such as dopamine  $D_2/A_{2A}$ , and  $D_3/A_{2A}$ , cannabinoid  $CB_1/A_{2A}$ , and glutamate mGluR5/ $A_{2A}$ , as well as  $CB_1/A_{2A}/D_2$  heterotrimers [7].

The pathways which give signals used by the  $A_{2A}$  receptor depend on the location of the cell and tissue, the specific G protein which couples it, and the signaling in the cell. The brain also carries the  $A_{2A}$  receptor in which it plays an important role in regulating the glutamate and releasing the dopamine [8]. In the striatopallidal neurons, dopamine  $D_2$  receptors are colocalized with adenosine  $A_{2A}$  receptors. Adenosine  $A_{2A}$  receptor activity that mediates stimulation and  $D_2$  receptors that mediate inhibition in the striatopallidal pathway are balanced [9]. The adenosine  $A_{2A}$ 

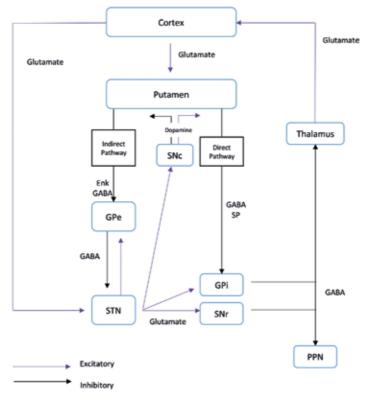


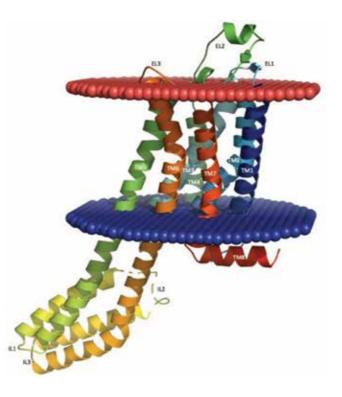
Figure 1. Basal ganglia circuitry in normal conditions.

# The Role and Development of the Antagonist of Adenosine $A_{2A}$ in Parkinson's Disease DOI: http://dx.doi.org/10.5772/intechopen.84272

likely affects motor activity by acting at different levels of the basal ganglia network. The basal ganglia comprise the striatum (putamen), the globus pallidus externa (GPe), the globus pallidus interna (GPi), substantia nigra pars compacta (SNc), substantia nigra reticulata (SNr), and the subthalamic nucleus (STN). The striatum is represented by medium-sized spiny projection neurons (MSNs), accounting for almost 95% of striatal neurons and using γ-aminobutyric acid (GABA) as neurotransmitter. The GABAergic spiny neurons give rise to the two main striatal efferent circuits: the striatonigral and the striatopallidal pathway. The neurons of the striatonigral (direct) pathway contain the neuropeptide substance P and dynorphin and mainly express D1 receptors; this pathway directly projects from the striatum to the GPi/SNr. The neurons of the striatopallidal (indirect) pathway containing the neuropeptide, enkephalin (ENK), predominantly express D<sub>2</sub> receptors; this circuit connects the striatum with the GPi/SNr via synaptic connections in the GPe and STN in Figure 1. Dopamine modulates motor coordination and fine movements by facilitating the action of the direct pathway on stimulatory D<sub>1</sub> receptors and by inhibiting indirect pathway function acting on inhibitory D<sub>2</sub> receptors [10].

The adenosine  $A_{2A}$  receptor has agonists and antagonists of which the roles are potentiating and inhibiting, respectively. The  $D_2$  receptor agonist has effects on motor activity, the releasing of neurotransmitter, and the expression of striatal of c-Fos, a factor of transcription which is used as neuronal activity's indirect marker [11]. The adenosine  $A_{2A}$  receptor has a key role in regulating the striatal dopaminergic neurotransmission which produces substances that are valuable to treat neurological disorders that are relevant with dopaminergic dysfunction.

The topology of G-protein-coupled receptor is displayed in the structure of the adenosine  $A_{2A}$  receptor. These receptors have a central core which consists of seven



#### Figure 2.

Crystal structure of the adenosine  $A_{2A}$  receptor (4EIY) shown in the membrane structure. The extracellular and intracellular parts of the membrane are shown in red and blue beads, respectively. The disorder residues of intracellular loop (IL2) are modeled in dashed line.

transmembrane helices (7TM). Each of the TM is mainly  $\alpha$ -helical and consists of 20–27 amino acids. Three intracellular (IL1, IL2, and IL3) and three extracellular (EL1, EL2, and EL3) loops connect each of the TM domain. A short helix TM8 runs parallel to the cytoplasmic surface of the membrane. The adenosine A<sub>2A</sub> receptor has differences in length and N-terminal extracellular domain function, their domain of C-terminal intracellular, and their loops of intracellular/extracellular. These differences are shown in **Figure 2**.

## 2. The role of adenosine A<sub>2A</sub> in Parkinson's disease

Parkinson's disease (PD) is a chronic neurodegenerative disorder in the brain, marked by motoric symptoms [12]. The motoric symptoms in PD are resting tremor, rigidity, bradykinesia, and postural disorder. Besides motoric symptoms, PD also has non-motoric symptoms such as depression, hallucination, sleeping disorder, and decreasing cognitive and sensory functions. The main pathological characteristic of PD is the loss of dopaminergic neurons in *substantia nigra pars compacta*, a region in the brain that controls all the body movement and forms the dopamine. The development of PD also includes the formation of Lewy body, a deposit of cytoplasmic, eosinophilic neuronal inclusions, composed of the presynaptic protein α-synuclein [13, 14].

The current therapy of PD is targeted at dopamine replacement, thereby decreasing the motor symptoms. It includes precursor of dopamine (levodopa), dopamine agonists [15, 16] monoamine oxidase type B (MAO-B) inhibitors [17], and catechol-O-methyltransferase (COMT) inhibitors [17, 18]. These agents produce undesirable side effects such as on-off effects, hallucinations, and dyskinesia. These effects get more severe as the treatment continued. The efficacy of these agents is also decreasing as the disease progressed [19].

Because of the undesirable side effects of dopamine replacement therapy, the non-dopaminergic therapy is continuously being explored. One of the approaches is selective adenosine  $A_{2A}$  antagonist [20, 21]. Adenosine  $A_{2A}$  receptors are found mainly in the striatum of rat [22, 23], which has similar distribution with the human brain [24, 25]. In the striatum, adenosine  $A_{2A}$  receptors are colocalized with dopamine  $D_2$  receptors. These two receptors have opposite effect on motoric function [26]. The activation of adenosine  $A_{2A}$  receptors will inhibit the signaling of dopamine  $D_2$  receptors, and conversely, the inhibition of signaling of adenosine  $A_{2A}$  receptors will increase the activation of dopamine  $D_2$  receptors, therefore facilitating dopamine  $D_2$ -mediated responses [11]. The inhibition of adenosine  $A_{2A}$  receptors showed motoric improvement in animal models of PD [27–30]. This also has desirable effect on long-term levodopa treatment such as decreasing the dyskinesia and increasing the therapeutic effect on levodopa [31, 32].

## 3. Adenosine A<sub>2A</sub> receptor antagonist as a neuroprotective

For years, adenosine-dopamine interactions have been investigated in order to observe their relevance for treatment of central nervous system (CNS) disorders [33]. It is assumed that adenosine  $A_1$  receptors ( $A_1Rs$ ) play an important role in neuroprotection as their activation at the onset of neuronal injury has shown to reduce brain damage in adult animal model. Vice versa, their blockade aggravates the damage. In other hand, adenosine  $A_2$  receptors ( $A_{2A}Rs$ ) are shown to be upregulated in harmful brain conditions, and their blockade shows brain neuroprotection in studied animals [34]. The blockade of  $A_{2A}Rs$  alleviates the long-term

# The Role and Development of the Antagonist of Adenosine $A_{2A}$ in Parkinson's Disease DOI: http://dx.doi.org/10.5772/intechopen.84272

burden of brain disorders in different neurodegenerative conditions, namely, ischemia, epilepsy, and Parkinson's and Alzheimer's disease, through its control on neuronal cell death [35].

A<sub>2A</sub>Rs have been shown to be viable in serving as alternative non-dopaminergic strategy of Parkinson's disease treatment because of their limited distribution in the striatum and the intense interaction between adenosine and dopamine receptors in the brain. A<sub>2A</sub>Rs antagonists were shown to improve motor function in different animal models (primates and rodents), alone or co-administered with dopamino-mimetic drugs, levodopa, or dopamine agonists [35]. Based on rigorous preclinical animal studies, istradefylline (KW6002) has shown its promising ability to increase motor activity in PD of the advanced stage in clinical phase IIB trial [36]. It became the first therapeutic agent developed to target A<sub>2A</sub>Rs, and other similar compounds will be available in near future [37].

The recent meta-analysis (n = 6) suggested that 20 mg of istradefylline improves unified Parkinson's disease ranking scale (UPDRS) III. Meanwhile at 40 mg per day, istradefylline could alleviate off time and motor symptoms derived from Parkinson's disease [38]. Phase 3 study (613 randomized patients), done by Isaacson et al. concluded that greater reduction from baseline in total hours off time/day were shown at all-time points for istradefylline 20 and 40 mg/day, compared to placebo. However, future development is needed as the study has not yet reached statistical significance [39].

In the case of Parkinson's disease, microglia has been suggested to be the most likely cell type to be targeted by  $A_{2A}Rs$  antagonists [40]. In vitro and in vivo studies showed that local neuroinflammation make glial cells (especially microglial cells) particularly sensitive to  $A_{2A}R$  modulation [41]. Previous research done by Gao and Phillis is the first study to demonstrate nonselective  $A_{2A}R$  antagonist action in reducing cerebral ischemic injury in the gerbil, following global forebrain ischemia [42]. After that, many studies have reported the neuroprotective of  $A_{2A}R$  antagonists in different models of ischemia [43].

Alzheimer's disease (AD) is a chronic neurodegenerative disorder that is indicated by the progressive loss of memory and other cognitive functions, leading to dementia [44, 45]. Adenosine can control and integrate cognition and memory [46]. Both A<sub>1</sub>Rs and A<sub>2A</sub>Rs, mainly located in synapses, control the release of neurotransmitters which are involved in memory or other cognitive processes [34, 47]. Methylxanthine was discovered to act as nonselective adenosine receptors antagonist. Caffeine, the most famous methylxanthine found in common beverages, is the most widely consumed psychoactive drug. Maia and de Mendonca presented the first epidemiological data showing that the incidence of AD is inversely proportioned with coffee consumption [48]. Several other studies also show this inverse relationship [49–51]. Animal models also shown that caffeine intake may be beneficial for AD. In a study, a 6-month period of 0.3 g/L caffeine intake alleviated the cognitive deficits found in AD transgenic mice (APPsw). Furthermore, these mice culture neurons showed the reduced production of  $A\beta_{1-40}$  and  $A\beta_{1-42}$  peptides [52].  $A_{2A}$ Rs antagonists and/or caffeine prophylactic and long-term neuroprotective process are suggested to be based on inhibition of reactive oxygen species activity, tau pathology, and A $\beta$  production by neuronal cells [53].

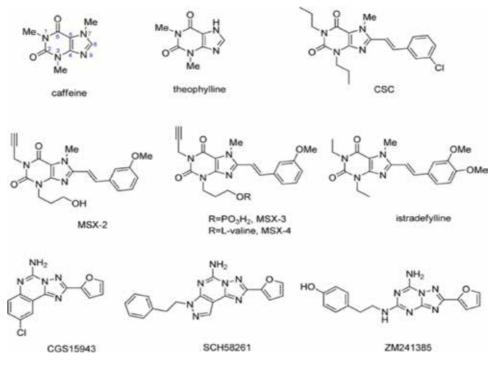
 $A_{2A}Rs$  antagonist may also serve as antidepressants, as observed in animal model of antidepressants screening test done by El-Yacoubi et al. [54, 55]. In both tests,  $A_{2A}Rs$  antagonists prolong escape-directed behavior. Additionally, potential role as antidepressants was also observed in attenuated behavioral despairs displayed in both tests [55]. The relation between adenosine and depression in preclinical models was obtained from the genetic manipulation model of  $A_{2A}Rs$ . Genetic depletion of  $A_{2A}Rs$  resulted in antidepressant-like phenotype in animal models [55]. The A<sub>2A</sub>Rs blockade also relieves stress-induced early hippocampal modifications [56]. However, the effect of adenosine neuromodulation system in depression is complex, as it has the ability to modulate several other neurotransmission systems [35].

As addressed in previous paragraphs,  $A_{2A}R$  emerges as potential target candidate in various disorders. This is majorly caused by its unique interaction with D2 receptors, a major psychoactive drug target. Important roles of  $A_{2A}R$  were also observed in its robust neuroprotective activity, in which it mainly acts in the normalization of glutaminergic synapses, the control of mitochondria-induced apoptosis, and the control of neuroinflammation [35].

## 4. Current sources of the adenosine A2A antagonist

The treatment of PD currently focuses on symptom management with dopaminergic therapy, such as dopamine precursor L-3,4-dihydroxyphenylalanine (L-DOPA) (in combination with peripheral decarboxylase inhibitors) and dopamine agonists [57]. Although L-DOPA is beneficial in patients with PD, with time, the span of the effect is shortened), the response becomes less probable, and involuntary muscle movements or, in a severe situation, dystonia can emerge [57]. These problems highlight the urgent medical need for an alternative mode of therapeutic intervention that can relieve the symptoms of the disorder while also allowing a decrease in the occurrence of side effects.

Among the non-dopaminergic therapies investigated for the treatment of PD, the adenosine  $A_{2A}$  receptor antagonists show very convincingly for two main reasons: their selective and restricted localization in the basal ganglia circuitry and their interaction with dopaminergic receptors. In another word, inhibition of the interaction of adenosine with the  $A_{2A}$  receptor may provide a potential treatment for PD.



**Figure 3.** Adenosine  $A_{2A}$  inhibitors.

# The Role and Development of the Antagonist of Adenosine $A_{2A}$ in Parkinson's Disease DOI: http://dx.doi.org/10.5772/intechopen.84272

Many highly selective  $A_{2A}$  antagonists, both xanthine and non-xanthine derivatives, have been created, and some of them are being investigated as treatment for subjects with PD in various stage of clinical trials (**Figure 3**) [7, 19, 58–61]. Caffeine as a xanthine derivate is developed as a lead compound for the design of antagonist of adenosine  $A_{2A}$  receptor [62]. Experimental model using 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-induced parkinsonism is known to be an evidence that caffeine have a protective effect in Parkinson's disease [36, 63]. Some  $A_{2A}$  antagonists have progressed to clinical trials by various pharmaceutical companies including istradefylline [59], PBS-509, ST1535 and its metabolite ST4206, tozadenant, V81444, preladenant, and vipadenant [64]. Several studies of novel series of 2-aminoimidazo[4,5-b]pyridine-derivatives [65], arylindenopyrimidine [66], and bicyclic aminoquinazoline derivatives [67] as adenosine  $A_{2A}$  antagonists are reported.

Various computational methods were used to study neuroprotective effect from adenosine  $A_{2A}$  antagonists such as pharmacophore model [68], QSAR, molecular docking [69–71], and molecular dynamics [72, 73]. Orally bioavailable adenosine  $A_{2A}$  receptor antagonists have been studied for its QSAR and pharmacokinetics properties [74].

The study of structure-kinetics relationship (SKR) is done as a complement to a SAR analysis at the adenosine  $A_{2A}$  receptor. The series of 24 triazolotriazine derivatives showing a similar binding kinetics to the putative antagonist ZM241385 (4-(2-((7-amino-2-(furan-2-yl)-[1,2,4]triazolo[1,5-a][1,3,5]triazin-5-yl)amino) ethyl)phenol) revealed minor affinity changes, although they varied substantially in their dissociation rates from the receptor [75].

# 5. Future direction of drug discovery of Parkinson's disease

Various studies have been conducted in the discovery of Parkinson's drugs against the target  $A_{2A}$  receptors. The discovery of drugs assisted by computers has accelerated in obtaining lead compounds. Apparently, this method takes a lot of consideration before entering the preclinical and clinical phases. It is because this computational method is more able to describe the answer in preparing the next design. This method can also make various predictions of activities that are difficult to do in the absence of chemical compounds before they are synthesized. In silico prediction of various pharmacokinetic parameters and toxicity can also be done faster. All of these things can provide a better picture of getting a cure for Parkinson's disease.

## 6. Conclusions

 $A_{2A}$  receptors emerge as potential target candidate in various disorders, caused by its unique interaction with D2 receptors, a major psychoactive drug target. Various studies have been conducted in the discovery of Parkinson's drugs against the target  $A_{2A}$  receptors. In silico study brings a new approach of study with  $A_{2A}$ receptors.

## Acknowledgements

This work was supported by *Hibah Publikasi Internasional Terindeks Untuk Tugas Akhir Mahasiswa UI* (PITTA) 2018 by Universitas Indonesia. Neuroprotection

# **Conflict of interest**

The authors declare that they have no conflict of interest or involvement with any organization of affiliation.

# **Author details**

Widya Dwi Aryati, Nabilah Nurtika Salamah, Rezi Riadhi Syahdi and Arry Yanuar<sup>\*</sup> Faculty of Pharmacy, Universitas Indonesia, Depok, Indonesia

\*Address all correspondence to: arry.yanuar@ui.ac.id

# IntechOpen

© 2019 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/ by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. The Role and Development of the Antagonist of Adenosine  $A_{2A}$  in Parkinson's Disease DOI: http://dx.doi.org/10.5772/intechopen.84272

# References

[1] Latini S, Pedata F. Adenosine in the central nervous system: Release mechanisms and extracellular concentrations. Journal of Neurochemistry. 2008;**79**:463-484

[2] Stiles GL. Adenosine receptors. Journal of Biological Chemistry. 1992;**267**:6451-6454

[3] Townsend-Nicholson A, Baker
E, Schofield PR, Sutherland
GR. Localization of the adenosine A<sub>1</sub>
receptor subtype gene (ADORA1)
to chromosome 1q32.1. Genomics.
1995;26:423-425

[4] Livingston M, Heaney LG, Ennis M. Adenosine, inflammation and asthma? A review. Inflammation Research. 2004;**53**:171-178

[5] Boison D, Singer P, Shen H-Y, Feldon J, Yee BK. Adenosine hypothesis of schizophrenia—Opportunities for pharmacotherapy. Neuropharmacology. 2012;**62**:1527-1543

[6] Fredholm BB, Cunha RA, Svenningsson P. Pharmacology of adenosine A<sub>2A</sub> receptors and therapeutic applications. Current Topics in Medicinal Chemistry. 2003;**3**:413-426

[7] de Lera Ruiz M, Lim Y-H, Zheng
 J. Adenosine A<sub>2A</sub> receptor as a drug discovery target. Journal of Medicinal Chemistry. 2014;57:3623-3650

[8] Kull B, Svenningsson P, Fredholm BB. Adenosine A<sub>2A</sub> receptors are colocalized with and activate golf in rat striatum. Molecular Pharmacology. 2000;**58**:771-777

[9] Torvinen M et al. Adenosine  $A_{2A}$ receptor and dopamine  $D_3$  receptor interactions: evidence of functional  $A_{2A}/D_3$  heteromeric complexes. Molecular Pharmacology. 2005;**67**:400-407 [10] Schapira AHV. Present and future drug treatment for Parkinson's disease. Journal of Neurology, Neurosurgery, and Psychiatry. 2005;**76**:1472-1478

[11] Pollack AE, Fink JS. Adenosine antagonists potentiate D2 dopaminedependent activation of Fos in the striatopallidal pathway. Neuroscience. 1995;**68**:721-728

[12] Lozano AM, Lang AE, Hutchison
WD, Dostrovsky JO. New developments in understanding the etiology of
Parkinson's disease and in its treatment.
Current Opinion in Neurobiology.
1998;8:783-790

[13] Savitt JM. Diagnosis and treatment of Parkinson disease: Molecules to medicine. Journal of Clinical Investigation. 2006;**116**:1744-1754

[14] Dauer W, Przedborski S. Parkinson's disease. Neuron. 2003;**39**:889-909

[15] Antonini A, Tolosa E, Mizuno Y, Yamamoto M, Poewe WH. A reassessment of risks and benefits of dopamine agonists in Parkinson's disease. Lancet Neurology. 2009;**8**:929-937

[16] Yamamoto M, SchapiraAH. Dopamine agonists inParkinson's disease. Expert Review ofNeurotherapeutics. 2008;8:671-677

[17] Olanow CW, Stocchi F. COMT inhibitors in Parkinson's disease: Can they prevent and/or reverse levodopainduced motor complications? Neurology. 2004;**62**:S72-S81

[18] Gordin A, Brooks DJ. Clinical pharmacology and therapeutic use of COMT inhibition in Parkinson's disease. Journal of Neurology. 2007;**254**:IV37-IV48

[19] Shook BC, Jackson PF. Adenosine A<sub>2A</sub> receptor antagonists and Parkinson's disease. ACS Chemical Neuroscience. 2011;2:555-567

[20] Schwarzschild MA, Agnati L, Fuxe K, Chen J-F, Morelli M. Targeting adenosine  $A_{2A}$  receptors in Parkinson's disease. Trends in Neurosciences. 2006;**29**:647-654

[21] Salamone JD. Facing dyskinesia in Parkinson disease: Nondopaminergic approaches. Drugs Future. 2010;**35**:567

[22] Rosin DL, Robeva A, Woodard RL, Guyenet PG, Linden J. Immunohistochemical localization of adenosine  $A_{2A}$  receptors in the rat central nervous system. Journal of Comparative Neurology. 1998;**401**:163-186

[23] Fredholm BB, Svenningsson
P. Striatal adenosine A<sub>2A</sub> receptors—
Where are they? What do they do?
Trends in Pharmacological Sciences.
1998;19:46-47

[24] Ishiwata K et al. First visualization of adenosine  $A_{2A}$  receptors in the human brain by positron emission tomography with [<sup>11</sup>C]TMSX. Synapse. 2005;55:133-136

[25] Svenningsson P, Hall H, Sedvall G, Fredholm BB. Distribution of adenosine receptors in the postmortem human brain: An extended autoradiographic study. Synapse. 1997;**27**:322-335

[26] Fink JS et al. Molecular cloning of the rat  $A_2$  adenosine receptor: Selective co-expression with  $D_2$  dopamine receptors in rat striatum. Molecular Brain Research. 1992;**14**:186-195

[27] Chen J-F et al. Neuroprotection by caffeine and  $A_{2A}$  adenosine receptor inactivation in a model of Parkinson's disease. Journal of Neuroscience. 2001;**21**:RC143-RC143

[28] Grondin R et al. Antiparkinsonian effect of a new selective adenosine  $A_{2A}$ 

receptor antagonist in MPTP-treated monkeys. Neurology. 1999;**52**:1673-1673

[29] Ongini E et al. Dual actions of A<sub>2A</sub> adenosine receptor antagonists on motor dysfunction and neurodegenerative processes. Drug Development Research. 2001;52:379-386

[30] Ikeda K, Kurokawa M, Aoyama S, Kuwana Y. Neuroprotection by adenosine A<sub>2A</sub> receptor blockade in experimental models of Parkinson's disease. Journal of Neurochemistry. 2002;80:262-270

[31] Kanda T et al. Combined use of the adenosine A<sub>2A</sub> antagonist KW-6002 with I-DOPA or with selective D1 or D2 dopamine agonists increases antiparkinsonian activity but not dyskinesia in MPTP-treated monkeys. Experimental Neurology. 2000;**162**:321-327

[32] Hauser RA, Hubble JP, Truong
 DD. Randomized trial of the adenosine
 A<sub>2A</sub> receptor antagonist istradefylline
 in advanced PD. Neurology.
 2003;61:297-303

[33] Fuxe K et al. Adenosine-dopamine interactions in the pathophysiology and treatment of CNS disorders. CNS Neuroscience and Therapeutics. 2010;**16**:e18-e42

[34] Cunha RA. Neuroprotection by adenosine in the brain: From  $A_1$  receptor activation to  $A_{2A}$  receptor blockade. Purinergic Signal. 2005;**1**:111-134

[35] Gomes CV, Kaster MP, Tomé AR, Agostinho PM, Cunha RA. Adenosine receptors and brain diseases: Neuroprotection and neurodegeneration. Biochimica et Biophysica Acta (BBA)— Biomembranes. 2011;**1808**:1380-1399

[36] Kalda A, Yu L, Oztas E, Chen J-F. Novel neuroprotection by caffeine and adenosine A<sub>2A</sub> receptor antagonists The Role and Development of the Antagonist of Adenosine  $A_{2A}$  in Parkinson's Disease DOI: http://dx.doi.org/10.5772/intechopen.84272

in animal models of Parkinson's disease. Journal of the Neurological Sciences. 2006;**248**:9-15

 [37] Yamada K, Kobayashi M, Kanda T. Chapter Fifteen - Involvement of Adenosine A<sub>2A</sub> Receptors in Depression and Anxiety. International Review of Neurobiology. 2014;119:373-393. DOI: 10.1016/B978-0-12-801022-8.00015-5

[38] Sako W, Murakami N, Motohama K, Izumi Y, Kaji R. The effect of istradefylline for Parkinson's disease: A meta-analysis. Scientific Reports. 2017;7:18018

[39] Isaacson S et al. Efficacy and safety of istradefylline in moderate to severe Parkinson's disease: A phase 3, multinational, randomized, doubleblind, placebo-controlled trial (i-step study). Journal of the Neurological Sciences. 2017;**381**:351-352

[40] Franco R, Navarro G. Adenosine  $A_{2A}$  receptor antagonists in neurodegenerative diseases: Huge potential and huge challenges. Frontiers in Psychiatry. 2018;**9**(1-5)

[41] Chen J-F, Pedata F. Modulation of ischemic brain injury and neuroinflammation by adenosine A<sub>2A</sub> receptors. Current Pharmaceutical Design. 2008;**14**:1490-1499

[42] Gao Y, Phillis JW. CGS 15943, An adenosine A2 receptor antagonist, reduces cerebral ischemic injury in the Mongolian gerbil. Life Sciences. 1994;**55**:PL61-PL65

[43] Pedata F et al. Adenosine A<sub>2A</sub>
 receptors modulate acute injury
 and neuroinflammation in brain
 ischemia. Mediators of Inflammation.
 2014;805198:2014

[44] Kalaria RN et al. Alzheimer's disease and vascular dementia in developing countries: Prevalence, management, and risk factors. The Lancet Neurology. 2008;7:812-826 [45] Lesne S. Toxic oligomer species of amyloid- $\beta$  in Alzheimer's disease, a timing issue. Swiss Medical Weekly. 2014;**298**:789-791

[46] Cunha RA, Agostinho PM. Chronic caffeine consumption prevents memory disturbance in different animal models of memory decline. Journal of Alzheimer's Disease. 2010;**20**:S95-S116

[47] Ribeiro JA, Sebastião AM, de Mendonça A. Adenosine receptors in the nervous system: Pathophysiological implications. Progress in Neurobiology. 2002;**68**:377-392

[48] Maia L, de Mendonça A. Does caffeine intake protect from Alzheimer's disease? Europen Journal of Neurology. 2002;**9**:377-382

[49] Ritchie K et al. The neuroprotective effects of caffeine: A prospective population study (the Three City Study). Neurology. 2007;**69**:536-545

[50] Santos C, Costa J, Santos J, Vaz-Carneiro A, Lunet N. Caffeine intake and dementia: systematic review and meta-analysis. Journal of Alzheimer's Disease. 2010;**20**:S187-S204

[51] Smith AP. Caffeine, cognitive failures and health in a non-working community sample. Human Psychopharmacology Clinical and Experimental. 2009;**24**:29-34

[52] Arendash GW et al. Caffeine protects Alzheimer's mice against cognitive impairment and reduces brain β-amyloid production. Neuroscience. 2006;**142**:941-952

[53] Marzagalli R, Castorina A. The seeming paradox of adenosine receptors as targets for the treatment of Alzheimer's disease: Agonists or antagonists? Neural Regeneration Research. 2015;**10**:205

[54] Cantwell R, Cox JL. Psychiatric disorders in pregnancy and the

puerperium. Current Obstetrics & Gynaecology. 2006;**16**:14-20

[55] El Yacoubi M et al. Adenosine A<sub>2A</sub> receptor antagonists are potential antidepressants: evidence based on pharmacology and A<sub>2A</sub> receptor knockout mice. British Journal of Pharmacology. 2001;**134**:68-77

[56] Cunha GMA, Canas PM, Oliveira CR, Cunha RA. Increased density and synapto-protective effect of adenosine  $A_{2A}$  receptors upon subchronic restraint stress. Neuroscience. 2006;**141**:1775-1781

[57] Olanow CW et al. Levodopa in the treatment of Parkinson's disease: Current controversies. Movement Disorders. 2004. DOI: 10.1002/ mds.20243

[58] Armentero MT et al. Past, present and future of A<sub>2A</sub> adenosine receptor antagonists in the therapy of Parkinson's disease. Pharmacology & Therapeutics. 2011. DOI: 10.1016/j. pharmthera.2011.07.004

[59] Jenner P. Istradefylline, a novel adenosine  $A_{2A}$  receptor antagonist, for the treatment of Parkinson's disease. Expert Opinion on Investigational Drugs. 2005. DOI: 10.1517/13543784.14.6.729

[60] Pinna A. Novel investigational adenosine  $A_{2A}$  receptor antagonists for Parkinson's disease. Expert Opinion on Investigational Drugs. 2009. DOI: 10.1517/13543780903241615

[61] Hickey P, Stacy M. Adenosine A<sub>2A</sub> antagonists in Parkinson's disease: What's next? Current Neurology and Neuroscience Reports. 2012;**12**:376-385

[62] Petzer JP, Petzer A. Caffeine as a lead compound for the design of therapeutic agents for the treatment of Parkinson's disease. Current Medicinal Chemistry. 2015. DOI: 10.2174/0929867 322666141215160015 [63] Munoz DG, Fujioka S. Caffeine and Parkinson disease: A possible diagnostic and pathogenic breakthrough. Neurology. 2018. DOI: 10.1212/ WNL.000000000004898

[64] Pinna A. Adenosine A<sub>2A</sub> receptor antagonists in Parkinson's disease: Progress in clinical trials from the newly approved istradefylline to drugs in early development and those already discontinued. CNS Drugs. 2014;28:455-474

[65] McGuinness BF et al. Discovery of
2-aminoimidazopyridine adenosine
A<sub>2A</sub> receptor antagonists. Bioorganic
& Medicinal Chemistry Letters.
2010;20:6845-6849

[66] Atack JR et al. JNJ-40255293, a novel adenosine  $A_{2A}/A_1$  antagonist with efficacy in preclinical models of Parkinson's disease. ACS Chemical Neuroscience. 2014;5:1005-1019

[67] Zhou G et al. Bioorganic & medicinal chemistry letters discovery of aminoquinazoline derivatives as human  $A_{2A}$  adenosine receptor antagonists. Bioorganic & Medicinal Chemistry Letters. 2016;**26**:1348-1354

[68] Khanfar MA, Al-Qtaishat S, Habash M, Taha MO. Discovery of potent adenosine  $A_{2A}$  antagonists as potential anti-Parkinson disease agents. Nonlinear QSAR analyses integrated with pharmacophore modeling. Chemico-Biological Interactions. 2016;**254**:93-101

[69] Anighoro A, Bajorath J. Binding mode similarity measures for ranking of docking poses: A case study on the adenosine  $A_{2A}$  receptor. Journal of Computer-Aided Molecular Design. 2016. DOI: 10.1007/s10822-016-9918-z

[70] Yang X et al. A covalent antagonist for the human adenosine  $A_{2A}$  receptor. Purinergic Signal. 2017. DOI: 10.1007/ s11302-016-9549-9 The Role and Development of the Antagonist of Adenosine  $A_{2A}$  in Parkinson's Disease DOI: http://dx.doi.org/10.5772/intechopen.84272

[71] Jaiteh M et al. Docking screens for dual inhibitors of disparate drug targets for Parkinson's disease. Journal of Medicinal Chemistry. 2018. DOI: 10.1021/acs.jmedchem.8b00204

[72] Sabbadin D et al. Bridging molecular docking to membrane molecular dynamics to investigate
GPCR-ligand recognition: The human
A<sub>2A</sub> adenosine receptor as a key study.
Journal of Chemical Information and Modeling. 2013. DOI: 10.1021/ci400532b

[73] Caliman AD, Swift SE, Wang Y,
Miao Y, McCammon JA. Investigation of the conformational dynamics of the apo A<sub>2A</sub> adenosine receptor. Protein Science.
2015. DOI: 10.1002/pro.2681

[74] Basu S et al. Design, synthesis of novel, potent, selective, orally bioavailable adenosine  $A_{2A}$  receptor antagonists and their biological evaluation. Journal of Medicinal Chemistry. 2017. DOI: 10.1021/acs. jmedchem.6b01584

 [75] Guo D et al. Binding kinetics of ZM241385 derivatives at the human adenosine A<sub>2A</sub> receptor.
 ChemMedChem. 2014. DOI: 10.1002/ cmdc.201300474

# **Chapter 4**

# Adrenergic Receptors as Pharmacological Targets for Neuroinflammation and Neurodegeneration in Parkinson's Disease

Monika Sharma and Patrick M. Flood

# Abstract

Inflammation is a key component of the dopaminergic neurodegeneration seen in progressive Parkinson's disease (PD). The presence of activated glial cells, the participation of innate immune system, increased inflammatory molecules such as cytokines and chemokines, and increased oxidative stress and reactive oxygen species are the main neuroinflammatory characteristics present in progressive PD. Therapeutic targets which suppress pro-inflammatory responses by glial cells (mainly microglia) have been shown to be effective treatments for slowing or eliminating the progressive degeneration of neurons within the substantia nigra. In this chapter, we will detail a specific anti-inflammatory therapy using agonists to  $\beta$ 2-adrenergic receptors that have been shown to be effective treatments for models of dopaminergic neurodegeneration and that have had efficacy in patients with progressive PD. We will also detail the possible molecular mechanisms of action of this therapeutic in stopping or reversing inflammation within the CNS.

**Keywords:** β2-adrenergic receptor, Parkinson's disease, microglia, neuroinflammation

# 1. Introduction

There are a number of neurological disorders that fall under the umbrella of neurodegeneration, with the major ones including Alzheimer's disease (AD), Parkinson's disease (PD), Huntington's disease (HD), amyotrophic lateral sclerosis (ALS), frontotemporal dementia (FTD), spinal cord injury (SCI), and others. Currently, there are no generally effective treatments available to slow down or reverse the debilitating effects of these diseases, and the long-term effects of these diseases are the progressive degeneration and death of neurons. A majority of the neurodegenerative diseases are linked with inflammation in CNS [1], and the presence of activated glial cells, infiltration and activation of adaptive and innate immune cells, increased presence of inflammatory molecules such as cytokines and chemokines, and increased oxidative stress and reactive oxygen species (ROS) are the main neuroinflammatory characteristics present in lesions associated with

#### Neuroprotection

these neurodegenerative disorders. Recent approaches found to be effective in the treatment of Parkinson's disease involve the use of anti-inflammatory agents and cytokines such as agonists to the  $\beta$ 2-adrenergic receptors ( $\beta$ 2-AR) to inhibit neuroinflammation and the progression of dopaminergic neurodegeneration. In this chapter, we will address the current understanding of therapeutic approaches targeting neuroinflammation linked with PD and the use of  $\beta$ 2-AR agonists as an effective treatment for PD.

# 2. Parkinson's disease: a chronic neurodegenerative and neuroinflammatory disease

Parkinson's disease (PD) is a progressive neurodegenerative disorder which leads to impaired motor skills. The major pathological feature of PD is the degeneration of dopaminergic (DA) neurons which project from substantia nigra (SN) to the striatum in the midbrain (nigro-striatal pathway) [2]. Another neuropathological feature of PD is the cytoplasmic inclusion of misfolded  $\alpha$ -synuclein protein in degenerating dopaminergic neurons called Lewy bodies [3]. The primary motor symptoms of PD, such as tremor, rigidity, and bradykinesia, are caused by inadequate formation and neurotransmission of dopamine within the nigro-striatal pathway [4, 5]. Dementia is reported in 28% of PD cases with the prevalence rising to 65% in those aged 85 years and above. Patients with PD also show non-motorrelated symptoms such as olfactory deficits, depression, cognitive deficits, sleep disorders, and autonomic dysfunction [6]. The majority of PD cases are idiopathic Parkinson's, and the disease mechanism that ultimately causes idiopathic PD is largely unknown. In the remainder of the cases of PD, about 10–15% of patients do have a family history and those patients are referred to as having the *familial* form of PD. For these patients, their PD appears to be caused by a mutation in one of a few selected genes (such as SNCA, Parkin, LRRK2, DJ-1, etc.) [7, 8]. Although the etiology of the idiopathic form of the disease remains elusive, there are some risk factors associated with the development of the disease. These risk factors include exposure to environmental toxins, severe cranial trauma, systemic or localized infections, and inherited genetic risk factors. These genetic and nongenetic risk factors have the potential to initiate neurodegeneration and subsequent chronic inflammation in the brain which eventually contributes to the pathophysiology of PD [9]. In addition, several cellular and molecular pathways such as oxidative stress [10], proteosomal dysfunction [11], excitotoxicity [12], and mitochondrial dysfunction [13] have also been identified which contributes to neuronal death.

The presence of activated glial cells, increased inflammatory molecules such as cytokines/chemokines, and increased oxidative stress and ROS are the main neuroinflammatory characteristics present in PD [14]. PD is now not only characterized as loss of DA-neurons and motor impairment, but also recognized to have an inflammatory component which plays a crucial role in the progression of the disease. Several inflammatory mediators such as TNF- $\alpha$ , IL-1 $\beta$ , ROS, and nitric oxide (NO), released from nonneuronal cells exacerbate the disease pathology [3, 15]. It has been suggested that  $\alpha$ -synuclein released from dying neurons also activate the microglia via TLR2 activation [16]. Furthermore, the elevated levels of inflammatory cytokines such as TNF- $\alpha$ , IL-1 $\beta$ , and IL-6 have been reported in serum, cerebrospinal fluid (CSF), and striatum of PD patients [17]. The influx of peripheral macrophages has been reported in brains of patients with PD, but the role of these cells in disease pathology remains to be tested [18]. Additionally, activation and increased number of glial cells and infiltrating peripheral lymphocytes such as cytotoxic CD4+ and CD8+ cells in SN also support the role of adaptive immunity in Adrenergic Receptors as Pharmacological Targets for Neuroinflammation and Neurodegeneration... DOI: http://dx.doi.org/10.5772/intechopen.81343

the etiology of the disease [8]. Overall, these studies and others suggest the contribution of the immune system in the pathophysiology of PD.

## 3. Microglial activation and neuroinflammation in PD

Microglia originate from erythromyeloid progenitors in the yolk sac which migrate and differentiate during development to form the central nervous system (CNS). Fully differentiated microglial cells are also considered to be the resident macrophages of the CNS [19], although some phenotypic and functional differences between microglia and macrophages have been found [20]. Growing evidence suggests that the activation of microglia in CNS plays an important role in the pathogenesis of PD. It is not well understood how microglia activation is either beneficial or detrimental to the neuron or how microglial activity is regulated. It has been found that microglial activation is required for neuronal survival by the removal of toxic substances through innate immunity [21]. On the other hand, it has been found that over-activated microglial cells are detrimental and neurotoxic [22]. Research studies of post-mortem brain tissue from patients with PD and related parkinsonian syndromes suggest the presence of activated microglia around degenerating DA-neurons in the SN [23] and these activated microglia are not only limited to the SN but also present in extended brain areas such as hippocampus, putamen, trans-entorhinal cortex, cingulate cortex, and temporal cortex [24]. Imaging of activated microglia in the striatum could be used as a biomarker for detecting neuroinflammation in neurodegenerative parkinsonian disorders [25]. The resting microglia switches to an activated microglia phenotype in response to pathogen invasion or release of toxic or inflammatory mediators and thereby promotes an inflammatory response [1]. Once activated, microglial cells produce a wide range of inflammatory mediators which serve to initiate an innate immune response or glial cell-propagated inflammation termed as neuroinflammation [26]. Also, the degenerating DA-neurons release many toxic factors that activate microglia and these degenerating neurons are vulnerable to inflammatory insult. Degenerating neurons will co-localize or attract an even larger population of microglia in the SN [27]. Collectively, these activated microglia and damaged neurons form a repetitive and vicious cycle that leads to chronic inflammation and continued extensive DA neurodegeneration over time, leading to the progression of PD [27]. These findings confirm neuroinflammation as a pivotal process in the progression of neurodegenerative disorders and the central role of microglia in this process [22]. Targeting neuroinflammatory pathways within microglia could be a significant step in the development of new therapeutics for neurodegenerative diseases, including PD.

## 4. Therapies targeting neurodegeneration/neuroinflammation in PD

Treatment for PD normally involves medications such as Levodopa to enhance the dopamine levels and deal with movement symptoms [28]. While none of our current treatments are able to stop the disease, medication and surgery can be helpful for managing the symptoms [29]. These treatments work well in patients initially, but they are also associated with unwanted side-effects and reduced efficacy over time [30]. On the other hand, many studies suggest that inflammatory mediators such as TNF, PGE<sub>2</sub>, NO, free radicals, and other immune mediators play role in the pathogenesis of PD and degeneration of dopamine-producing neurons and that targeting these mediators can be an effective treatment for PD. This opens up the potential of using anti-inflammatory drugs as an effective and long-term treatment in PD. These anti-inflammatory drugs can act by arresting the disease onset (primary prevention) or by interrupting or even reversing the disease progression (secondary prevention). Epidemiological and observational studies suggest that the use of anti-inflammatory drugs lower the risk of developing PD [31]. Observations which demonstrated that inflammation in SN plays a role in PD have led many investigators to initially consider the potential use of both steroidal and nonsteroidal anti-inflammatory drugs for the treatment of PD. Steroidal anti-inflammatory drugs (SAIDs), such as dexamethasone, have shown neuroprotective effects in LPS-induced neurotoxicity in the SN in LPS models of PD [32]. Nonsteroidal anti-inflammatory drugs (NSAIDs) have also been used as analgesics and antipyretics to suppress the adverse effects of inflammation [33]. The neuroprotective effects of Ibuprofen have been studied in PD pathogenesis and these studies demonstrate the protective effect on dopaminergic neurons against glutamate toxicity in vitro [34, 35]. Previously, we have established several therapies targeting neuroinflammation and neurodegeneration in an animal model of PD and these therapies include D-morphinan-related compounds [36], anti-inflammatory cytokines such as TGF- $\beta$  (transforming growth factor-beta) [37] and IL-10 [38, 39], IKK (inhibitor of kappa B (IkB) kinase) inhibitors [40], NADPH (nicotinamide adenine dinucleotide phosphate) oxidase inhibitors [41], and  $\beta$ 2-AR (beta 2adrenergic receptor) agonists [42, 43].

We have conducted a number of experiments using different classes of antiinflammatory compounds to determine their efficacy in preventing dopaminergic neurotoxicity by activated microglial cells both in vitro and in vivo. First, it was found that morphinan compounds and their stereoisomers (L-morphine and its D stereo enantiomers) can inhibit microglial activation and LPS- or MPP<sup>+</sup>-induced neurotoxicity in rat primary mesencephalic cultures. We and others observed that several dextrorotatory isomers of morphine compounds, including D-morphine, dextromethorphan, and sinomenine, showed neuroprotective effects against LPS and MPP+ (1-methyl-4-phenylpyridinium) which were mediated through the inhibition of microglial PHOX activity [36, 44, 45]. Furthermore, these studies also suggest that these morphinan compounds bind to the catalytic subunit of PHOX, inhibit its activity, and reduce the production of superoxide and other pro-inflammatory cytokines [44]. In another set of studies using a different anti-inflammatory approach, a specific inhibitor of IKK- $\beta$  (IkappaB kinase-beta) protects dopaminergic neurons against LPS-induced neurotoxicity both in vitro and *in vivo* through inhibition of NF-κB activation, resulting in the decreased production of ROS and inflammatory cytokines [40]. We have also developed therapies targeting neuroinflammation in PD models by using anti-inflammatory cytokines such as IL-10 and TGF- $\beta$ 1, and found that treatment with IL-10 on rat mesencephalic neuron-glia culture protects against LPS-induced neurotoxicity via suppression of pro-inflammatory mediators and superoxide production [38]. Similarly, the neuroprotective effect of TGF $\beta$ 1 is primarily due to its ability to inhibit ERK phosphorylation, the serine phosphorylation on p47<sup>phox</sup>, and the production of ROS from microglia during activation by LPS [37].

## 5. Adrenergic receptors

One of the most potent and successful therapeutic treatments for inflammationmediated dopaminergic neurotoxicity is the use of long-acting agonists to the  $\beta$ 2-AR. Adrenergic receptors (AR) are seven-transmembrane proteins that serve as adrenoreceptors for catecholamines such as norepinephrine and epinephrine on multiple cell types, and cells within the CNS that express AR include neurons, immune cells, and Adrenergic Receptors as Pharmacological Targets for Neuroinflammation and Neurodegeneration... DOI: http://dx.doi.org/10.5772/intechopen.81343

Receptor Type	Tissue Distribution	Mechanism of Action	Agonist Potency	Physiological Effects	Agonist	Antagonist
α1	Vascular Smooth Muscles, Visceral smooth Muscles	Gq-protein coupled activates Phospholipase C, IP3+DAG	Epi ≥ NE >> Iso	Smooth muscle contractions, Gluconeogenesis, Vasoconstriction	Norepinephrine, Phenylephrine, Methoxamine	Doxazosin, Phentolamine, Prazosin
α2	Pre-synaptic terminals, pancreas, platelets, Ciliary epithelium, Salivary Glands	Gi-protein coupled inhibits Adenyl cyclase	Epi ≥ NE ≫lso	Inhibits release of Neurotransmitter	Clonidine, Monoxidine	Yohimbine, Idazoxan, Tolazoline
β1	Heart, Kidney, some pre- synaptic terminals	Gs-protein coupled activates Adenyl cyclase +PKA	lso > Epi ≥ NE	Increase heart rate and Renin secretion	Isoproterenol, Norepinephrine, Dobutamine	Propranolol, Metoproiol, Atenoiol
β <b>2</b>	Visceral smooth muscles, Bronchioles, Liver, Skeletal Muscles	Gs-protein coupled activates Adenyl cyclase +PKA, Ca- channels	lso > Epi >> NE	Vasodilation, Bronchodilation, Inhibits insulin secretion	Isoproterenol, Salbutamol, Salmeterol, Albuterol, Formoterol, Terbutaline, Levalbuterol	Propranolol, Kl- 118,551, Nadolol, Butoxamine
β3	Adipose Tissue	Gs-protein coupled activates Adenyl cyclase +PKA	lso = NE > Epi	Increase lipolysis	Isoproterenol, Amibegron, Solabegron	SR59230A

NE: Norepinephrine, Epi: Epinephrine and Iso: Isoproterenol

#### Table 1.

Characteristics of adrenergic receptors.

astrocytes. Pharmacological classification of the adrenergic receptor was first introduced in 1948 and broadly classified as  $\alpha$  and  $\beta$  adrenergic receptors [46] by Ahlquist. The classification was based on the order of potency and specificity of natural and synthetic agonist and blocking agents. The  $\alpha$ -AR response corresponds to mainly excitatory response, while β-AR responses were correlated mainly with the inhibitory response. The  $\alpha$ -AR response showed the order of potency: norepinephrine > epinephrine > isoproterenol and  $\beta$ -AR-mediated response exhibited order of potency: isoproterenol > epinephrine > norepinephrine [47, 48]. After the discovery of new drugs which have a high affinity to adrenergic receptors, these receptors were sub-classified. α-AR were subdivided into  $\alpha 1$  and  $\alpha 2$  adrenergic receptors [49]. Further studies subdivided  $\beta$ -AR into  $\beta 1$  and  $\beta 2$  which are normally present on immune cells, cardiac muscles, and airway smooth muscles, respectively [50]. A third  $\beta$ -AR, now called as  $\beta$ 3-AR was identified on adipose tissues [51]. Tissue distribution, physiological effects, mechanism of action, and the major agonists/antagonists of ARs are summarized in Table 1. Pharmacological compounds that serve as short, long, and ultra-long-acting agonists for these receptors have now been developed, and they are normally thought to stimulate adrenergic receptors by four different mechanisms: (1) by direct receptor binding, the most common mechanism where drugs activate peripheral adrenergic receptors via direct binding to receptor and mimic the actions of endogenous agonists (NE, epinephrine), (2) by ameliorating NE release, where drugs act on sympathetic nerve terminals and results into NE release, (3) by inhibition of NE reuptake, where these drugs can cause NE to accumulate within synaptic gaps at sympathetic nerve terminals, (4) by blockade of NE inactivation where drugs inhibit the activity of monoamine oxidase (MAO) which inhibits the activity of monoamines such as NE and dopamine [52].

# 6. General properties of β2-adrenergic receptors: a G-protein-coupled receptor

## 6.1 Structure

The  $\beta$ 2-ARs belong to a diverse superfamily of human cell surface seven transmembrane receptors for hormones and neurotransmitters called G-protein-coupled receptors (GPCRs). GPCRs are divided into six classes on basis of sequence homology: class A (Rhodopsin-like), class B (Secretin receptor family), class C (Metabotropic glutamate), class D (Fungal mating pheromone receptor), class E (Cyclic AMP receptor), and class F (Frizzled/smoothened) [53]. GPCRs are one of the most extensively studied proteins for the development of pharmaceutical drugs and target for approximately 50% of the marketed pharmaceutical drugs [54]. The adrenergic receptor family belongs to the rhodopsin-like subfamily, the largest class of the GPCR. The  $\beta$ 2-AR is an intron-less gene is present on the long arm of chromosome 5 (5q31) and encodes for 413 amino acid polypeptide of 46kD [55]. Similar to all GPCRs,  $\beta$ 2-AR is composed of seven transmembrane spanning  $\alpha$ -helices with an intracellular C-terminus and an extracellular N-terminus. The  $\beta$ 2-AR was the first GPCR to be cloned [56] and the first GPCR structure to be solved [57]. The  $\beta$ 2-AR has been studied extensively and also serves as a model system for investigating the regulation and signal transduction of GPCRs. The study of the 3D protein structure of this family of GPCRs took a giant leap forward when rhodopsin was first crystallized in 2000 and this crystalline structure has been used as an important template for modeling other GPCRs in this family [58]. The crystalline structure of human  $\beta$ 2-AR was not solved until 2007, when a nonactive structure of β2-AR was identified [57]. Post-translational modifications such as glycosylation, pamitoylation, disulfide bond formation, and phosphorylation have now been found to affect receptor functions. Interestingly,  $\beta$ 2-AR is glycosylated at amino acid 6, 15, and 187 which is important for the trafficking of the  $\beta$ 2-AR from the endoplasmic reticulum to the plasma membrane [59]. Mutation in these sites also results in reduced expression of receptor on the cell membrane, suggesting a role for glycosylation in cell surface expression [60]. Conversely, the cysteine amino acid in the cytoplasmic tail at position 341 is palmitoylated, and is now found to be an important residue for the adequate coupling of the receptor to the  $G_s$ -protein [61]. Finally,  $\beta$ 2-ARs have disulfide bonds which are essential for agonist binding and also for maintaining their tertiary structure [62].

# 6.2 Localization

Adrenergic receptors are widely distributed on human body organs and regulate physiologic functions such as bronchodilation [63], vasodilation, glycogenolysis in the liver, and relaxation of uterine and bladder muscles [64]. The human  $\beta$ 2-AR are widely expressed not only on airway smooth muscles, but also on the wide variety of cells such as epithelial cells, endothelial cells, brain cells, and immune cells including mast cells, macrophages, adaptive immune cells, and eosinophils [65]. The expressions of  $\beta$ 1- and  $\beta$ 2-AR have also been found on microglial cells, suggesting that microglia, the brain's resident immune cell, is predominantly regulated by NE since NE is the predominant catecholamine in the CNS. Conversely, peripheral immune cells such as macrophages and T cells, which also express high levels of  $\beta$ 1 and  $\beta$ -2 AR, are thought to be regulated primarily by epinephrine [66].

## 6.3 β2-AR activation and signaling pathways in inflammation

Activation of adrenergic receptors could result into both pro- and antiinflammatory actions, depending on certain parameters such as the type of cell, duration of ligand exposure to the receptor, and type of the adrenergic receptor [67]. It is the diversity of the  $\beta$ 2-AR that leads to the complexity of signaling mechanisms and to this duality of function. Activation of  $\beta$ 2-AR by receptor agonists initiate intracellular signaling pathways that function either via G-proteins or through  $\beta$ -arrestins. Like other GPCR,  $\beta$ 2-AR can activate either canonical (traditional)

## Adrenergic Receptors as Pharmacological Targets for Neuroinflammation and Neurodegeneration... DOI: http://dx.doi.org/10.5772/intechopen.81343

or noncanonical (nontraditional) signal transduction pathway. In the canonical pathway, similar to a typical GPCR the  $\beta$ 2-AR signals via a heterotrimeric G-protein complex, and when the receptor is coupled to inactive GDP-bound G-protein, it appears to have high affinity to the agonist or ligand. After ligand binding, the transmembrane domains of the receptor undergo conformational change with the exchange of GDP to GTP. Further, this conformational change reduces the affinity of the ligand to its receptor, increasing the possibility of retraction of ligand from the receptor, thereby preventing the over-activation of G-protein. This provides evidence that  $\beta$ 2-AR appear to oscillate between an active and inactive form under normal conditions. After the exchange of GDP to GTP, the  $G_{\alpha}$ -subunit dissociates from  $G_{\beta\gamma}$ -subunit which remains associated with plasma membrane and the  $G_{\alpha}$ -subunit activates effector proteins. The downstream signaling of this process normally results in the production of intracellular second messengers which further activates the cAMP-PKA-mediated intracellular signaling pathway. The activated  $\beta$ 2-AR binds with the  $\alpha$ -subunit of the G-protein together with a guanosine triphosphate (GTP) molecule. Further, the receptor coupled with adenylate cyclase (AC) which catalyzes the conversion of ATP into cAMP (a second messenger for  $\beta$ 2-AR) by hydrolysis of GTP into GDP. The cAMP activates and regulates protein kinase A (PKA) which further mediates the transcription of genes and degradation of cAMP by phosphodiesterase (PDE) leading to termination of signaling [68].

Earlier it was determined that  $\beta$ 2-AR exhibits their inhibitory signals in immune cells via the canonical (PKA) signaling pathway. It has now been found that GPCR can also signal through a noncanonical pathway in addition to their classical signaling pathway [69]. Activation through the noncanonical signaling pathway is cell type dependent and G-protein independent, but rather the G-protein-coupled receptor kinases (GRKs) and  $\beta$ -arrestins are involved in activation of this noncanonical signaling pathway. Various types of GRKs phosphorylate specifically serine and threonine at C-terminal of the  $\beta$ 2-AR which further determines whether receptors undergo desensitization or initiate noncanonical signaling [70]. For example, phosphorylation of receptor by GRK5/6 initiates  $\beta$ -arrestin-mediated noncanonical signaling, while phosphorylation by GRK2 leads to  $\beta$ -arrestin-mediated desensitization of the receptor [71]. During noncanonical signaling,  $\beta$ -arrestin2 couples  $\beta$ 2-AR to MAPK signaling pathways which induces activation of transcription factors and allows their nuclear translocation. Activation of  $\beta$ 2-AR with high agonist concentration can lead to sustained activation of ERK1/2 via  $\beta$ -arrestin2. This explains why β2-AR activation can either enhance or suppress the proliferation of immune cells and cytokine production particularly at a high concentration of agonists [67, 72]. Studies suggest that during inflammatory conditions immune cells can switch from canonical to the noncanonical pathway [67, 68]. Engagement of  $\beta$ 2-AR receptors by agonists can result in immunomodulatory actions. Depending on the type of immune stimuli and timing of  $\beta$ 2-AR activation relative to immune activation,  $\beta$ 2-AR stimulation can positively or negatively regulate the response of immune activator [67, 73]. The initial data obtained in animal models of dopaminergic neurotoxicity suggests that the primary immunomodulatory mechanism of  $\beta$ 2-AR activation that regulates CNS inflammation in microglial cells occurs through the noncanonical  $\beta$ -arrestin2 pathway of activation.

# 7. β2-agonists

 $\beta$ -agonists are a group of pharmaceutical compounds or sympathomimetic drugs that mimic the effects of endogenous catecholamines such as epinephrine, norepinephrine, and dopamine. These drugs do not comprise a similar structure to

catecholamines but still directly or indirectly activate the  $\beta$ 2-adrenergic receptor. The first  $\beta$ -agonist was used around 5000 years ago in Chinese medicine where an ephedrine containing plant, Ma-huang, was used to treat respiratory problems [74]. Further research in the twentieth century has led to increased use of  $\beta$ -agonists for the treatment of respiratory diseases. The first  $\beta$ 2-AR selective agonist, Salbutamol was synthesized by Glaxo in 1968 [75]. Later, the same team at Glaxo modified Salbutamol into Salmeterol with long-lasting effects and reduced side effects. Recently, they have synthesized  $\beta$ 2-agonists with ultra-long-lasting effects such as Indacaterol [76]. After successful trials, these  $\beta$ 2-agonists were approved by the US Food and Drug Administration (FDA) for the treatment of respiratory diseases such as asthma and chronic obstructive pulmonary disease (COPD). Since 1968, a number of companies have labored to develop  $\beta$ 2-AR agonists, and some have now been commercialized for use in the treatment of COPD. A list of some of these agonists is given below and in **Table 1**.

## 7.1 Classification of β2-agonists

A pharmacogenetic study of  $\beta$ 2-agonists has summarized the relationship between polymorphisms in the  $\beta$ 2-adrenoreceptor (*ADRB2*) gene and the effects of select  $\beta$ 2-agonists [77]. Two hypotheses aim to account for the differences in functioning and *in vivo* half-lives of these compounds: exosite/exoreceptor or plasmalemma diffusion microkinetics. Briefly, the exosite hypothesis focuses on the ability of the side-chain of these compounds to interact with a distinct site on the receptor such that it allows the active component to "swing backand-forth" to activate the receptor. The plasmalemma diffusion microkinetic hypothesis suggests that high concentrations of agonists are achieved in close proximity to the receptor and allows for a longer duration of action [78]. Both of these hypotheses require further investigation and need to be studied within the CNS. Depending upon their mechanism and duration of action, all  $\beta$ 2-agonists are grouped into three major classes: short-acting, long-acting, and ultra-longacting  $\beta$ 2-agonists.

## 7.1.1 Short-acting $\beta$ -agonists (SABA)

These drugs are mostly hydrophilic in nature, access the active site of  $\beta$ -AR directly from the aqueous extracellular area and show the fast onset of action [79]. These SABAs bind to the receptor for short time; therefore, their duration of action is short. Some of the more common SABAs include Salbutamol (Ventolin), Albuterol (AccuNeb), Pirbuterol (Maxair), and Levalbuterol (Xopenex).

## 7.1.2 Long-acting $\beta$ -agonists (LABA)

These drugs are a frontline treatment for COPD, and usually prescribed alone or in combination with inhaled corticosteroids. LABAs are lipophilic in nature and taken up by cell membrane as a reservoir, progressively seep out and interact with the active site of the receptor [79]. They diffuse in the plasma membrane, where they interact with the active site of the  $\beta$ 2-AR which allows for the close proximity with the receptor and longer duration of action. The onset of action of these drugs is slower as compared to SABAs, but the duration of action is prolonged thereby, called as LABAs. The duration of action is also dependent on the concentration of the agonist. Salmeterol, Salmeterol with an inhaled corticosteroid, Formoterol, and Formoterol with an inhaled corticosteroid are commercially available LABAs and used in medication for asthma and COPD [80]. Adrenergic Receptors as Pharmacological Targets for Neuroinflammation and Neurodegeneration... DOI: http://dx.doi.org/10.5772/intechopen.81343

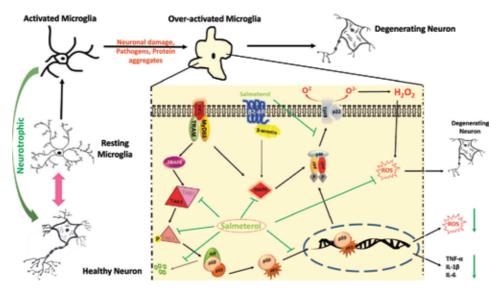
### 7.1.3 Ultra-long-acting $\beta$ -agonists

These agonists are also lipophilic in nature and onset of action is similar to LABAs, but the duration of action lasts longer than LABAs. Vilanterol with an inhaled corticosteroid and Indacaterol are ultra-LABAs, approved by FDA for the treatment of COPD [81].

## 8. β2-adrenergic receptors agonists in neuroprotection

The majority of adrenergic neurons are present in brainstem locus coeruleus (LC) nuclei, which is a predominant site for the production of norepinephrine (NE) in the brain. LC neurons play a key role in the regulation of cognitive behavior such as attention, mood, and arousal [82]. These neurons also play role in the development of the brain, mainly the neocortex [83]. The degeneration of LC-neurons has been identified in patients with PD and AD [84]. Also, the classical "monoamine hypothesis of depression" says that the deficiency of NE is a culprit for the cognitive impairment [85]. NE/noradrenaline, the primary neurotransmitter released by the LC neurons targets the adrenergic receptors present on the microglia and astrocytes in the brain [86]. NE-activated ARs on glial cells stimulate the second messenger system and maintain the homeostasis in the brain. Activation of AR on glial cells elicits anti-inflammatory actions, inhibits neuroinflammation, and thereby limits the degeneration of neurons [87]. Moreover, drugs that stimulate the release of NE/NA have potential to reduced inflammation and amyloid pathology in a mouse model of AD [88]. According to Braak's hypothesis, early stage of progression starts in LC before it spreads to SN [89]. Overall, these and many other studies suggest the role of the adrenergic signaling in neurodegeneration. Therefore, enhancing NE/NA signaling, transplanting noradrenergic neurons, or use of drugs that mimic the activity of NA/NE on glial cells have great potential to reverse or halt the progressive degeneration of neurons [90]. The endogenous agonist/ligand for  $\beta$ 2-AR is norepinephrine which acts as a neurotrophic factor and can influence protein/DNA synthesis in developing adult brain [91, 92]. NE protects cholinergic and dopaminergic cultured neurons against oxidative stress and catechol moiety of NE plays role in neuroprotection [93, 94]. It suggests that a compound containing catechol moiety, such as  $\beta$ -agonists, can mimic the neuroprotective effects of NE. Treatment with NE stimulates the synthesis of BDNF in astrocytes and neuron *in vitro* and *in vivo* [95, 96] and these neuroprotective effects were reversed by the antagonist of  $\alpha 1$ ,  $\beta 1$ , and  $\beta 2$ -AR [97].

The use of  $\beta$ 2-agonists as an adjunct therapy to L-DOPA in PD was first described in 1994 [98]. Chai et al. showed that the β2-AR activation enhances hippocampal neurogenesis, ameliorates memory deficits, and increases dendritic branching and spine density in a mouse model of Alzheimer's disease [99]. Recently, Mittal et al. have found that  $\beta$ 2-AR activation regulates the gene expression of  $\alpha$ -synuclein in various animal and *in vitro* models of PD. Salbutamol, a blood-brain-barrier-permeable  $\beta$ 2-agonist, reduces expression of SNCA gene via histone-3-lysine-27 acetylation of its promoter and enhancer. They also analyzed the pharmacological history of 4 million Norwegians over 11 years and found that Salbutamol was also associated with reduced risk of developing PD [100]. In a mouse model of Down syndrome, Formoterol, a long-acting β2-AR agonist, causes significant improvement in synaptic density and cognitive functions [101]. Salmeterol (Sal) is an inhaled long-acting highly selective β2-AR agonist which is currently being used as the active ingredient in Advair<sup>@</sup> as a bronchodilator. Our previous studies and others have shown that Salmeterol has anti-inflammatory and DA-neuroprotective activities, even at very low doses. Pre-treatment with Salmeterol protects DA neurons against LPS- and 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-induced toxicity in both in vitro and in vivo animal models of PD [42, 102]. The mechanism how Salmeterol regulates the activation of microglia is described in **Figure 1**. Collectively, these studies suggest that  $\beta$ 2-AR agonists not only protect neurons against degeneration, but also have anti-inflammatory effects, and therefore, hold significant promise for the treatments of a wide variety of neurodegenerative conditions including PD [43]. The clinical efficacy of  $\beta$ 2-AR agonists have been examined in various neurological disorders and few of them are summarized in **Table 2**.



#### Figure 1.

Schematic of microglia-mediated neurotoxicity and inhibitory effects of Salmeterol on microglial activation.

Disease Condition	Design	Doses	Drug	References
Spinal Cord Injury	Randomized controlled	4mg twice/day for 1st week then 8mg twice/day for 15 weeks	Albuterol	[129]
Alzheimer's Disease	Randomized controlled	20mg/2ml for 12 months	Formoterol	[130]
Multiple Sclerosis	Blinded controlled	4mg/day	Albuterol	[131]
Neuropathic pain	Controlled, double blinded	5mg twice/day for 28 days	Terbutaline	[132]
Memory and Cognition	Randomized controlled	4mg, single oral administration	Salbutamol	[133]
SMA	Uncontrolled	3-8mg/day for 6 months	Albuterol	[134]
ALS	Uncontrolled	60ug/day for 6 months	Clenbuterol	[135]
SBMA	Uncontrolled	20ug/day for 2days, then 40ug/day	Clenbuterol	[136]

SMA: Spinal Muscular Atrophy, SBMA: Spinal and Bulbar Muscular Atrophy, ALS: Amyotrophic Lateral Sclerosis.

# Table 2.

Clinical trials using  $\beta_2$ -agonist in neurological conditions.

Adrenergic Receptors as Pharmacological Targets for Neuroinflammation and Neurodegeneration... DOI: http://dx.doi.org/10.5772/intechopen.81343

## 9. \beta2-adrenergic receptors and neuroinflammation

Extensive previous investigations into the etiology of PD demonstrate a central role for the inflammatory microglial cell in the progression of PD. Thus, targeting neuroinflammation mediated by microglia may serve as a potential therapeutic benefit in the treatment of PD. Since traditional treatment for PD is aimed only at controlling the disease symptoms, the search for more effective neuroprotective therapies which target the cause of the disease is now receiving significant attention. Studies targeting neuroinflammation are aimed to promote the development of a novel therapeutic approach and aid in the drug discovery for neurodegenerative conditions such as PD.

One such anti-inflammatory approach that has been found to be effective in protection against dopaminergic neurodegeneration is accomplished by natural and therapeutic compounds that activate the  $\beta$ 2-AR. Brain cells including neurons, microglia, and astrocytes as well as immune cells express a high density of  $\beta$ 2-AR on their surface [66, 103]. Catecholamines such as epinephrine (adrenaline), norepinephrine (noradrenaline), and dopamine are the most abundant catecholamines found in the nervous system. As evidenced by many unrelated studies, catecholamines can modulate the immune response [87, 104]. Further studies have found that the endogenous agonist of  $\beta$ 2-AR, norepinephrine (NE), controls microglial motility and functions during pathogenic conditions [105]. NE also protects cortical neurons against microglia-mediated inflammation, while decreased levels of NE enhance microglial activation [106]. One study showed that  $\beta$ 2-AR negatively regulates NF-κB activation and stabilizes the NF-κB/IκBα complex via β-arrestin 2 in LPS activated murine macrophages [107]. Interestingly, activation of  $\beta$ 2-AR in astrocytes modulates TNF- $\alpha$ -induced inflammatory gene expression in vitro and *in vivo*. In addition, an *in vivo* study demonstrated increased expression of β2-AR in glial cells in response to neuronal injury. This suggests that β2-AR may provide a therapeutic target for regulation of glial cell functioning and the inflammatory response in the brain [108]. Activation of  $\beta$ 2-AR on astrocytes stimulates the release of trophic factors such as BDNF, bFGF, NGF-1, and TGF-β1 via canonical signaling, showing anti-apoptotic and neuroprotective effects in animal models of cerebral ischemia and excitotoxicity [109, 110]. It has also been shown that noradrenaline acting on β2-AR enhances the expression of anti-inflammatory and neurotrophic cytokine IL-10 in the brain. This suggests an endogenous ligand of  $\beta$ 2-AR is neuroprotective during inflammatory conditions in CNS disease pathology [108, 111]. Both canonical and noncanonical signaling of  $\beta$ 2-AR can selectively regulate the adaptive immune response [67], since  $\beta$ 2-AR are expressed by naïve CD4+ T (T-helper (Th0)) and Th1 cells but absent on Th2 cells [112, 113]. Naïve CD4+ T-cell treated with a  $\beta$ 2-AR agonist or NE suppresses the production of interferon (IFN)- $\gamma$ and IL-2 and affects their differentiation [114]. Collectively, these studies and several others suggest the role of  $\beta$ 2-AR in the regulation of immune response.

# 10. Molecular mechanism of inflammation in PD or molecular mediators of inflammation in PD

### 10.1 Effect of β2-AR agonists on NF-κB pathway

We have characterized and examined the effects of  $\beta$ 2-AR agonists including Salbutamol, Salmeterol, Indacaterol, and Vilanterol on neuroinflammation in models of PD *in vitro* and *in vivo*. However, the short-acting agonists were neuroprotective and able to reduce inflammation *in vitro* at higher doses, but the long-acting agonist showed beneficial effects at low concentration  $(10^{-9} \text{ M})$  in neurotoxicity and inflammatory models of PD. Salmeterol, a  $\beta$ 2-AR agonist, can effectively serve as a therapeutic treatment for PD by inhibiting microglia-mediated inflammatory responses *in vivo*. We have found that Salmeterol functions to inhibit innate pro-inflammatory response in both murine macrophages and microglia through its inhibition of the NF- $\kappa$ B signaling pathways [42]. We have also investigated whether Salmeterol is specific to neuroinflammation in PD or if it can be used as a universal anti-inflammatory drug against other chronic inflammatory diseases. To test this, we used murine macrophages stimulated with LPS from *Porphyromonas gingivalis* (*Pg*LPS), an oral pathogen as an *in vitro* model for the periodontal disease. We have found that Salmeterol shows similar anti-inflammatory effects on *Pg*LPS-stimulated macrophages [115]. Additionally, Feng et al. have also shown neuroprotective effects of  $\beta$ -arrestin2 via endogenous opioid arrest in inflammatory microglial cells [116].

## 10.2 Effect of β2-AR agonists on MAPK pathway

The agonist-activated β2-AR stimulates MAPK signaling pathway via noncanonical and G-protein independent pathway. Agonist-activated  $\beta$ 2-AR reduces phosphorylation of ERK1/2 and p38 MAPK in macrophages stimulated with LPS. In contrast,  $\beta$ 2-AR activation stimulates MAPK signaling and TNF- $\alpha$ , IL-12, and NO production in murine macrophages treated with PMA (phorbol 12-myristate-13-acetate) [73]. Similarly, our previous studies have shown that activation of  $\beta$ 2-AR with the high concentration of agonist (up to  $10^{-5}$  M) leads to sustained phosphorylation of ERK1/2 and enhanced production inflammatory mediators in murine microglia and macrophages [117]. High-dose treatment of  $\beta$ 2-AR agonists on mixed neuroglia culture enhances neurotoxicity via NADPH oxidase activity in the ERK-dependent manner [118]. Like others, we have found that the low-doses of the  $\beta$ 2-AR agonist Salmeterol reduces the MAPK activity, NF- $\kappa$ B activation and production of TNF- $\alpha$  in LPS-activated primary microglia [42]. We have also found that low-dose Salmeterol inhibits the phosphorylation of TAK1 (TGF- $\beta$ -activated kinase1) which is an upstream regulator of NF- $\kappa$ B signaling in LPS-stimulated microglia. We have also found that Salmeterol increases the expression of  $\beta$ -arrestin2 and enhances the interaction between  $\beta$ -arrestin2 and TAB1 (TAK1-binding protein), reduced TAK1/TAB1 mediated activation of NFKB and expression of pro-inflammatory genes. Furthermore, silencing of β-arrestin2 abrogates the anti-inflammatory effects of Salmeterol in LPS-stimulated BV2 cells [119]. These studies suggest that the anti-inflammatory effects of Salmeterol work through the inhibition of pro-inflammatory pathways in microglial cells.

## 10.3 The $\beta$ -arrestin-mediated biased effects of $\beta$ 2-AR agonist

Previous findings show that high dose Salmeterol enhances the expression of IL-1 $\beta$  and IL-6 mRNA and protein in unstimulated human monocytes and murine macrophages. These effects were  $\beta$ -arrestin2-dependent but PKA and NF- $\kappa$ B independent, while treatment with ERK1/2 and p38 MAPK inhibitor could reverse this effect [117]. This finding and several others suggest Salmeterol or other long-acting agonist have  $\beta$ -arrestin "biased" signaling of  $\beta$ 2-AR. These agonists activate receptors via  $\beta$ -arrestin signaling with a much greater extent than their effect on G-protein-dependent signaling [120]. Our studies suggest that a very low concentration of Salmeterol does not enhance cAMP signaling and its downstream mediators, while it activates the  $\beta$ -arrestin2-mediated signaling events [42].  $\beta$ -arrestin2 has been shown as a novel regulator of IkB stability via the direct interaction of  $\beta$ -arrestin2 and IkB in HEK293 cells [121]. In addition,  $\beta$ -arrestin2 negatively regulates the activation of NF- $\kappa$ B via direct binding with IkB $\alpha$  [122]. One study showed

## Adrenergic Receptors as Pharmacological Targets for Neuroinflammation and Neurodegeneration... DOI: http://dx.doi.org/10.5772/intechopen.81343

that overexpression of  $\beta$ -arrestin2 significantly reduces L-DOPA-induced dyskinesia in animal models of PD [123]. Collectively, these studies suggest that  $\beta$ 2-AR agonists can be used therapeutically not only to inhibit chronic inflammation and progressive degeneration of neurons, but also to treat some of the most debilitating neurologic symptoms in PD.

# 10.4 cAMP/PKA/CREB pathway induced by β2-AR

After binding with an agonist or endogenous ligand,  $\beta$ 2-AR normally activates the classical cAMP-dependent signaling pathway. The downstream effect of the cAMP/PKA pathway is the phosphorylation and nuclear translocation of the CREB transcription factor which further enhances the expression of cAMP-inducible genes [79]. Activation of CREB via this pathway regulates the synthesis of proteins which are mandatory for neuronal homeostasis [124]. The classical signaling of  $\beta$ 2-AR also increases the activity of PGC-1 $\alpha$  (Peroxisome proliferator-activated receptor gamma coactivator 1-alpha), which is a key regulator of mitochondrial biogenesis and ROS metabolism [125]. Activation of  $\beta$ 2-AR also elevated the release of neurotrophic factors via cAMP/PKA/CREB pathway and provides neuroprotective benefits against degeneration [126]. An endogenous agonist of  $\beta$ 2-AR (NE) affects immune cell functions, production of cytokines, and antibody secretion [112]. β2-AR agonists have anti-inflammatory activity and inhibit release of proinflammatory mediators via cAMP/PKA/CREB pathway and also by alternate cAMP-dependent pathway (cAMP/Epac1/2) [42, 127, 128]. We have also found that pro-inflammatory effects of high-dose of Salmeterol are through cAMP/Epac pathway, while the anti-inflammatory effects of low-dose of Salmeterol are independent on cAMP and Epac activation [42, 118].

# 11. From bench to bedside: challenges in translation to the clinic

The  $\beta$ 2-AR agonists discussed above are FDA-approved for the treatment of respiratory diseases such as asthma and COPD, but none of these  $\beta$ 2-AR agonists are specifically developed for PD. Although, Mittal et al. have found in a Norwegian population that using Salbutamol, a SABA, lower the risk of developing PD whereas the use of Propranolol, a β2-AR antagonist (commonly used to treat hypertension and certain other forms of heart disease) was associated with increased risk of PD [100]. Furthermore, this risk of developing PD was dependent on the duration of Salbutamol intake in those patients. In the patient population who used Salbutamol for at least 6 months, it was expected that 43 would develop PD, but only 23 patients were ultimately diagnosed with the disease (rate ratio 0.66). On the other hand, in the cohort who used Salbutamol for 2 months or less, there was no decreased risk of developing PD in this population [100]. In contrast, patients on Propranolol (which is also used as therapeutic for tremors in PD) for at least 1 year showed a significantly increased risk of developing PD compared to patients not on propranolol (rate ratio 2.2). Therefore, it is clear that patients on long-term Salbutamol (a  $\beta$ 2-AR agonist) had significantly decreased the risk of developing PD, while patients on long-term propranolol (a  $\beta$ 2-AR antagonist) therapy had significantly higher rates of PD, suggesting that  $\beta$ 2-AR inhibition is a highly significant risk factor in developing PD. When we compared the effectiveness of Salbutamol to Salmeterol (a more lipophilic drug) in animal models of PD, Salmeterol was much more effective both in vitro and in vivo in dopaminergic neuroprotection [42]. More importantly, we found that animals given Salmeterol treatment well before the appearance of symptoms in a long-term model of PD showed little evidence of dopaminergic neurodegeneration

compared to untreated animals. Taken together, this data suggests that administration of  $\beta$ 2-AR agonists may have a profound preventative effect on the development of PD. Since the blood-brain-barrier penetration is a major obstacle in the development of therapeutics targeting CNS disorders, it will be important to consider the importance of lipophilic properties, concentration within the CNS, as well as the specificity, half-life and safety in using  $\beta$ 2-AR agonists in older patients before and after the initial appearance of symptoms associated with PD. Consequently, these drugs require further investigation in a large cohort study to assess their utility as a potential therapeutic for PD and other neurodegenerative diseases.

# 12. Conclusion

Natural or synthetic activation or inhibition of the  $\beta$ 2-AR can have profound effects on the development and progression of Parkinson's disease, a chronic neurodegenerative disorder which involves both neuroinflammatory and cellular mechanisms in dopaminergic neurotoxicity. It is now clear that the therapeutic use of  $\beta$ 2-AR agonists can both inhibit the cause of neurodegeneration and activate a mechanism that can enhance recovery of patients with this disease, and serves as an important new therapeutic approach to the treatment of chronic neurodegenerative disorders.

# **Conflict of interest**

Authors declare no "conflict of interest."

# **Author details**

Monika Sharma<sup>1</sup> and Patrick M. Flood<sup>2,3,4\*</sup>

1 Department of Medical Microbiology and Immunology, Faculty of Medicine and Dentistry, University of Alberta, Edmonton, Alberta, Canada

2 Department of Dentistry, Neuroscience and Mental Health Institute, Faculty of Medicine and Dentistry, University of Alberta, Edmonton, Alberta, Canada

3 Department of Medical Microbiology and Immunology, Neuroscience and Mental Health Institute, Faculty of Medicine and Dentistry, University of Alberta, Edmonton, Alberta, Canada

4 School of Dentistry, Katz Group Centre for Pharmacy and Health Research, University of Alberta, Edmonton, Alberta, Canada

\*Address all correspondence to: pflood@ualberta.ca

# IntechOpen

© 2018 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/ by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. Adrenergic Receptors as Pharmacological Targets for Neuroinflammation and Neurodegeneration... DOI: http://dx.doi.org/10.5772/intechopen.81343

### References

[1] Gao HM, Hong JS. Why neurodegenerative diseases are progressive: Uncontrolled inflammation drives disease progression. Trends in Immunology. 2008;**29**(8):357-365

[2] Poewe W, Seppi K, Tanner CM, Halliday GM, Brundin P, Volkmann J, et al. Parkinson disease. Nature Reviews Disease Primers. 2017;**3**:17013

[3] Nolan YM, Sullivan AM, Toulouse
A. Parkinson's disease in the nuclear age of neuroinflammation.
Trends in Molecular Medicine.
2013;19(3):187-196

[4] Moustafa AA, Chakravarthy S, Phillips JR, Gupta A, Keri S, Polner B, et al. Motor symptoms in Parkinson's disease: A unified framework. Neuroscience and Biobehavioral Reviews. 2016;**68**:727-740

[5] Parkinson's Disease. PubMed -NCBI [Internet]. Available from: https://www.ncbi.nlm.nih.gov/ pubmed/?term=25904081[Cited: August 2, 2018]

[6] Löhle M, Storch A, Reichmann H. Beyond tremor and rigidity: Nonmotor features of Parkinson's disease. Journal of Neural Transmission. 2009;**116**(11):1483-1492

[7] Gasser T. Molecular pathogenesis of Parkinson disease: Insights from genetic studies. Expert Reviews in Molecular Medicine. 2009;**11**:e22

[8] Spatola M, Wider C. Genetics of Parkinson's disease: The yield.Parkinsonism & Related Disorders.2014;20(Suppl 1):S35-S38

[9] Chade AR, Kasten M, Tanner CM. Nongenetic causes of Parkinson's disease. Journal of Neural Transmission. Supplementum. 2006;**70**:147-151 [10] Wei Z, Li X, Li X, Liu Q, Cheng Y. Oxidative stress in Parkinson's disease: A systematic review and meta-analysis. Frontiers in Molecular Neuroscience. 2018;**11**:236

[11] Doehner J, Genoud C, Imhof C, Krstic D, Knuesel I. Extrusion of misfolded and aggregated proteins—A protective strategy of aging neurons? The European Journal of Neuroscience. 2012;**35**(12):1938-1950

[12] Blaylock RL. Parkinson's disease: Microglial/macrophage-induced immunoexcitotoxicity as a central mechanism of neurodegeneration. Surgical Neurology International. 2017;**8**:65

[13] Ryan BJ, Hoek S, Fon EA, Wade-Martins R. Mitochondrial dysfunction and mitophagy in Parkinson's: From familial to sporadic disease. Trends in Biochemical Sciences.
2015;40(4):200-210

[14] Molteni M, Rossetti C. Neurodegenerative diseases: The immunological perspective. Journal of Neuroimmunology. 2017;**313**:109-115

[15] Lehnardt S. Innate immunity and neuroinflammation in the CNS: The role of microglia in toll-like receptormediated neuronal injury. Glia. 2010;**58**(3):253-263

[16] Kim C, Ho DH, Suk JE, You S, Michael S, Kang J, et al. Neuronreleased oligomeric  $\alpha$ -synuclein is an endogenous agonist of TLR2 for paracrine activation of microglia. Nature Communications. 2013;**4**:1562

[17] Hirsch EC, Hunot S. Neuroinflammation in Parkinson's disease: A target for neuroprotection? Lancet Neurology. 2009;**8**(4):382-397 [18] Funk N, Wieghofer P, Grimm S, Schaefer R, Bühring HJ, Gasser T, et al. Characterization of peripheral hematopoietic stem cells and monocytes in Parkinson's disease. Movement Disorders. 2013;**28**(3):392-395

[19] Ginhoux F, Garel S. The mysterious origins of microglia. Nature Neuroscience. 2018;**21**(7):897-899

[20] Li Q, Barres BA. Microglia and macrophages in brain homeostasis and disease. Nature Reviews. Immunology. 2018;**18**(4):225-242

[21] Glezer I, Simard AR, Rivest S. Neuroprotective role of the innate immune system by microglia. Neuroscience. 2007;**147**(4):867-883

[22] Block ML, Zecca L, Hong JS.Microglia-mediated neurotoxicity:Uncovering the molecular mechanisms.Nature Reviews. Neuroscience.2007;8(1):57-69

[23] McGeer PL, Itagaki S, Boyes BE, McGeer EG. Reactive microglia are positive for HLA-DR in the substantia nigra of Parkinson's and Alzheimer's disease brains. Neurology. 1988;**38**(8):1285-1291

[24] Imamura K, Hishikawa N, Sawada M, Nagatsu T, Yoshida M, Hashizume Y. Distribution of major histocompatibility complex class II-positive microglia and cytokine profile of Parkinson's disease brains. Acta Neuropathologica. 2003;**106**(6):518-526

[25] Koshimori Y, Ko JH, Mizrahi R, Rusjan P, Mabrouk R, Jacobs MF, et al. Imaging striatal microglial activation in patients with Parkinson's disease. PLoS One. 2015;**10**(9):e0138721

[26] Perry VH, Holmes C. Microglial priming in neurodegenerative disease. Nature Reviews. Neurology. 2014;**10**(4):217-224 [27] Wang Q, Liu Y, Zhou J. Neuroinflammation in Parkinson's disease and its potential as therapeutic target. Translational Neurodegeneration. 2015;4:19

[28] Tambasco N, Romoli M, Calabresi
P. Levodopa in Parkinson's disease:
Current status and future developments.
Current Neuropharmacology.
2017;16(8):1239-1252

[29] Connolly BS, Lang AE.Pharmacological treatment ofParkinson disease: A review. Journal of the American Medical Association.2014;**311**(16):1670-1683

[30] You H, Mariani LL, Mangone G, de Nailly DLF, Charbonnier-Beaupel F, Corvol JC. Molecular basis of dopamine replacement therapy and its side effects in Parkinson's disease. Cell and Tissue Research. 2018;**373**(1):111-135

[31] Sharma M, Arbabzada N, Flood PM. Inflammation: Role in Parkinson's disease and target for therapy. In: Challenges in Parkinson's Disease. Rijeka, Croatia: InTech; 2016

[32] Castaño A, Herrera AJ, Cano J, Machado A. The degenerative effect of a single intranigral injection of LPS on the dopaminergic system is prevented by dexamethasone, and not mimicked by rh-TNFalpha, IL-1beta and IFN-gamma. Journal of Neurochemistry. 2002;**81**(1):150-157

[33] Manthripragada AD, Schernhammer ES, Qiu J, Friis S, Wermuth L, Olsen JH, et al. Nonsteroidal anti-inflammatory drug use and the risk of Parkinson's disease. Neuroepidemiology. 2011;**36**(3):155-161

[34] Gao X, Chen H, Schwarzschild MA, Ascherio A. Use of ibuprofen and risk of Parkinson disease. Neurology. 2011;**76**(10):863-869 Adrenergic Receptors as Pharmacological Targets for Neuroinflammation and Neurodegeneration... DOI: http://dx.doi.org/10.5772/intechopen.81343

[35] Casper D, Yaparpalvi U, Rempel
N, Werner P. Ibuprofen protects
dopaminergic neurons against glutamate
toxicity in vitro. Neuroscience Letters.
2000;289(3):201-204

[36] Qian L, Xu Z, Zhang W, Wilson B, Hong JS, Flood PM. Sinomenine, a natural dextrorotatory morphinan analog, is anti-inflammatory and neuroprotective through inhibition of microglial NADPH oxidase. Journal of Neuroinflammation. 2007;**4**:23

[37] Qian L, Wei SJ, Zhang D, Hu X, Xu Z, Wilson B, et al. Potent anti-inflammatory and neuroprotective effects of TGFbeta1 are mediated through the inhibition of ERK and p47phox-Ser345 phosphorylation and translocation in microglia. Journal of Immunology. 2008;**181**(1):660-668

[38] Qian L, Block ML, Wei SJ, Lin CF, Reece J, Pang H, et al. Interleukin-10 protects lipopolysaccharide-induced neurotoxicity in primary midbrain cultures by inhibiting the function of NADPH oxidase. The Journal of Pharmacology and Experimental Therapeutics. 2006;**319**(1):44-52

[39] Zhu Y, Liu Z, Peng YP, Qiu
YH. Interleukin-10 inhibits
neuroinflammation-mediated apoptosis
of ventral mesencephalic neurons via
JAK-STAT3 pathway. International
Immunopharmacology. 2017;50:353-360

[40] Zhang F, Qian L, Flood PM, Shi JS, Hong JS, Gao HM. Inhibition of IkappaB kinase-beta protects dopamine neurons against lipopolysaccharideinduced neurotoxicity. The Journal of Pharmacology and Experimental Therapeutics. 2010;**333**(3):822-833

[41] Qian L, Gao X, Pei Z, Wu X, Block M, Wilson B, et al. NADPH oxidase inhibitor DPI is neuroprotective at femtomolar concentrations through inhibition of microglia over-activation. Parkinsonism & Related Disorders. 2007;**13**(Suppl 3):S316-S320

[42] Qian L, Wu H, Chen SH, Zhang D, Ali SF, Peterson L, et al.  $\beta$ 2-adrenergic receptor activation prevents rodent dopaminergic neurotoxicity by inhibiting microglia via a novel signaling pathway. Journal of Immunology. 2011;**186**(7):4443-4454

[43] Peterson L, Ismond KP, Chapman E, Flood P. Potential benefits of therapeutic use of  $\beta$ 2-adrenergic receptor agonists in neuroprotection and Parkinsonµs disease. Journal of Immunology Research. 2014;**2014**:103780

[44] Qian L, Tan KS, Wei SJ, Wu HM, Xu Z, Wilson B, et al. Microgliamediated neurotoxicity is inhibited by morphine through an opioid receptorindependent reduction of NADPH oxidase activity. Journal of Immunology. 2007;**179**(2):1198-1209

[45] Li G, Cui G, Tzeng NS, Wei SJ, Wang T, Block ML, et al. Femtomolar concentrations of dextromethorphan protect mesencephalic dopaminergic neurons from inflammatory damage. The FASEB Journal. 2005;**19**(6):489-496

[46] Ahlquist RP. The adrenergic receptor. Journal of Pharmaceutical Sciences. 1966;55(4):359-367

[47] Ahlquist RP. Historical perspective.Classification of adrenoreceptors.Journal of Autonomic Pharmacology.1980;1(1):101-106

[48] Ahlquist RP. Adrenergic receptors: A personal and practical view.Perspectives in Biology and Medicine.1973;17(1):119-122

[49] Berthelsen S, Pettinger WA. A functional basis for classification of alpha-adrenergic receptors. Life Sciences. 1977;**21**(5):595-606

[50] Fraser CM, Venter JC. Monoclonal antibodies to beta-adrenergic receptors: Use in purification and molecular characterization of beta receptors. Proceedings of the National Academy of Sciences of the United States of America. 1980;77(12):7034-7038

[51] Emorine LJ, Marullo S, Briend-Sutren MM, Patey G, Tate K,
Delavier-Klutchko C, et al. Molecular characterization of the human beta
3-adrenergic receptor. Science.
1989;245(4922):1118-1121

[52] Siegel GJ, Agranoff BW, Albers RW, et al. Basic Neurochemistry: Molecular, Cellular and Medical Aspects. 6th ed. Philadelphia, PA: Lippincott-Raven; 1999

[53] Fredriksson R, Lagerström MC, Lundin LG, Schiöth HB. The G-proteincoupled receptors in the human genome form five main families. Phylogenetic analysis, paralogon groups, and fingerprints. Molecular Pharmacology. 2003;**63**(6):1256-1272

[54] Garland SL. Are GPCRs still a source of new targets? Journal of Biomolecular Screening. 2013;**18**(9):947-966

[55] Kobilka BK, MacGregor C, Daniel K, Kobilka TS, Caron MG, Lefkowitz RJ. Functional activity and regulation of human beta 2-adrenergic receptors expressed in Xenopus oocytes. The Journal of Biological Chemistry. 1987;**262**(32):15796-15802

[56] Dixon RA, Kobilka BK, Strader DJ, Benovic JL, Dohlman HG, Frielle T, et al. Cloning of the gene and cDNA for mammalian beta-adrenergic receptor and homology with rhodopsin. Nature. 1986;**321**(6065):75-79

[57] Cherezov V, Rosenbaum DM, Hanson MA, Rasmussen SGF, Thian FS, Kobilka TS, et al. Highresolution crystal structure of an engineered human beta2-adrenergic G protein-coupled receptor. Science. 2007;**318**(5854):1258-1265

[58] Palczewski K, Kumasaka T, Hori T, Behnke CA, Motoshima H, Fox BA, et al. Crystal structure of rhodopsin: A G protein-coupled receptor. Science. 2000;**289**(5480):739-745

[59] Collins S, Lohse MJ, O'Dowd B, Caron MG, Lefkowitz RJ. Structure and regulation of G protein-coupled receptors: The beta 2-adrenergic receptor as a model. Vitamins and Hormones. 1991;**46**:1-39

[60] Rands E, Candelore MR, Cheung AH, Hill WS, Strader CD, Dixon
RA. Mutational analysis of betaadrenergic receptor glycosylation.
The Journal of Biological Chemistry.
1990;265(18):10759-10764

[61] O'Dowd BF, Hnatowich M, Caron MG, Lefkowitz RJ, Bouvier M. Palmitoylation of the human beta 2-adrenergic receptor. Mutation of Cys341 in the carboxyl tail leads to an uncoupled nonpalmitoylated form of the receptor. The Journal of Biological Chemistry. 1989;**264**(13):7564-7569

[62] Dohlman HG, Caron MG, DeBlasi A, Frielle T, Lefkowitz RJ. Role of extracellular disulfide-bonded cysteines in the ligand binding function of the beta 2-adrenergic receptor. Biochemistry. 1990;**29**(9):2335-2342

[63] Carstairs JR, Nimmo AJ, Barnes PJ. Autoradiographic visualization of beta-adrenoceptor subtypes in human lung. The American Review of Respiratory Disease. 1985;**132**(3):541-547

[64] Simon V, Robin MT, Legrand C, Cohen-Tannoudji J. Endogenous G protein-coupled receptor kinase 6 triggers homologous beta-adrenergic receptor desensitization in primary uterine smooth muscle cells. Endocrinology. 2003;**144**(7):3058-3066 Adrenergic Receptors as Pharmacological Targets for Neuroinflammation and Neurodegeneration... DOI: http://dx.doi.org/10.5772/intechopen.81343

[65] Barnes PJ. Beta-adrenoceptors on smooth muscle, nerves and inflammatory cells. Life Sciences. 1993;**52**(26):2101-2109

[66] Tanaka KF, Kashima H, Suzuki H, Ono K, Sawada M. Existence of functional beta1- and beta2adrenergic receptors on microglia. Journal of Neuroscience Research. 2002;**70**(2):232-237

[67] Lorton D, Bellinger DL. Molecular mechanisms underlying  $\beta$ -adrenergic receptor-mediated cross-talk between sympathetic neurons and immune cells. International Journal of Molecular Sciences. 2015;**16**(3):5635-5665

[68] Barisione G, Baroffio M, Crimi E, Brusasco V. Beta-adrenergic agonists. Pharmaceuticals. 2010;**3**(4):1016-1044

[69] Shukla AK, Xiao K, Lefkowitz
 RJ. Emerging paradigms of
 β-arrestin-dependent seven
 transmembrane receptor signaling.
 Trends in Biochemical Sciences.
 2011;36(9):457-469

[70] Luttrell LM, Gesty-Palmer D. Beyond desensitization: Physiological relevance of arrestin-dependent signaling. Pharmacological Reviews. 2010;**62**(2):305-330

[71] Claing A, Laporte SA, Caron MG, Lefkowitz RJ. Endocytosis of G protein-coupled receptors: Roles of G protein-coupled receptor kinases and beta-arrestin proteins. Progress in Neurobiology. 2002;**66**(2):61-79

[72] Shenoy SK, Drake MT, Nelson
CD, Houtz DA, Xiao K, Madabushi
S, et al. Beta-arrestin-dependent, G
protein-independent ERK1/2 activation
by the beta2 adrenergic receptor.
The Journal of Biological Chemistry.
2006;281(2):1261-1273

[73] Szelenyi J, Selmeczy Z, Brozik A, Medgyesi D, Magocsi M. Dual beta-adrenergic modulation in the immune system: Stimulus-dependent effect of isoproterenol on MAPK activation and inflammatory mediator production in macrophages. Neurochemistry International. 2006;**49**(1):94-103

[74] Chu EK, Drazen JM. Asthma: One hundred years of treatment and onward. American Journal of Respiratory and Critical Care Medicine. 2005;**171**(11):1202-1208

[75] Brittain RT. A comparison of the pharmacology of salbutamol with that of isoprenaline, orciprenaline and trimetoquinol. Postgraduate Medical Journal. 1971;**47**(Suppl):11-16

[76] Billington CK, Penn RB, Hall IP. β2 Agonists. Handbook of Experimental Pharmacology. 2017;**237**:23-40

[77] Hizawa N. Pharmacogenetics of
 β2-agonists. Allergology International.
 2011;60(3):239-246

[78] Morgan DJ. Clinical pharmacokinetics of beta-agonists.Clinical Pharmacokinetics.1990;18(4):270-294

[79] Johnson M. Molecular mechanisms of beta(2)-adrenergic receptor function, response, and regulation. The Journal of Allergy and Clinical Immunology. 2006;**117**(1): 18-24; quiz 25

[80] Johnson M. Beta2-adrenoceptors: Mechanisms of action of beta2-agonists.Paediatric Respiratory Reviews.2001;2(1):57-62

[81] Chowdhury BA, Dal Pan G. The FDA and safe use of long-acting betaagonists in the treatment of asthma. The New England Journal of Medicine. 2010;**362**(13):1169-1171

[82] Schwarz LA, Luo L. Organization of the locus coeruleus-norepinephrine

system. Current Biology. 2015;**25**(21):R1051-R1056

[83] Foote SL, Bloom FE, Aston-Jones
G. Nucleus locus ceruleus: New evidence of anatomical and physiological specificity. Physiological Reviews.
1983;63(3):844-914

[84] Peterson AC, Li CSR. Noradrenergic dysfunction in Alzheimer's and Parkinson's diseases—An overview of imaging studies. Frontiers in Aging Neuroscience. 2018;**10**:127

[85] Schildkraut JJ. The catecholamine hypothesis of affective disorders: A review of supporting evidence. The American Journal of Psychiatry. 1965;122(5):509-522

[86] Zorec R, Parpura V, Verkhratsky A. Preventing neurodegeneration by adrenergic astroglial excitation. The FEBS Journal. 2018;**285**(19):3645-3656

[87] Feinstein DL, Kalinin S, Braun D. Causes, consequences, and cures for neuroinflammation mediated via the locus coeruleus: Noradrenergic signaling system. Journal of Neurochemistry. 2016;139(Suppl 2):154-178

[88] Heneka MT, Nadrigny F, Regen T, Martinez-Hernandez A, Dumitrescu-Ozimek L, Terwel D, et al. Locus ceruleus controls Alzheimer's disease pathology by modulating microglial functions through norepinephrine. Proceedings of the National Academy of Sciences of the United States of America. 2010;**107**(13):6058-6063

[89] Braak H, Del Tredici K, Rüb U, de Vos RAI, Jansen Steur ENH, Braak E. Staging of brain pathology related to sporadic Parkinson's disease. Neurobiology of Aging. 2003;**24**(2):197-211

[90] Weinshenker D. Long road to ruin: Noradrenergic dysfunction in

neurodegenerative disease. Trends in Neurosciences. 2018;**41**(4):211-223

[91] Lauder JM. Neurotransmitters as growth regulatory signals: Role of receptors and second messengers. Trends in Neurosciences.1993;16(6):233-240

[92] Day JS, O'Neill E, Cawley C, Aretz NK, Kilroy D, Gibney SM, et al. Noradrenaline acting on astrocytic β2-adrenoceptors induces neurite outgrowth in primary cortical neurons. Neuropharmacology. 2014;77:234-248

[93] Traver S, Salthun-Lassalle B, Marien M, Hirsch EC, Colpaert F, Michel PP. The neurotransmitter noradrenaline rescues septal cholinergic neurons in culture from degeneration caused by low-level oxidative stress. Molecular Pharmacology. 2005;**67**(6):1882-1891

[94] Troadec JD, Marien M, Darios F, Hartmann A, Ruberg M, Colpaert F, et al. Noradrenaline provides long-term protection to dopaminergic neurons by reducing oxidative stress. Journal of Neurochemistry. 2001;**79**(1):200-210

[95] Musazzi L, Rimland JM, Ieraci A, Racagni G, Domenici E, Popoli M. Pharmacological characterization of BDNF promoters I, II and IV reveals that serotonin and norepinephrine input is sufficient for transcription activation. The International Journal of Neuropsychopharmacology. 2014;17(5):779-791

[96] Chen MJ, Nguyen TV, Pike CJ, Russo-Neustadt AA. Norepinephrine induces BDNF and activates the PI-3K and MAPK cascades in embryonic hippocampal neurons. Cellular Signalling. 2007;**19**(1):114-128

[97] Juric DM, Loncar D, Carman-Krzan M. Noradrenergic stimulation of BDNF synthesis in astrocytes: Mediation via alpha1- and beta1/beta2-adrenergic Adrenergic Receptors as Pharmacological Targets for Neuroinflammation and Neurodegeneration... DOI: http://dx.doi.org/10.5772/intechopen.81343

receptors. Neurochemistry International. 2008;**52**(1-2):297-306

[98] Alexander GM, Schwartzman RJ, Nukes TA, Grothusen JR, Hooker MD. Beta 2-adrenergic agonist as adjunct therapy to levodopa in Parkinson's disease. Neurology. 1994;**44**(8):1511-1513

[99] Chai GS, Wang YY, Yasheng A, Zhao P. Beta 2-adrenergic receptor activation enhances neurogenesis in Alzheimer's disease mice. Neural Regeneration Research. 2016;**11**(10):1617-1624

[100] Mittal S, Bjørnevik K, Im DS, Flierl A, Dong X, Locascio JJ, et al.  $\beta$ 2-Adrenoreceptor is a regulator of the  $\alpha$ -synuclein gene driving risk of Parkinson's disease. Science. 2017;**357**(6354):891-898

[101] Dang V, Medina B, Das D, Moghadam S, Martin KJ, Lin B, et al. Formoterol, a long-acting  $\beta 2$ adrenergic agonist, improves cognitive function and promotes dendritic complexity in a mouse model of Down syndrome. Biological Psychiatry. 2014;75(3):179-188

[102] Abdelmotilib H, West AB.Breathing new life into an old target:Pulmonary disease drugs for Parkinson's disease therapy. Genome Medicine.2017;9(1):88

[103] Cash R, Raisman R, Lanfumey L, Ploska A, Agid Y. Cellular localization of adrenergic receptors in rat and human brain. Brain Research. 1986;**370**(1):127-135

[104] Braun D, Madrigal JLM, Feinstein DL. Noradrenergic regulation of glial activation: Molecular mechanisms and therapeutic implications. Current Neuropharmacology. 2014;**12**(4):342-352

[105] Gyoneva S, Traynelis SF. Norepinephrine modulates the motility of resting and activated microglia via different adrenergic receptors. The Journal of Biological Chemistry. 2013;**288**(21):15291-15302

[106] Madrigal JLM, Feinstein DL, Dello Russo C. Norepinephrine protects cortical neurons against microglial-induced cell death. Journal of Neuroscience Research. 2005;**81**(3):390-396

[107] Kizaki T, Izawa T, Sakurai T, Haga S, Taniguchi N, Tajiri H, et al. Beta2adrenergic receptor regulates toll-like receptor-4-induced nuclear factorkappaB activation through beta-arrestin 2. Immunology. 2008;**124**(3):348-356

[108] Laureys G, Gerlo S, Spooren A, Demol F, De Keyser J, Aerts JL.  $\beta$ 2-adrenergic agonists modulate TNF- $\alpha$ induced astrocytic inflammatory gene expression and brain inflammatory cell populations. Journal of Neuroinflammation. 2014;**11**:21

[109] Culmsee C, Semkova I, Krieglstein J. NGF mediates the neuroprotective effect of the beta2-adrenoceptor agonist clenbuterol in vitro and in vivo: Evidence from an NGF-antisense study. Neurochemistry International. 1999;**35**(1):47-57

[110] Gleeson LC, Ryan KJ, Griffin EW, Connor TJ, Harkin A. The  $\beta$ 2-adrenoceptor agonist clenbuterol elicits neuroprotective, antiinflammatory and neurotrophic actions in the kainic acid model of excitotoxicity. Brain, Behavior, and Immunity. 2010;**24**(8):1354-1361

[111] McNamee EN, Ryan KM, Griffin EW, González-Reyes RE, Ryan KJ, Harkin A, et al. Noradrenaline acting at central beta-adrenoceptors induces interleukin-10 and suppressor of cytokine signaling-3 expression in rat brain: Implications for neurodegeneration. Brain, Behavior, and Immunity. 2010;**24**(4):660-671 [112] Kin NW, Sanders VM. It takes nerve to tell T and B cells what to do. Journal of Leukocyte Biology. 2006;**79**(6):1093-1104

[113] Kohm AP, Sanders VM. Norepinephrine and beta 2-adrenergic receptor stimulation regulate CD4+ T and B lymphocyte function in vitro and in vivo. Pharmacological Reviews. 2001;**53**(4):487-525

[114] Swanson MA, Lee WT, Sanders VM. IFN-gamma production by Th1 cells generated from naive CD4+ T cells exposed to norepinephrine.
Journal of Immunology.
2001;166(1):232-240

[115] Sharma M, Patterson L, Chapman E, Flood PM. Salmeterol, a long-acting  $\beta$ 2-adrenergic receptor agonist, inhibits macrophage activation by lipopolysaccharide from *Porphyromonas gingivalis*. Journal of Periodontology. 2017;**88**(7):681-692

[116] Feng X, Wu CY, Burton FH, Loh HH, Wei L-N.  $\beta$ -Arrestin protects neurons by mediating endogenous opioid arrest of inflammatory microglia. Cell Death and Differentiation. 2014;**21**(3):397-406

[117] Tan KS, Nackley AG, Satterfield K, Maixner W, Diatchenko L, Flood PM. Beta2 adrenergic receptor activation stimulates pro-inflammatory cytokine production in macrophages via PKA- and NF-kappaB-independent mechanisms. Cellular Signalling. 2007;**19**(2):251-260

[118] Qian L, Hu X, Zhang D, Snyder A, Wu HM, Li Y, et al. Beta2 adrenergic receptor activation induces microglial NADPH oxidase activation and dopaminergic neurotoxicity through an ERKdependent/protein kinase A-independent pathway. Glia. 2009;**57**(15):1600-1609 [119] Sharma M, Flood PM.  $\beta$ -arrestin2 regulates the anti-inflammatory effects of Salmeterol in lipopolysaccharidestimulated BV2 cells. Journal of Neuroimmunology. 2018;**325**:10-19

[120] Drake MT, Violin JD, Whalen EJ, Wisler JW, Shenoy SK, Lefkowitz RJ. Beta-arrestin-biased agonism at the beta2-adrenergic receptor. The Journal of Biological Chemistry. 2008;**283**(9):5669-5676

[121] Gao H, Sun Y, Wu Y, Luan B, Wang Y, Qu B, et al. Identification of beta-arrestin2 as a G protein-coupled receptor-stimulated regulator of NF-kappaB pathways. Molecular Cell. 2004;**14**(3):303-317

[122] Witherow DS, Garrison TR, Miller WE, Lefkowitz RJ. Beta-Arrestin inhibits NF-kappaB activity by means of its interaction with the NF-kappaB inhibitor IkappaBalpha. Proceedings of the National Academy of Sciences of the United States of America. 2004;**101**(23):8603-8607

[123] Urs NM, Bido S, Peterson SM, Daigle TL, Bass CE, Gainetdinov RR, et al. Targeting  $\beta$ -arrestin2 in the treatment of L-DOPA-induced dyskinesia in Parkinson's disease. Proceedings of the National Academy of Sciences of the United States of America. 2015;**112**(19):E2517-E2526

[124] Zhou HC, Sun YY, Cai W, He XT, Yi F, Li BM, et al. Activation of  $\beta$ 2-adrenoceptor enhances synaptic potentiation and behavioral memory via cAMP-PKA signaling in the medial prefrontal cortex of rats. Learning and Memory (Cold Spring Harbor, N.Y.). 2013;**20**(5):274-284

[125] Peterson YK, Cameron RB, Wills LP, Trager RE, Lindsey CC, Beeson CC, et al.  $\beta$ 2-adrenoceptor agonists in the regulation of mitochondrial biogenesis. Bioorganic and Medicinal Chemistry Letters. 2013;**23**(19):5376-5381 Adrenergic Receptors as Pharmacological Targets for Neuroinflammation and Neurodegeneration... DOI: http://dx.doi.org/10.5772/intechopen.81343

[126] Counts SE, Mufson EJ. Noradrenaline activation of neurotrophic pathways protects against neuronal amyloid toxicity. Journal of Neurochemistry. 2010;**113**(3):649-660

[127] Theron AJ, Steel HC, Tintinger GR, Feldman C, Anderson R. Can the anti-inflammatory activities of  $\beta$ 2-agonists be harnessed in the clinical setting? Drug Design, Development and Therapy. 2013;7:1387-1398

[128] Carnevale D, De Simone R, Minghetti L. Microglia-neuron interaction in inflammatory and degenerative diseases: Role of cholinergic and noradrenergic systems. CNS & Neurological Disorders: Drug Targets. 2007;**6**:388-397

[129] Schilero GJ, Hobson JC, Singh K, Spungen AM, Bauman WA, Radulovic M. Bronchodilator effects of ipratropium bromide and albuterol sulfate among subjects with tetraplegia. Journal of Spinal Cord Medicine. 2018;**41**(1):42-47

[130] Phillips C, Fahimi A, Das D, Mojabi FS, Ponnusamy R, Salehi A. Noradrenergic system in down syndrome and alzheimer's disease a target for therapy. Current Alzheimer Research. 2016;**13**(1):68-83

[131] Khoury SJ, Healy BC, Kivisäkk P, Viglietta V, Egorova S, Guttmann CRG, et al. A randomized controlled double-masked trial of albuterol add-on therapy in patients with multiple sclerosis. Archives of Neurology. 2010;**67**(9):1055-1061

[132] de Quervain D, Schwabe L, Roozendaal B. Stress, glucocorticoids and memory: Implications for treating fear-related disorders. Nature Reviews Neuroscience. 2017;**18**(1):7-19

[133] Salvat E, Yalcin I, Muller A, Barrot M. A comparison of early and late treatments on allodynia and its chronification in experimental neuropathic pain. Molecular Pain. 2018;**14**:17448069-17749683

[134] Sorarù G, Pegoraro E, Spinella P, Turra S, D'Ascenzo C, Baggio L, et al. A pilot trial with clenbuterol in amyotrophic lateral sclerosis. Amyotrophic Lateral Sclerosis : Official Publication of the World Federation of Neurology Research Group on Motor Neuron Diseases. 2006;7(4):246-248

[135] Kinali M, Mercuri E, Main M, De Biasia F, Karatza A, Higgins R, et al. Pilot trial of albuterol in spinal muscular atrophy. Neurology. 2002;**59**(4):609-610

[136] Querin G, D'Ascenzo C, Peterle E, Ermani M, Bello L, Melacini P, et al. Pilot trial of clenbuterol in spinal and bulbar muscular atrophy. Neurology. 2013;**80**(23):2095-2098

### Section 2

# Neuroprotection to the CNS

### Chapter 5

## Protecting the Aging Retina

Shen Nian and Amy C.Y. Lo

### Abstract

Aging retina, notably the aging macula, is prone to develop degenerative diseases, such as age-related macular degeneration (AMD), the leading cause of visual loss in individuals aged 65 or above in developed countries. However, current treatments are very limited. Since degeneration, dysfunction, and death of retinal neurons are demonstrated in the pathogenesis of AMD, neuroprotective strategies could serve as a possible way to treat AMD. In this chapter, we will briefly introduce risk factors, pathophysiology, affected neurons, classification, clinical manifestation, and current treatments of AMD. Finally, neuroprotection in both AMD animal models and patients will be discussed.

**Keywords:** neuroprotection, degeneration, photoreceptor, age-related macular degeneration, vision loss

### 1. Introduction

Retina, which forms the innermost layer of the eyeball, is considered as the end organ of the central nervous system. Macula, located in the central and posterior part of retina, possesses the highest concentration of photoreceptors and, therefore, is responsible for central vision and high-resolution visual acuity (VA). The fovea is a tiny pit in the center of the macula and in charge of the central, sharpest vision. Unfortunately, the macula is more prone to experience degenerative changes with age, such as age-related macular degeneration (AMD), leading to visual impairment.

Due to the increase of life expectancy worldwide, the size of aging population will become larger in the coming decades [1]. According to "World Population Prospects: The 2017 Revision" released by the United Nations, the number of persons aged 60 or above will more than double, from 962 million in 2017 to 2.1 billion by 2050. The number of people aged 80 or above is predicted to triple by 2050, from 137 million in 2017 to 425 million by 2050 [2]. In mainland of China, people aged above 65 represented 11.4% of the total population in 2017 [3]. As a consequence, the increasing prevalence of AMD will be foreseen globally in the future.

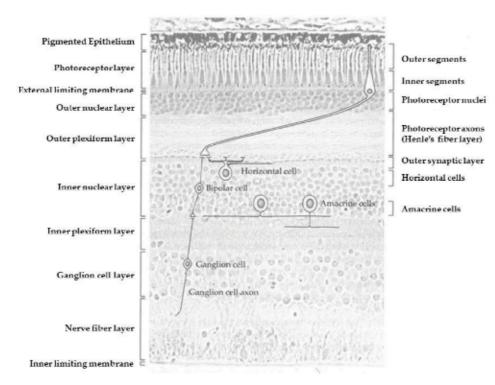
In 2015, it was estimated that AMD was the fourth leading cause of blindness and the third most common cause of moderate to severe visual impairment globally [4]. The meta-analysis performed by Wong and collaborators has estimated that the number of persons with AMD will increase from 196 million in 2020 to 288 million in 2040 worldwide [5]. A similar trend is observed in the projected number of individuals affected by AMD in China, rising from 31.23 million in 2020 to 55.19 million in 2050 [6]. In addition, the prevalence of late AMD did not show significant difference among Asian, European, and North American studies, whereas the number of individuals with early AMD was more in European and North American studies than in Asian studies [7].

### 2. Anatomy and function of the retina

The eye is composed of three layers, which are the inner retina layer, middle vascular choroid layer, and outer fibrous sclera layer, respectively. Retina, the innermost layer of the eye, consists of two parts: the inner transparent neurosensory retina and outer pigmented epithelial layer—the retinal pigment epithelium (RPE). There is a potential space between neural retina and RPE, called subretinal space. In the neural retina, the neural cell bodies are situated in three layers (**Figure 1**), including the outer nuclear layer (ONL) occupied with nuclei of photoreceptors; the inner nuclear layer (INL) filled with nuclei of horizontal, bipolar, and most of the amacrine cells; as well as ganglion cell layer (GCL) containing nuclei of retinal ganglion cells and the rest of displaced amacrine cells. Additionally, axons and dendrites of these retinal neurons constitute two synaptic layers: the inner plexiform layer (IPL) and outer plexiform layer (OPL) [8]. The RPE cells form a continuous polarized cell monolayer with its apical surface adjacent to the outer segment apices of the photoreceptors and its basal aspect lying on supportive substrate Bruch's membrane.

Retinal neurons are mainly distributed in three layers: ONL with nuclei of photoreceptors; INL with nuclei of horizontal, bipolar, and most of the amacrine cells; and GCL with nuclei of retinal ganglion cells and the rest of displaced amacrine cells. Additionally, axons and dendrites of these retinal neurons constitute two synaptic layers, including IPL and OPL [8].

Photoreceptors are divided into two types: rods, which are dominated in the peripheral retina and responsible for dim light vision and detecting movement and contrast, and cones, which are dominated in the macula, especially the fovea (only cones), and are responsible for bright light vision and sensing color vision



**Figure 1.** *Retinal layers of the human eye.* 

and resolution. Outer segments of the photoreceptor are generated by the cell body, and the distal parts of outer segments are phagocytosed by RPE every day. There are abundant mitochondria in inner segments to supply energy for these cells with high metabolic rate. Bruch's membrane, where the RPE cells are tightly attached, consists of five layers including, from inner to outer, the RPE basal lamina, inner collagenous zone, elastic layer, outer collagenous zone, and basement membrane of the choriocapillaris [9].

In brief, the function of the retina is to convert the external light signals into electrical impulses by photoreceptors. The impulses, which are partly integrated locally by horizontal cells and amacrine cells, are then processed by bipolar cells and sent to ganglion cells which further transmit them to the brain. Moreover, RPE is crucial for maintaining the microenvironment of neural retina by exchanging nutrients and wastes between neural retina and choroid, secreting numerous growth factors, phagocytosing shed photoreceptor outer segments (POS), absorbing light and converting all-trans-retinal into 11-cis-retinal [10]. Any defects in any of these functions may result in retinal degeneration, deficits of visual function, and eventually loss of sight.

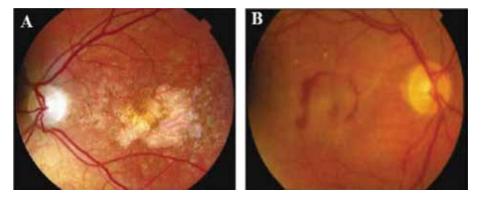
### 3. Current knowledge of AMD

### 3.1 Classification and clinical manifestations

A variety of classification systems are employed for both clinical and basic research of AMD. However, the most commonly used classification is the one defined by the Age-Related Eye Disease Study (AREDS) in terms of the characteristics of drusen (yellow deposits in the macula), abnormal hypo- or hyperpigmentation in the first eye, and how it affected the fellow eye. Based on the above criteria, AMD is classified into 4 categories: (1) no clinical manifestation of AMD if there was no drusen or only non-extensive, small drusen (<63  $\mu$ m in diameter) in both eyes; (2) mild AMD classified by extensive small drusen, non-extensive intermediate drusen (63–124  $\mu$ m in diameter), or abnormalities of pigments in at least one eye; (3) intermediate AMD characterized with large drusen (>124  $\mu$ m in diameter), extensive intermediate drusen, or noncentral geographic atrophy (GA) in at least one eye; (4) advanced AMD defined as central GA or choroidal neovascularization (CNV) resulting in VA less than 20/32 [11–13].

There are two clinical types of advanced AMD: a non-exudative or atrophic (dry) form, accounting for 90% of AMD, and an exudative (wet) form, accounting for only 10% of AMD. The atrophic form is characterized by progressive degeneration of RPE cells and photoreceptors in the macula, affecting central vision to varying degrees over months or years [14, 15]. The exudative form is associated with CNV in the submacular area and subsequent retinal hemorrhage due to leakage of these newly formed fragile blood vessels, leading to severe central vision loss within a very short period of time [15].

At the early stage of AMD, patients are normally asymptomatic and may be diagnosed by the presence of round, yellowish drusen through the routine ophthalmological examinations. Patients with CNV usually suffer from sudden loss of vision, describing unexpected deterioration of central visual field, distorted straight line (metamorphopsia), and/or a dark area in the central visual field (scotoma). On fundus examination, macular edema and hemorrhage are observed and fluorescein angiography shows leakage. In non-exudative AMD, it takes years for patients to develop visual loss gradually, and fundus examination shows a well-defined RPE atrophic area with depigmentation (**Figure 2**) [15].



#### Figure 2.

Representative fundus of AMD patients. (A) Non-exudative form of AMD with a well-defined atrophic region and (B) exudative form of AMD with retinal hemorrhage and macula edema [16].

#### 3.2 Risk factors

A number of risk factors are thought to be related to the pathogenesis of AMD, such as increasing age, smoking, family history of AMD, overweight or obesity, cataract and cataract surgery, eye exposure to the sunlight, etc. Of these factors, age is the strongest risk factor, with around 30% of cases older than 85 years diagnosed with AMD in US [17, 18]. The risk of developing AMD was reported to be more than triple in patients above 75 years than patients aged between 65 and 74 years [18, 19]. In populations of European ancestry, the projected prevalence of late AMD was 0.08% at age 50, 0.33% at age 60, 1.38% at age 70, 5.60% at age 80, and 20.10% at age 90, respectively [20]. Similarly, the estimated prevalence of any AMD increased from 2.44% in persons at the age of 45–49 years to 18.98% in persons at the age of 85–89 years [6].

Ethnicity and family history are also strongly related with AMD prevalence. The highest rate was reported in Caucasians, followed by Hispanics and Asians, while African Americans showed the lowest morbidity rate [21]. Increased risk of AMD was observed in individuals with positive family history, approximately three to six times higher than it was for those from the general population [22]. Furthermore, genetic studies have revealed 34 loci linked to AMD until now, of which complement factor H (CFH) on 1q31.3, age-related maculopathy susceptibility 2 (ARMS2) on 10q26, and CFB/C2 on 6q21.3 were the most discussed genes [15, 23, 24].

The strong relationship between cigarette smoking and development of AMD has been widely accepted [17, 25, 26]. Several studies revealed that current smoking was strongly associated with the progression of AMD [27]. Moreover, past smoking was reported to be a risk factor to develop AMD as well, especially in the rural areas [28]. There was an increased risk for individuals with high intake of several types of fat, such as omega-6 polyunsaturated fatty acids and saturated fats, whereas diets rich in monounsaturated fatty acids might reduce the prevalence of AMD [29]. Inadequate uptake of antioxidants also resulted in AMD, and numerous studies have suggested that antioxidant supplementation, including vitamins, lutein, zeaxan-thin, beta-carotene, zinc, etc., helped to halt the progression of AMD to some extent [12, 30, 31].

The effects of sunlight exposure on the occurrence of AMD are controversial. A study performed by Khan and colleagues demonstrated that no significant association between light exposure and AMD was observed when comparing patients at the end stage of AMD with spouse controls [32]. However, other studies suggested that damages induced by either ultraviolet (UV) or visible sunlight might lead to

AMD [33–35]. There was conflicting evidence regarding the role of cataract surgery in the progression of AMD [36]. Earlier, a large population-based research indicated a stronger relationship between cataract surgery with the development of end-stage AMD, especially neovascular AMD, in older patients [36, 37]. On the contrary, several recent clinical studies failed to demonstrate similar results [36, 38].

In addition to the risk factors mentioned above, other relevant factors included hypertension, cardiovascular diseases, iris color, hyperlipidemia, diabetes, alcohol consumption, and so on [22]. According to a systematic review of 18 prospective and cross-sectional studies and 6 case-control studies engaging 113,780 individuals, advancing age, cigarette smoking, previous cataract surgery, and family history of AMD were considered as the strong risk factors of AMD, while hypertension, cardiovascular disease, overweight or obesity, and elevated plasma fibrinogen were regarded as the moderate risk factors [17].

### 3.3 Pathophysiology

AMD is regarded as a complicated multifactorial disease, although the exact pathogenesis still remains poorly understood. It is generally recognized that impairment of aged RPE cell functions served an important role in the progression of non-exudative AMD. The aged RPE cells were less efficient in phagocytosing and degrading POS, resulting in the progressive accumulation of lipofuscin consisting of phagosomal and lysosomal constituents in the cytoplasm. The cytotoxic elements in lipofuscin, such as bisretinoid fluorophore, were able to generate reactive oxygen species leading to the damage of DNA, lipids, and proteins [39].

Another age-dependent change was the formation of focal extracellular yellow deposits known as drusen, the hallmark of AMD, between the RPE and Bruch's membrane. Accumulation of drusen relied on the changes in the permeability of Bruch's membrane due to the decline of RPE cell functions with aging [40]. In addition, it has been estimated that breakdown of choriocapillaris, which was next to RPE and Bruch's membrane, resulted in insufficient elimination of extracellular wastes causing drusen [41]. According to their shape and size, drusen are separated into small (<125  $\mu$ m), round hard drusen with well-defined borders, and relatively large, (125–250  $\mu$ m) soft drusen with poorly defined borders. Soft drusen, especially with depigmentation or pigment abnormalities, was considered to be an indication to give rise to severe vision loss at the late stage.

The mechanism of drusen causing adjacent RPE and photoreceptors' damage was not only dependent on the structural disturbance of RPE and photoreceptor by them, but also on the indirect effects through stimulation of local inflammation and immune system [42, 43]. Components in complement pathway and inflammatory processes were observed in drusen and aged RPE cells that are closely related to drusen [42]. It was demonstrated that in AMD, genetic variation of factor H gene (HF1/CFH) induced the abnormal activities of factor H (the inhibitor of complement cascade) in drusen, leading to the activation of complement pathway and subsequent inflammation in subretinal tissues [44]. Furthermore, in AMD patients, decreased CFH level was reported in smokers than nonsmokers, suggesting the activation of complement cascade might contribute to the significantly higher risk of AMD progression in cases with smoking [45].

DICER1, the ribonucleic acid (RNA)-cleaving enzyme, was decreased in RPE cells in cases with non-exudative AMD. A group of researchers in the University of Kentucky suggested that the lower level of DICER1 led to the lower rate of Alu RNA degradation in RPE cells. Therefore, accumulation of Alu RNA in cytoplasm resulted in RPE cytotoxicity and consequent degeneration via activating NLRP3 inflammasome [46, 47].

In exudative AMD, local nonspecific inflammation stimulated the upregulation of angiogenic factors, such as vascular endothelial growth factor (VEGF), and/or downregulation of anti-angiogenic factors, such as pigment epithelium-derived factor (PEDF), causing CNV development in the avascular outer 1/3 retina. The new vessels are fragile, thus macular edema and hemorrhage occur due to the leakage of these new blood vessels, which finally resulted in the fibrovascular scars in the macula [41, 48].

### 3.4 Affected neurons

Due to the dysfunction of aged RPE cells, especially the failure of POS phagocytosis, photoreceptor loss occurs subsequently. Although the most remarkable clinical and pathological injuries are present in RPE and its underlying Bruch's membrane, it is the structural and functional disruptions, even the death of photoreceptors, via either atrophic or neovascular process, that take responsibilities for the visual impairment in AMD. Furthermore, the condition of photoreceptors directly reflects the significance of lesions in RPE/Bruch's membrane complex.

It is crucial to identify which type of photoreceptors are most severely damaged in AMD, not only for the potential therapeutic strategies targeting the most affected cells, but also for investigation of mechanism of these pathological changes. The rate of rod and cone degeneration is representative in different situations affecting photoreceptors. In aging retina without age-related maculopathy, the number of cells in the cone-dominated fovea remained stable, while the number of rods in the parafovea was reduced by 30% [49]. In both non-exudative and exudative forms of AMD, photoreceptors were lost. In addition, more rod loss was observed than that of cones; gradually, only degenerated cones were left; finally, all photoreceptors might die [50]. The pathological changes mentioned above were consistent with the functional research exhibiting that scotopic sensitivity decreased more than photopic sensitivity in cases with AMD [51]. Maeda and colleagues revealed the apoptosis of photoreceptors after RPE damage caused by intravitreal injection of ornithine in rats, indicating the important role of RPE cells in maintaining photoreceptor integrity [52]. In AMD patients, apoptotic photoreceptors and RPE cells (TUNEL positive) were observed as well. Most of TUNEL-positive photoreceptors were rods and located at the edge of RPE atrophy. Moreover, Fas was upregulated in apoptotic photoreceptors, indicating Fas/FasL might be involved in the apoptosis process [53]. Studies performed by Kim and collaborators demonstrated that significant reduction of photoreceptors was shown in the areas where RPE cells were totally lost in GA, and where disciform scar formed in wet AMD [54, 55]. In terms of disciform scar, thickness of the scar was closely associated with photoreceptor loss, which means that the thicker the disciform scar, the less photoreceptors survived [56].

In addition to photoreceptors, other retinal neurons are also affected. Joshua et al. first reported that TUNEL-positive cells were detected in the inner side of INL, indicating that these cells might be amacrine cells [53]. In cases with GA, retinal ganglion cells were significantly decreased by 30.7% compared to agematched control. However, cell nuclei in INL were not significantly different [54]. In wet form of AMD, a decrease of ganglion cells and increase of cells in INL were observed, but there was no significant difference [55].

#### 3.5 Current treatments

Several treatments have been adopted for AMD management. However, current therapeutics can only slow the progression of the disease, trying to delay the onset of vision loss as much as possible.

To date, there are no approved drugs for the dry form of AMD. Therefore, much effort has been made to reduce risk factors. Among various risk factors, oxidative stress induced by inflammation, light exposure, and so on is considered as one of the most important risk factors for the occurrence and progression of AMD. Numerous studies have been conducted to evaluate the relationship between antioxidant nutrition supplementation and AMD. In the Age-Related Eye Disease Study (AREDS), it was demonstrated that oral dietary supplementation containing vitamin C (500 mg/day) and E (400 IU/day), zinc oxide (80 mg/day), cupric oxide (2 mg/day), and beta-carotene (15 mg/day) can decrease the risk of development of AMD from intermediate stage to advanced stage [11]. Since beta-carotene was observed to increase the risk of lung cancer in cigarette smokers, and 80 mg/day zinc is out of tolerance for individuals, there were elimination of beta-carotene, decreased dose of zinc (25 mg/day), and adding of lutein and zeaxanthin in the AREDS2 formula [12, 57, 58]. A 10% reduction in developing to advanced AMD was present in patients treated with AREDS2 formula containing lutein and zeaxanthin [59]. In addition, healthy diet rich in fish, green leafy vegetables, and nuts together with healthy lifestyle are strongly recommended to reduce AMD risk factors [15].

For the wet form of AMD, therapies mainly focus on halting the progression of CNV. Thermal laser photocoagulation is the first treatment to stop the progression of CNV successfully, but with no significant vision improvement and high recurrence of CNV. Photodynamic therapy with verteporfin can selectively damage the CNV tissue without additional injuries of neighboring tissue, but this therapy has no effects on visual improvement either. In terms of upregulation of VEGF in the development of CNV, intravitreal injection of anti-VEGF drugs has been widely used by ophthalmologists as a standard treatment. Anti-VEGF drugs (Pegaptanib sodium, Ranibizumab, and Bevacizumab) for exudative AMD have demonstrated exciting results: the vision in the majority of patients remained stable for 1 year, of which 40% of patients had visual improvement. Surgical intervention to remove CNV and submacular hemorrhage did not improve VA, which is the result of recurrence of CNV [15, 60].

### 4. Neuroprotection in AMD experimental studies and clinical approaches

Neuroprotection comprises a large number of therapeutic interventions to improve survival of neurons by modifying the structure and function of neurons, and/or their microenvironment. Initially, neuroprotective therapies are focused on central nervous system diseases including stroke, Alzheimer's disease, Parkinson's disease, etc. Since retina is regarded as the end part of central nervous system, a series of neuroprotective strategies has been applied to prevent vision loss by protecting retinal neurons. Furthermore, tremendous neuroprotective strategies are under investigation in both experimental and clinical research.

### 4.1 Studies in animal models

Neurotrophic factors, belonging to the family of growth factors, have the ability to promote survival of retinal neurons. Ciliary neurotrophic factor (CNTF) is one of the most extensively studied neurotrophic factors for neural retina protection. La Vail and collaborators first reported that intraocular injection of CNTF obviously prevented photoreceptor death from light-induced damage in Sprague Dawley rats [61]. Subsequently, intravitreal injection of adenoviral vector containing CNTF cDNA in rd1 mice, a naturally occurring mouse model for

retinal degenerative diseases, demonstrated the reduction of photoreceptor loss, conservation of ONL thickness, and increase of photoreceptor segments' length. Moreover, the amplitudes of a-wave and b-wave in electroretinogram (ERG) were significantly increased compared with those of the control group, suggesting the preservation of retinal functions [62, 63]. Later, long-term protective effects of photoreceptors were shown using adeno-associated virus to deliver CNTF to the retina [64]. In order to sustainably deliver neurotrophic factors, encapsulated human RPE cells secreting CNTF were transplanted into the vitreous of rcd1 dog (a dog model of retinal degeneration). A significant increase of ONL thickness was observed in the treated eye as a result of continuous release of CNTF at the nano-gram level [65]. No adverse effects were exhibited in the retina of transplanted eyes during the whole experiment period (7 weeks). In addition, CNTF was proved to protect loss of cone outer segments, an early sign of cone degeneration, indicating that CNTF could not only slow or halt progression of degeneration but also might reverse degeneration [66].

Placental growth factor (PIGF), one of the members of vascular endothelial growth factor family, was believed to prevent neuronal injury in the brain. In the retina, the role of PIGF was exhibited quite differently in *in vitro* and *in vivo* studies. Blue light-induced murine photoreceptor cell death was significantly attenuated after the treatment of PIGF by suppressing caspase-3/7 activity through the mitogen-activated protein kinase (MEK) and phosphoinositide 3-kinase (PI3K) pathway. Anti-PIGF antibody eliminated these protective effects [67]. However, in the light-induced retina-damaged mouse model, PIGF induced decreased ONL thickness and dysfunction of retina. Anti-PIGF antibody diminished neuroretinal injury and disruption of RPE cell-cell junctions after exposure to the white light for 3 h [68]. The opposite effects of PIGF and its antibody in mice were later found to be due to the hyperpermeability of RPE induced by PIGF, leading to the breakdown of retina-blood barrier and subsequent damages [68].

Ursodeoxycholic acid (UDCA) and its taurine-conjugated derivative tauroursodeoxycholic acid (TUDCA) were first found in the bile acid of hibernating bears. They have been used for liver detoxification, dissolution of gallstone and kidney stone, suppression of convulsions, and visual improvement in traditional Chinese medicine for a very long time. According to the theory of modern medicine, UDCA and TUDCA exhibit neuroprotective effects through prevention of cell apoptosis [69]. TUDCA treatment significantly preserved the number and structure of photoreceptors and retinal functions in different murine models of photoreceptor degeneration, including rd10 mice, rd1 mice, BALB/c mice, Bardet-Biedl syndrome type 1 mice, and transgenic P23H rats [69]. Furthermore, TUDCA manifested greater protective effects in cones [70]. In vitro studies using photoreceptor 661 W cells revealed that reduced endoplasmic reticulum (ER) stress and improved trafficking of cyclic nucleotide-gated channels in cones contributed to neuroprotective effects of TUDCA [71]. In addition, TUDCA improved phagocytosis of POS in H<sub>2</sub>O<sub>2</sub>-treated RPE cells via activating Mer tyrosine kinase receptor (MerTK), which indirectly protected photoreceptors [72].

Endogenous and exogenous progesterone have been certified to have neuroprotective effects in brain and retina for several decades. A broad range of studies have been conducted in either light-damaged or genetic murine models of retinal degeneration, demonstrating improvement in photoreceptor survival, decreased gliosis, and reduced retinal dysfunction after administration of progesterone or synthetic progestins [69]. A group of researchers in Spain revealed that rd1 mice orally administered with progesterone (100 mg/kg body weight) at postnatal day 7 (P7) exhibited significantly decreased number of apoptotic cells in ONL in the far peripheral retina and increased amplitude of ERG b-wave at P15, but no

significant change was observed at P17. There was also a transient reduced gliosis in the treated rd1 mice [73]. Similar results were observed with oral administration of synthetic progestin, known as the FDA-approved Norgestrel, showing reduction of photoreceptor death by 70 and 75% in light-damaged mouse model and rd10 mice, respectively [74]. The rescue effects were achieved by increasing production of basic fibroblast growth factor and its downstream pro-survival reactive oxygen species [74, 75].

Crystallins, critical family members of small heat shock proteins, have been identified to have novel functions in both retina and RPE as in the lens, such as antiapoptosis and anti-inflammation. *aB-Crystallin* is secreted in exosomes released from apical surface of polarized RPE cells and accumulates in the matrix among photoreceptors, and therefore may protect neighboring RPE and photoreceptors [76]. RPE cells in  $\alpha$ B-crystallin knockout mice exhibited high susceptibility to oxidative stress and endoplasmic reticulum stress compared to the RPE cells from wild-type mice. Furthermore, RPE cells overexpressing *aB*-crystallin were more resistant to apoptosis, indicating the protective effects of  $\alpha$ B-crystallin [77]. In the mouse model of AMD induced by sodium iodate (NaIO3), absence of  $\alpha$ B-crystallin accelerated RPE apoptosis with subsequent death of photoreceptors through upregulation of AKT phosphorylation and expression of peroxisome proliferatoractivator receptor- $\gamma$ , suggesting  $\alpha$ B-crystallin, especially the small peptide called mini cry, may play an important role in the protection of retinal degeneration [77]. In order to prolong the life of mini cry in the vitreous, free mini cry was fused to form an elastin-like polypeptide (ELP), which could be detected in the vitreous for up to 2 weeks. One intravitreal injection of ELP-linked peptide prevented RPE cells from apoptosis, inhibited activation of caspase-3 activation, and protected neural retina for up to 1 month after NaIO3 challenge [78].

NF-E2-related factor 2 (Nrf2) is a transcription factor that regulates antioxidant responses in many tissues and cell types, providing protection against oxidative stress. Under the oxidative stress, Nrf2 is translocated from cytoplasm to nucleus, and subsequently binds to the corresponding sites to activate transcription of a wide range of antioxidant genes. In the central nervous system, Nrf2 was proved to slow the neurodegeneration by means of antioxidative stress and neuroinflammation [79]. In the mice undergoing optic nerve crush (an animal model of glaucoma), retinal ganglion cells were significantly decreased than in the wild-type mice. With the treatment of Nrf2 activator, retinal ganglion cell loss was decreased by upregulating gene expression of antioxidant and phase II detoxifying enzymes [80]. After retinal ischemia-reperfusion injury, Nrf2 knockout mice showed greater loss of retinal ganglion cells when compared with the wild-type mice. Moreover, after ischemia-reperfusion injury, Nrf2 activator increased survival of retinal ganglion cells in wild-type mice, but not Nrf2 knockout mice, indicating the neuroprotective effects of Nrf2 [81]. An *in vitro* study has demonstrated that siRNA knockdown of Nrf2 led to significant increase of reactive oxygen species and cell death after blue light exposure in murine photoreceptor cells, suggesting that Nrf2 could be used to protect photoreceptors in AMD and other retinal degeneration [82].

There is no direct evidence that dopamine protects retinal neurons through antiinflammation, apoptosis, and oxidative stress. However, decreased expression of dopamine may play a negative role in neuron survival in the brain [83]. Thus, dopamine receptor agonists may achieve the goal to promote retinal neuron survival and function by restoring the lost dopamine resulting from certain diseases. Dopamine receptor agonist pramipexole was shown to ameliorate structural and functional injuries in the light-damaged mice, exhibiting decreased photoreceptor death, damage of photoreceptor outer and inner segments, TUNEL-positive cells in ONL, and preservation of a-wave and b-wave in ERG [84]. Additionally, pramipexole inhibited ARPE-19 cell (an immortalized RPE cell line) death after H<sub>2</sub>O<sub>2</sub> treatment, suggesting its antioxidative effects [84].

Regular general physical exercise is not only a part of healthy life style, but also a rehabilitation strategy showing neuroprotective effects in numerous diseases. Recently, exercise has been proven to be neuroprotective in animal models of retinal degeneration [85–87]. Wild-type BALB/c mice were forced to exercise for 5 days/ week for 2 weeks before being exposed to bright light. Exercised mice showed greater improved amplitude of ERG b-wave and photoreceptor nuclei than the mice without exercise after light exposure [85]. In order to exclude the impact of stress caused by forced exercise, voluntary wheel running was adopted in rd10 mice, which also demonstrated protection of VA, and the number of cones and total photoreceptors [86]. No matter if it was involuntary or voluntary exercise, either one benefited damaged photoreceptors in animal models of retinal degeneration through increased expression of brain-derived neurotrophic factor (BDNF) and activation of tropomyosin-related kinase B (TrkB) signaling pathway [85, 86].

Electrical neurostimulation has developed rapidly in recent years, covering a range of neurological diseases, such as neurostimulation for epilepsy, spinal cord stimulation for chronic pain, brain stimulation for Parkinson's disease, and so on [88]. Electrical neurostimulation in vision research has also made a great improvement, from transcorneal, subretinal, to whole-eye electrical stimulation. Transcorneal electrical stimulation (TES) was performed in SD rats before or after exposure to the intense light for 14 days. Both the stimulation before and after light exposure slowed the progression of photoreceptor degeneration. Furthermore, TES after light exposure exhibited a longer and better protective effect. The neuroprotection effects may result from anti-apoptosis (upregulation of Bcl-2 and downregulation of Bax) and increased expression of neurotrophic factors (CNTF and BDNF) [89]. Subretinal electrical stimulation (SES) in the eye of Royal College of Surgeons (RCS) rats, a commonly used model of retinal degeneration, significantly preserved amplitudes of b-wave and oscillatory potential, and implicit times of a-wave and b-wave, suggesting the preservation of not only photoreceptors, but also signal transmission in the retina [90]. Although the whole-eye electrical stimulation did not provide protection for rod structure in P23H rats, b-wave amplitudes and rod sensitivity were significantly increased [91].

Although the neuroprotective strategies mentioned above have acquired promising results in animal models, damaged human retina can be only partially rescued. Miller and colleagues have investigated the underlying reasons using many animal models. There are three types of cell death including caspase-mediated apoptosis, autophagy-mediated cell death, and necrosis which is regulated by receptor-interacting protein kinases (RIPK). In the animal model of retinal detachment, elevated expression and phosphorylation of RIPK together with activation of caspases were observed. When either RIPK or caspases were inhibited, no obvious rescue effects were demonstrated. However, inhibition of both RIPK and caspases resulted in significant protective effects. Similar results were found in the animal model of retinal degeneration which experienced both photoreceptor and RPE cell death. In this animal model, the dominant type of cell death in photoreceptors was apoptosis, while necrotic RPE cells were mainly exhibited. Therefore, therapies that block both RIPK and caspase pathways may provide more satisfactory neuroprotection [92].

### 4.2 Current medicine/neuroprotective agents in clinical trials

Neurotech Pharmaceuticals developed an intraocular drug delivery system using encapsulated cell technology, called NT-501, to consistently release CNTF in the vitreous for more than 2 years [93]. A double-masked, randomized,

sham-controlled, phase II study enrolling 51 GA patients who were randomly divided into 3 groups, high-dose NT-501 implant, low-dose NT-501 implant, and sham control group, demonstrated promising outcomes after a 2-year evaluation [94]. Another phase II study using intraocular implant secreting CNTF in GA patients has also been completed. In this multicenter, double-masked, randomized, sham-controlled, 1-year, dose-ranging phase II study, VA stabilization, defined as loss of less than 15 letters, was observed in the high-dose group (96.3%) versus low-dose group (83.3%) and sham surgery (75%). All the patients with best corrected VA  $\geq$ 20/63 in the high-dose group lost less than 15 letters, while it was only 55.6% in the combined group of patients treated with low-dose implant and sham surgery. Additionally, increase of retinal thickness was consistent with the stabilization of VA [95].

Brimonidine, an  $\alpha$ -2 agonist, is usually used to treat glaucoma patients in ophthalmology. Since brimonidine has been reported to protect neuroretinal cells in murine models, the effects of this drug on dry AMD patients were under evaluation [96]. A randomized, double-masked, sham-controlled, phase II study involved 119 patients with bilateral GA who were randomly divided into 3 groups including 200 µg treatment, 400 µg treatment, and sham control group. The efficacy and safety of brimonidine on biodegradable implant were evaluated after intravitreal transplantation [97]. But the results were not reliable, so another multicenter study is currently performed with larger samples (311 eyes receiving either treatment of 400 µg brimonidine on biodegradable implant or sham treatment) and longer evaluation period (up to 24 months) [98].

A retrospective analysis was performed to analyze the association between intake of L-DOPA and incidence of AMD. The results showed that the onset of AMD in individuals prescribed with L-DOPA was 8 years later when compared with those without the uptake of L-DOPA. The protective effects of L-DOPA might be through GPR143 (the only known L-DOPA receptor) pathway [99, 100].

There are few studies about exercise as a clinical intervention in AMD, and the published papers are observational research showing the correlation between exercise and prevalence of AMD. A study lasting for 15 years demonstrated that active lifestyle with physical exercise 3 times per week or more was associated with reduced risk to develop wet AMD [101]. In another cohort study, an inverse relation was observed between vigorous exercise ( $\geq$ 3 times/week) and occurrence of intermediate AMD in women, not in men [102]. In addition, low physical exercise was related to the formation of drusen larger than 63 µm [103].

Electrical neurostimulation has already been applied to patients using less invasive approaches, such as transcorneal or whole-eye electrical stimulation, to protect the structure and function of retinal neurons. In a clinical trial conducted by Anastassiou and colleagues, 22 patients with dry AMD received transpalpebral electrical stimulation twice a day for 5 days. Most of the patients demonstrated the improvement in VA and contrast sensitivity at 4 weeks after the treatment; however, only contrast sensitivity was significantly different when compared with sham control group [104]. Similarly, microcurrent stimulation (150  $\mu$ A for 35 min) once a week for 3 months was applied to both dry and wet AMD. Significantly increased VA was demonstrated in dry AMD, but not in wet AMD. Moreover, the number of patients showing increased VA was twice as those exhibiting deterioration [105].

### 5. Conclusions

AMD is believed to have stronger relationship with age. With increase of aging population worldwide, more individuals are suffering from visual damage,

resulting in poor quality of life for the aged people and elevated cost of medical care. In AMD, it is not only the degeneration of RPE cells, but also the neural retina degeneration, especially photoreceptors, that leads to the visual impairment. Therefore, neuroprotection can be one of the therapeutic strategies to slow, halt, or even reverse the progression of retina degeneration. Although the neuroprotective interventions that are currently investigated in both animal models and patients demonstrate promising results, it is of importance to identify the long-term efficacy and safety of these interventions. Only after that, the therapeutics will be provided to patients to help them maintain vision and further improve the quality of life.

### Acknowledgements

This work is supported by the Project of Dominant Discipline Construction in Universities of Shaanxi Province-Basic Medicine, Shaanxi Province, China, awarded to Xi'an Medical University as well as Health and Medical Research Fund, Hong Kong (04150746), awarded to Amy C.Y. Lo.

### **Conflict of interest**

The authors declare no conflict of interest.

### **Author details**

Shen Nian<sup>1</sup> and Amy C.Y. Lo<sup>2\*</sup>

1 Department of Pathology, Xi'an Medical University, Xi'an, Shaanxi Province, China

2 Department of Ophthalmology, LKS Faculty of Medicine, The University of Hong Kong, Hong Kong

\*Address all correspondence to: amylo@hku.hk

### IntechOpen

© 2019 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/ by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

### References

[1] Zimmer Z, Jagger C, Chiu CT, Ofstedal MB, Rojo F, Saito Y. Spirituality, religiosity, aging and health in global perspective: A review. SSM—Population Health. 2016;2:373-381. DOI: 10.1016/j. ssmph.2016.04.009

 [2] United Nations. World Population Prospects: The 2017 Revision. 2017.
 Available from: https://www.un.org/ development/desa/publications/ world-population-prospects-the-2017revision.html [Accessed: September 12, 2018]

[3] National Bureau of Statistics of China. National Population Projections. 2017. Available from: http://data.stats.gov.cn/easyquery. htm?cn=C01&zb=A0306&sj=2016 [Accessed: September 12, 2018]

[4] Bourne RRA, Flaxman SR, Braithwaite T, Cicinelli MV, Das A, Jonas JB, et al. Magnitude, temporal trends, and projections of the global prevalence of blindness and distance and near vision impairment: A systematic review and metaanalysis. The Lancet Global Health. 2017;5(9):e888-ee97. DOI: 10.1016/ S2214-109X(17)30293-0

[5] Wong WL, Su X, Li X, Cheung CM, Klein R, Cheng CY, et al. Global prevalence of age-related macular degeneration and disease burden projection for 2020 and 2040: A systematic review and metaanalysis. The Lancet Global Health. 2014;**2**(2):e106-e116. DOI: 10.1016/ S2214-109X(13)70145-1

[6] Song P, Du Y, Chan KY, Theodoratou E, Rudan I. The national and subnational prevalence and burden of age-related macular degeneration in China. Journal of Global Health. 2017;7(2):020703. DOI: 10.7189/ jogh.07.020703 [7] Jonas JB, Cheung CMG, Panda-Jonas S. Updates on the epidemiology of age-related macular degeneration. Asia-Pacific Journal of Ophthalmology (Phila). 2017;**6**(6):493-497. DOI: 10.22608/APO.2017251

[8] Oyster CW. Retina I: Photoreceptors and functional organization. In: Oyster CW, editor. The Human Eye: Structure and Function. Sunderland, Mass: Sinauer Associates; 1999. pp. 555-557

[9] Sheridan C, Williams R, Grierson I. Basement membranes and artificial substrates in cell transplantation. Graefe's Archive for Clinical and Experimental Ophthalmology. 2004;**242**(1):68-75. DOI: 10.1007/ s00417-003-0800-z

[10] Strauss O. The retinal pigment epithelium in visual function.Physiological Reviews. 2005;85(3):845-881. DOI: 10.1152/physrev.00021.2004

[11] Age-Related Eye Disease Study Research Group. Risk factors associated with age-related macular degeneration. A case-control study in the age-related eye disease study: Age-related eye disease study report number 3. Ophthalmology. 2000;**107**(12):2224-2232

[12] Age-Related Eye Disease Study Research Group. A randomized, placebo-controlled, clinical trial of highdose supplementation with vitamins C and E, beta carotene, and zinc for agerelated macular degeneration and vision loss: AREDS report no. 8. Archives of Ophthalmology. 2001;**119**(10): 1417-1436

[13] Bird AC, Bressler NM, Bressler SB, Chisholm IH, Coscas G, Davis MD, et al. An international classification and grading system for age-related maculopathy and age-related macular degeneration. The international ARM epidemiological study group. Survey of Ophthalmology. 1995;**39**(5):367-374

[14] Sunness JS, Rubin GS, Applegate CA, Bressler NM, Marsh MJ, Hawkins BS, et al. Visual function abnormalities and prognosis in eyes with age-related geographic atrophy of the macula and good visual acuity. Ophthalmology. 1997;**104**(10):1677-1691

[15] Lim LS, Mitchell P, Seddon JM, Holz FG, Wong TY. Age-related macular degeneration. Lancet.
2012;**379**(9827):1728-1738. DOI: 10.1016/S0140-6736(12)60282-7

[16] Arnold J, Sarks S. Age related macular degeneration. Clinical Evidence. 2003;**10**:727-743

[17] Chakravarthy U, Wong TY, Fletcher A, Piault E, Evans C, Zlateva G, et al. Clinical risk factors for age-related macular degeneration: A systematic review and meta-analysis. BMC Ophthalmology. 2010;**10**:31. DOI: 10.1186/1471-2415-10-31

[18] Klein R, Klein BE, LintonKL. Prevalence of age-relatedmaculopathy. The beaver dam eye study.Ophthalmology. 1992;99(6):933-943

[19] Leibowitz HM, Krueger DE, Maunder LR, Milton RC, Kini MM, Kahn HA, et al. The Framingham eye study monograph: An ophthalmological and epidemiological study of cataract, glaucoma, diabetic retinopathy, macular degeneration, and visual acuity in a general population of 2631 adults, 1973-1975. Survey of Ophthalmology. 1980;**24**(Suppl):335-610

[20] Rudnicka AR, Jarrar Z, Wormald R, Cook DG, Fletcher A, Owen CG. Age and gender variations in age-related macular degeneration prevalence in populations of European ancestry: A meta-analysis. Ophthalmology. 2012;**119**(3):571-580. DOI: 10.1016/j. ophtha.2011.09.027 [21] Chou R, Dana T, Bougatsos C, Grusing S, Blazina I. Screening for impaired visual acuity in older adults: Updated evidence report and systematic review for the US preventive services task force. Journal of the American Medical Association. 2016;**315**(9):915-933. DOI: 10.1001/ jama.2016.0783

[22] Al-Zamil WM, Yassin SA. Recent developments in age-related macular degeneration: A review. Clinical Interventions in Aging. 2017;**12**: 1313-1330. DOI: 10.2147/CIA.S143508

[23] Hughes AE, Orr N, Esfandiary H, Diaz-Torres M, Goodship T, Chakravarthy U. A common CFH haplotype, with deletion of CFHR1 and CFHR3, is associated with lower risk of age-related macular degeneration. Nature Genetics. 2006;**38**(10):1173-1177. DOI: 10.1038/ng1890

[24] Fritsche LG, Igl W, Bailey JN, Grassmann F, Sengupta S, Bragg-Gresham JL, et al. A large genome-wide association study of age-related macular degeneration highlights contributions of rare and common variants. Nature Genetics. 2016;**48**(2):134-143. DOI: 10.1038/ng.3448

[25] Chakravarthy U, Augood C, Bentham GC, de Jong PT, Rahu M, Seland J, et al. Cigarette smoking and age-related macular degeneration in the EUREYE study. Ophthalmology. 2007;**114**(6):1157-1163. DOI: 10.1016/j. ophtha.2006.09.022

[26] Khan JC, Thurlby DA, Shahid H, Clayton DG, Yates JR, Bradley M, et al. Smoking and age related macular degeneration: The number of pack years of cigarette smoking is a major determinant of risk for both geographic atrophy and choroidal neovascularisation. The British Journal of Ophthalmology. 2006;**90**(1):75-80. DOI: 10.1136/ bjo.2005.073643

[27] Singh N, Srinivasan S, Muralidharan V, Roy R, Jayprakash V, Raman R. Prevention of age-related macular degeneration. Asia-Pacific Journal of Ophthalmology (Phila). 2017;6(6): 520-526. DOI: 10.22608/APO.2017416

[28] Raman R, Pal SS, Ganesan S, Gella L, Vaitheeswaran K, Sharma T. The prevalence and risk factors for age-related macular degeneration in rural-urban India, Sankara Nethralaya rural-urban age-related macular degeneration study, report No. 1. Eye (London, England). 2016;**30**(5):688-697. DOI: 10.1038/eye.2016.14

[29] Parekh N, Voland RP, Moeller SM, Blodi BA, Ritenbaugh C, Chappell RJ, et al. Association between dietary fat intake and age-related macular degeneration in the carotenoids in agerelated eye disease study (CAREDS): An ancillary study of the Women's Health Initiative. Archives of Ophthalmology. 2009;**127**(11):1483-1493. DOI: 10.1001/ archophthalmol.2009.130

[30] Chew EY, Clemons TE, Agron E, Sperduto RD, Sangiovanni JP, Kurinij N, et al. Long-term effects of vitamins C and E, beta-carotene, and zinc on age-related macular degeneration: AREDS report no. 35. Ophthalmology. 2013;**120**(8):1604-11.e4. DOI: 10.1016/j. ophtha.2013.01.021

[31] Age-Related Eye Disease Study 2 Research Group. Lutein + zeaxanthin and omega-3 fatty acids for age-related macular degeneration: The agerelated eye disease study 2 (AREDS2) randomized clinical trial. JAMA. 2013;**309**(19):2005-2015. DOI: 10.1001/ jama.2013.4997

[32] Khan JC, Shahid H, Thurlby DA, Bradley M, Clayton DG, Moore AT, et al. Age related macular degeneration and sun exposure, iris colour, and skin sensitivity to sunlight. British Journal of Ophthalmology. 2006;**90**(1):29-32. DOI: 10.1136/bjo.2005.073825 [33] Glazer-Hockstein C, Dunaief JL. Could blue light-blocking lenses decrease the risk of age-related macular degeneration? Retina. 2006;**26**(1):1-4

[34] Margrain TH, Boulton M, Marshall J, Sliney DH. Do blue light filters confer protection against age-related macular degeneration? Progress in Retinal and Eye Research. 2004;**23**(5):523-531. DOI: 10.1016/j.preteyeres.2004.05.001

[35] Liu IY, White L, LaCroix AZ. The association of age-related macular degeneration and lens opacities in the aged. American Journal of Public Health. 1989;**79**(6):765-769

[36] Qian CX, Young LH. The impact of cataract surgery on AMD development and progression. Seminars in Ophthalmology. 2014;**29**(5-6):301-311. DOI: 10.3109/08820538.2014.962166

[37] Wang JJ, Klein R, Smith W, Klein BE, Tomany S, Mitchell P. Cataract surgery and the 5-year incidence of latestage age-related maculopathy: Pooled findings from the beaver dam and blue mountains eye studies. Ophthalmology. 2003;**110**(10):1960-1967

[38] Rim TH, Lee CS, Lee SC, Kim S, Kim SS. Association between previous cataract surgery and age-related macular degeneration. Seminars in Ophthalmology. 2017;**32**(4):466-473. Epub 2016/04/30

[39] Sparrow JR, Boulton M. RPE lipofuscin and its role in retinal pathobiology. Experimental Eye Research. 2005;**80**(5):595-606. DOI: 10.3109/08820538.2015.1119861

[40] Green WR, McDonnell PJ, YeoJH. Pathologic features of senile macular degeneration. Ophthalmology. 1985;92(5):615-627

[41] Biesemeier A, Taubitz T, Julien S, Yoeruek E, Schraermeyer U. Choriocapillaris breakdown precedes retinal degeneration in age-related macular degeneration. Neurobiology of Aging. 2014;**35**(11):2562-2573. DOI: 10.1016/j.neurobiolaging.2014.05.003

[42] Johnson LV, Leitner WP, Staples MK, Anderson DH. Complement activation and inflammatory processes in Drusen formation and age related macular degeneration. Experimental Eye Research. 2001;**73**(6):887-896. DOI: 10.1006/exer.2001.1094

[43] Anderson DH, Mullins RF, Hageman GS, Johnson LV. A role for local inflammation in the formation of drusen in the aging eye. American Journal of Ophthalmology. 2002;**134**(3):411-431

[44] Hageman GS, Anderson DH, Johnson LV, Hancox LS, Taiber AJ, Hardisty LI, et al. A common haplotype in the complement regulatory gene factor H (HF1/CFH) predisposes individuals to age-related macular degeneration. Proceedings of the National Academy of Sciences of the United States of America. 2005;**102**(20):7227-7232. DOI: 10.1073/ pnas.0501536102

[45] Sepp T, Khan JC, Thurlby DA, Shahid H, Clayton DG, Moore AT, et al. Complement factor H variant Y402H is a major risk determinant for geographic atrophy and choroidal neovascularization in smokers and nonsmokers. Investigative Ophthalmology & Visual Science. 2006;47(2):536-540. DOI: 10.1167/ iovs.05-1143

[46] Tarallo V, Hirano Y, Gelfand BD, Dridi S, Kerur N, Kim Y, et al. DICER1 loss and Alu RNA induce age-related macular degeneration via the NLRP3 inflammasome and MyD88. Cell. 2012;**149**(4):847-859. DOI: 10.1016/j. cell.2012.03.036

[47] Kaneko H, Dridi S, Tarallo V, Gelfand BD, Fowler BJ, Cho WG, et al. DICER1 deficit induces Alu RNA toxicity in age-related macular degeneration. Nature. 2011;**471**(7338):325-330. DOI: 10.1038/ nature09830

[48] Ng EW, Adamis AP. Targeting angiogenesis, the underlying disorder in neovascular age-related macular degeneration. Canadian Journal of Ophthalmology. 2005;**40**(3):352-368. DOI: 10.1016/S0008-4182(05)80078-X

[49] Curcio CA, Millican CL, Allen KA, Kalina RE. Aging of the human photoreceptor mosaic: Evidence for selective vulnerability of rods in central retina. Investigative Ophthalmology & Visual Science. 1993;**34**(12):3278-3296

[50] Curcio CA, Medeiros NE, Millican CL. Photoreceptor loss in age-related macular degeneration. Investigative Ophthalmology & Visual Science. 1996;**37**(7):1236-1249

[51] Curcio CA. Photoreceptor topography in ageing and age-related maculopathy. Eye (London, England). 2001;**15**(Pt 3):376-383. DOI: 10.1038/ eye.2001.140

[52] Maeda H, Ogata N, Yi X, Takeuchi M, Ohkuma H, Uyama M. Apoptosis of photoreceptor cells in ornithine-induced retinopathy. Graefe's Archive for Clinical and Experimental Ophthalmology. 1998;**236**(3): 207-212

[53] Dunaief JL, Dentchev T, Ying GS, Milam AH. The role of apoptosis in age-related macular degeneration. Archives of Ophthalmology. 2002;**120**(11):1435-1442

[54] Kim SY, Sadda S, Humayun MS, de Juan E Jr, Melia BM, Green WR. Morphometric analysis of the macula in eyes with geographic atrophy due to age-related macular degeneration. Retina. 2002;**22**(4):464-470

[55] Kim SY, Sadda S, Pearlman J, Humayun MS, de Juan E Jr, Melia BM, et al. Morphometric analysis of the macula in eyes with disciform agerelated macular degeneration. Retina. 2002;**22**(4):471-477

[56] Green WR, Enger C. Age-related macular degeneration histopathologic studies. The 1992 Lorenz
E. Zimmerman lecture. Ophthalmology. 1993;100(10):1519-1535

[57] Albanes D, Heinonen OP, Taylor PR, Virtamo J, Edwards BK, Rautalahti M, et al. Alpha-Tocopherol and betacarotene supplements and lung cancer incidence in the alpha-tocopherol, beta-carotene cancer prevention study: Effects of base-line characteristics and study compliance. Journal of the National Cancer Institute. 1996;**88**(21):1560-1570

[58] Johnson AR, Munoz A, Gottlieb JL, Jarrard DF. High dose zinc increases hospital admissions due to genitourinary complications. The Journal of Urology. 2007;**1**77(2):639-643. DOI: 10.1016/j. juro.2006.09.047

[59] Chew EY, Clemons TE, Sangiovanni JP, Danis RP, Ferris FL 3rd, Elman MJ, et al. Secondary analyses of the effects of lutein/ zeaxanthin on age-related macular degeneration progression: AREDS2 report No. 3. JAMA Ophthalmology. 2014;**132**(2):142-149. DOI: 10.1001/ jamaophthalmol.2013.7376

[60] Abd AJ, Kanwar RK, Kanwar JR. Aged macular degeneration: Current therapeutics for management and promising new drug candidates. Drug Discovery Today. 2017;**22**(11):1671-1679. DOI: 10.1016/j.drudis.2017.07.010

[61] LaVail MM, Unoki K, Yasumura D, Matthes MT, Yancopoulos GD, Steinberg RH. Multiple growth factors, cytokines, and neurotrophins rescue photoreceptors from the damaging effects of constant light. Proceedings of the National Academy of Sciences of the United States of America. 1992;**89**(23):11249-11253

[62] Cayouette M, Behn D, Sendtner M, Lachapelle P, Gravel C. Intraocular gene transfer of ciliary neurotrophic factor prevents death and increases responsiveness of rod photoreceptors in the retinal degeneration slow mouse. The Journal of Neuroscience. 1998;**18**(22):9282-9293

[63] Cayouette M, Gravel C. Adenovirusmediated gene transfer of ciliary neurotrophic factor can prevent photoreceptor degeneration in the retinal degeneration (rd) mouse. Human Gene Therapy. 1997;8(4): 423-430. DOI: 10.1089/hum

[64] Schlichtenbrede FC, MacNeil A, Bainbridge JW, Tschernutter M, Thrasher AJ, Smith AJ, et al. Intraocular gene delivery of ciliary neurotrophic factor results in significant loss of retinal function in normal mice and in the Prph2Rd2/Rd2 model of retinal degeneration. Gene Therapy. 2003;**10**(6):523-527. DOI: 10.1038/ sj.gt.3301929

[65] Tao W, Wen R, Goddard MB, Sherman SD, O'Rourke PJ, Stabila PF, et al. Encapsulated cell-based delivery of CNTF reduces photoreceptor degeneration in animal models of retinitis pigmentosa. Investigative Ophthalmology & Visual Science. 2002;**43**(10):3292-3298

[66] Li Y, Tao W, Luo L, Huang D, Kauper K, Stabila P, et al. CNTF induces regeneration of cone outer segments in a rat model of retinal degeneration. PLoS One. 2010;5(3):e9495. DOI: 10.1371/ journal.pone.0009495

[67] Inoue Y, Shimazawa M, Nakamura S, Imamura T, Sugitani S, Tsuruma K, et al. Protective effects of placental growth factor on retinal neuronal cell damage. Journal of Neuroscience Research. 2014;**92**(3):329-337. DOI: 10.1002/jnr.23316

[68] Izawa H, Inoue Y, Ohno Y, Ojino K, Tsuruma K, Shimazawa M, et al. Protective effects of Antiplacental growth factor antibody against lightinduced retinal damage in mice. Investigative Ophthalmology & Visual Science. 2015;**56**(11):6914-6924. DOI: 10.1167/iovs.15-16748

[69] Pardue MT, Allen RS. Neuroprotective strategies for retinal disease. Progress in Retinal and Eye Research. 2018;**65**:50-76. DOI: 10.1016/j.preteyeres.2018.02.002

[70] Oveson BC, Iwase T, Hackett SF, Lee SY, Usui S, Sedlak TW, et al. Constituents of bile, bilirubin and TUDCA, protect against oxidative stress-induced retinal degeneration. Journal of Neurochemistry.
2011;116(1):144-153. DOI: 10.1111/j.1471-4159.2010.07092.x

[71] Duricka DL, Brown RL, Varnum MD. Defective trafficking of cone photoreceptor CNG channels induces the unfolded protein response and ER-stressassociated cell death. The Biochemical Journal. 2012;441(2):685-696. DOI: 10.1042/BJ20111004

[72] Murase H, Tsuruma K, Shimazawa M, Hara H. TUDCA promotes phagocytosis by retinal pigment epithelium via MerTK activation. Investigative Ophthalmology & Visual Science. 2015;**56**(4):2511-2518. DOI: 10.1167/iovs.14-15962

[73] Sanchez-Vallejo V, Benlloch-Navarro S, Lopez-Pedrajas R, Romero FJ, Miranda M. Neuroprotective actions of progesterone in an in vivo model of retinitis pigmentosa. Pharmacological Research. 2015;**99**:276-288. DOI: 10.1016/j.phrs.2015.06.019

[74] Doonan F, O'Driscoll C, Kenna P, Cotter TG. Enhancing survival of photoreceptor cells in vivo using the synthetic progestin norgestrel. Journal of Neurochemistry. 2011;**118**(5):915-927. DOI: 10.1111/j.1471-4159.2011.07354.x

[75] Ruiz Lopez AM, Roche SL, Wyse Jackson AC, Moloney JN, Byrne AM, Cotter TG. Pro-survival redox signalling in progesterone-mediated retinal neuroprotection. The European Journal of Neuroscience. 2017;**46**(1):1663-1672. DOI: 10.1111/ejn.13604

[76] Kannan R, Sreekumar PG, Hinton DR. Novel roles for alpha-crystallins in retinal function and disease.
Progress in Retinal and Eye Research.
2012;**31**(6):576-604. DOI: 10.1016/j.
preteyeres.2012.06.001

[77] Zhou P, Kannan R, Spee C,
Sreekumar PG, Dou G, Hinton
DR. Protection of retina by alphaB
crystallin in sodium iodate induced
retinal degeneration. PLoS One.
2014;9(5):e98275. DOI: 10.1371/journal.
pone.0098275

[78] Sreekumar PG, Li Z, Wang W, Spee C, Hinton DR, Kannan R, et al. Intra-vitreal alphaB crystallin fused to elastin-like polypeptide provides neuroprotection in a mouse model of age-related macular degeneration. Journal of Controlled Release. 2018;**283**:94-104. DOI: 10.1016/j. jconrel.2018.05.014

[79] Buendia I, Michalska P, Navarro E, Gameiro I, Egea J, Leon R. Nrf2-ARE pathway: An emerging target against oxidative stress and neuroinflammation in neurodegenerative diseases. Pharmacology & Therapeutics. 2016;**157**:84-104. DOI: 10.1016/j. pharmthera.2015.11.003

[80] Himori N, Yamamoto K, Maruyama K, Ryu M, Taguchi K, Yamamoto M, et al. Critical role of Nrf2 in oxidative stress-induced retinal ganglion cell death. Journal of Neurochemistry. 2013;**127**(5):669-680. DOI: 10.1111/ jnc.12325

[81] Xu Z, Cho H, Hartsock MJ, Mitchell KL, Gong J, Wu L, et al. Neuroprotective role of Nrf2 for retinal ganglion cells in ischemia-reperfusion. Journal of Neurochemistry. 2015;**133**(2):233-241. DOI: 10.1111/jnc.13064

[82] Chen WJ, Wu C, Xu Z, Kuse Y, Hara H, Duh EJ. Nrf2 protects photoreceptor cells from photo-oxidative stress induced by blue light. Experimental Eye Research. 2017;**154**:151-158. DOI: 10.1016/j.exer.2016.12.001

[83] Lotharius J, Brundin P. Pathogenesis of Parkinson's disease: Dopamine, vesicles and alpha-synuclein. Nature Reviews Neuroscience. 2002;3(12):932-942. DOI: 10.1038/nrn983

[84] Shibagaki K, Okamoto K, Katsuta O, Nakamura M. Beneficial protective effect of pramipexole on light-induced retinal damage in mice. Experimental Eye Research. 2015;**139**:64-72. DOI: 10.1016/j.exer.2015.07.007

[85] Lawson EC, Han MK, Sellers
JT, Chrenek MA, Hanif A, Gogniat
MA, et al. Aerobic exercise protects
retinal function and structure from
light-induced retinal degeneration.
The Journal of Neuroscience.
2014;34(7):2406-2412. DOI: 10.1523/
JNEUROSCI.2062-13.2014

[86] Hanif AM, Lawson EC, Prunty M, Gogniat M, Aung MH, Chakraborty R, et al. Neuroprotective effects of voluntary exercise in an inherited retinal degeneration mouse model. Investigative Ophthalmology & Visual Science. 2015;**56**(11):6839-6846. DOI: 10.1167/iovs.15-16792

[87] Chrenek MA, Sellers JT, Lawson EC, Cunha PP, Johnson JL, Girardot PE, et al. Exercise and cyclic light preconditioning protect against lightinduced retinal degeneration and evoke similar gene expression patterns. Advances in Experimental Medicine and Biology. 2016;**854**:443-448. DOI: 10.1007/978-3-319-17121-0\_59 [88] Edwards CA, Kouzani A, Lee KH, Ross EK. Neurostimulation devices for the treatment of neurologic disorders. Mayo Clinic Proceedings. 2017;**92**(9):1427-1444. DOI: 10.1016/j. mayocp.2017.05.005

[89] Ni YQ, Gan DK, Xu HD, Xu GZ, Da CD. Neuroprotective effect of transcorneal electrical stimulation on light-induced photoreceptor degeneration. Experimental Neurology. 2009;**219**(2):439-452. DOI: 10.1016/j. expneurol.2009.06.016

[90] Ciavatta VT, Mocko JA, Kim MK, Pardue MT. Subretinal electrical stimulation preserves inner retinal function in RCS rat retina. Molecular Vision. 2013;**19**:995-1005

[91] Rahmani S, Bogdanowicz L, Thomas J, Hetling JR. Chronic delivery of low-level exogenous current preserves retinal function in pigmented P23H rat. Vision Research. 2013;**76**:105-113. DOI: 10.1016/jvisres.2012.10.016

[92] Miller JW, Bagheri S, Vavvas DG. Advances in age-related macular degeneration understanding and therapy. US Ophthalmic Review. 2017;**10**(2):119-130. DOI: 10.17925/ USOR.2017.10.02.119

[93] U.S. National Library of Medicine, ClinicalTrials.gov. A Study of an Encapsulated Cell Technology (ECT) Implant for Patients with Atrophic Macular Degeneration. 2009. Available from: https://clinicaltrials.gov/ct2/ show/NCT00447954 [Accessed: September 23, 2018]

[94] Birch DG, Weleber RG, Duncan JL, Jaffe GJ, Tao W. Randomized trial of ciliary neurotrophic factor delivered by encapsulated cell intraocular implants for retinitis pigmentosa. American Journal of Ophthalmology. 2013;**156**(2):283-92. e1. DOI: 10.1016/j.ajo.2013.03.021

[95] Zhang K, Hopkins JJ, Heier JS, Birch DG, Halperin LS, Albini TA, et al. Ciliary neurotrophic factor delivered by encapsulated cell intraocular implants for treatment of geographic atrophy in age-related macular degeneration. Proceedings of the National Academy of Sciences of the United States of America. 2011;**108**(15):6241-6245. DOI: 10.1073/pnas.1018987108

[96] U.S. National Library of Medicine, ClinicalTrials.gov. Safety and Efficacy of Brimonidine Intravitreal Implant in Patients with Geographic Atrophy Due to Age-related Macular Degeneration (AMD). 2013. Available from: https://clinicaltrials.gov/ct2/show/ NCT00658619 [Accessed: October 17, 2018]

[97] U.S. National Library of Medicine, ClinicalTrials.gov. A Safety and Efficacy Study of Brimonidine Intravitreal Implant in Geographic Atrophy Secondary to Age-related Macular Degeneration (BEACON). 2014. Available from: https://clinicaltrials. gov/ct2/show/NCT02087085 [Accessed: October 17, 2018]

[98] Lynn SA, Keeling E, Munday R, Gabha G, Griffiths H, Lotery AJ, et al. The complexities underlying age-related macular degeneration: Could amyloid beta play an important role? Neural Regeneration Research. 2017;**12**(4): 538-548. DOI: 10.4103/1673-5374. 205083

[99] McKay BS, Schwartz SG. Pigmentation and macular degeneration: Is there a role for GPR143? Journal of Ocular Pharmacology and Therapeutics. 2016;**32**(1):3-4. DOI: 10.1089/jop.2016.29007.bsm

[100] Brilliant MH, Vaziri K, Connor TB Jr, Schwartz SG, Carroll JJ, McCarty CA, et al. Mining retrospective data for virtual prospective drug repurposing: L-DOPA and age-related macular degeneration. The American Journal of Medicine. 2016;**129**(3):292-298. DOI: 10.1016/j.amjmed.2015.10.015 [101] Knudtson MD, Klein R, Klein BE. Physical activity and the 15-year cumulative incidence of age-related macular degeneration: The beaver dam eye study. The British Journal of Ophthalmology. 2006;**90**(12):1461-1463. DOI: 10.1136/bjo.2006.103796

[102] McGuinness MB, Karahalios A, Simpson JA, Guymer RH, Robman LD, Hodge AM, et al. Past physical activity and age-related macular degeneration: The Melbourne collaborative cohort study. The British Journal of Ophthalmology. 2016;**100**(10):1353-1358. DOI: 10.1136/ bjophthalmol-2015-307663

[103] Munch IC, Linneberg A, Larsen M. Precursors of age-related macular degeneration: Associations with physical activity, obesity, and serum lipids in the inter99 eye study. Investigative Ophthalmology & Visual Science. 2013;54(6):3932-3940. DOI: 10.1167/ iovs.12-10785

[104] Anastassiou G, Schneegans AL, Selbach M, Kremmer S. Transpalpebral electrotherapy for dry age-related macular degeneration (AMD): An exploratory trial. Restorative Neurology and Neuroscience. 2013;**31**(5):571-578. DOI: 10.3233/RNN-130322

[105] Chaikin L, Kashiwa K, Bennet M, Papastergiou G, Gregory W. Microcurrent stimulation in the treatment of dry and wet macular degeneration. Clinical Ophthalmology.
2015;9:2345-2353. DOI: 10.2147/OPTH.
S92296



### Edited by Raymond Chuen-Chung Chang and Yuen-Shan Ho

Neuroprotection is a strategy to prevent or delay the progression of chronic neurodegenerative diseases, acute neurological disorders, or even mental disorders. The major aim of this book is to focus on different approaches to achieve neuroprotection. In this book, there are chapters discussing imidazoline ligands and opioid ligands in Alzheimer's disease, the beneficial effects of adenosine A2A receptor antagonist, adrenergic receptor agonists and antagonists modulating microglial responses, and different approaches to achieve neuroprotection against agingassociated macular degeneration. This book will give insights to scientists in the field to stimulate their research, medical professionals to review their clinical practices, and others who would like to learn more about different neuroprotective approaches.

Published in London, UK © 2019 IntechOpen © ktsimage / iStock

IntechOpen



